

GUIDELINE OPEN ACCESS

European Society for the Study of Coeliac Disease (ESsCD) 2025 Updated Guidelines on the Diagnosis and Management of Coeliac Disease in Adults. Part 2: Management, Follow-Up, and Complex Disease Courses

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ABSTRACT

Introduction: Since the publication of the first European Society for the Study of Coeliac Disease (ESsCD) guidelines in 2019, substantial advances have been made in understanding the management and complex disease courses of coeliac disease (CeD)

Abbreviations: AGA, Antigliadin Antibodies; AGREE-II, Appraisal of Guidelines for Research and Evaluation II; auto-HSCT, Autologous Hematopoietic Stem Cell Transplantation; BMD, Bone Mineral Density; CeD, Coeliac Disease; CoE, certainty of evidence; DAE, Device-Assisted Enteroscopy; DGP, Deamidated Gluten Peptides; DH, Dermatitis Herpetiformis; DXA, Dual Energy Absorptiometry Measurement; EATL, Enteropathy-associated T cell Lymphoma; ESPGHAN, European Society Paediatric Gastroenterology, Hepatology and Nutrition; ESsCD, European Society for the Study of Coeliac Disease; FODMAPs, fermentable oligo di-mono-saccharides and polyols; GFD, Gluten-Free Diet; GR, grade of recommendation; GRADE, Grading of Recommendations Assessment, Development, and Evaluation; HLA, Human Leucocyte Antigen; IBD, Inflammatory Bowel Disease; IBS, Irritable Bowel Syndrome; ICI, immune checkpoint inhibitor; IEL, Intraepithelial Lymphocytes; IgA anti-EMA, anti-Endomysial Antibodies; LE, Level of Evidence; NA, Not Applicable; NCWS, Non-Coeliac Wheat Allergy; NPV, Negative Predictive Value; POCT, Point-of-care testing; PPV, Positive Predictive Value; QoL, Quality of Life; RCD-I, Refractory Coeliac Disease type 1; RCD-II, Refractory Coeliac Disease type 2; SNCD, Seronegative Coeliac Disease; T1DM, Type 1 Diabetes Mellitus; TG2, Tissue Transglutaminase 2; UGPS, ungraded good practice statement; ULN, Upper Limit of Normal; VCE, Video Capsule Endoscopy.

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in adults. These 2025 updated guidelines aim to integrate new evidence, refine management strategies, and promote a personalised and multidisciplinary approach to care.

Methods: The ESSCD convened a multidisciplinary panel of experts to revise the 2019 guidelines using the Appraisal of Guidelines for Research and Evaluation II (AGREE II) framework. Evidence was appraised and graded according to the Grading of Recommendations Assessment, Development, and Evaluation (GRADE) methodology. Statements and recommendations were draughted within working groups and finalised through a structured Delphi consensus process.

Results: The updated guidelines are presented in two parts. *Part 1*, which has already been published, addresses the diagnostic approach to CeD in adults, whereas *Part 2* focuses on disease management, structured follow-up, and the evaluation and treatment of persistent symptoms despite a gluten-free diet or refractory disease. New or expanded sections include guidance on the safe inclusion of oats, use of low-FODMAP diets in patients with persistent symptoms, management of exocrine pancreatic insufficiency, recognition of functional asplenia and related vaccination recommendations, and stratified bone-health screening. The guidelines also discuss nutritional and psychosocial support, digital models of care, and structured transition from paediatric to adult services. Updated therapeutic strategies for refractory CeD are provided, including immunosuppressive and novel pharmacologic options.

Conclusions: These updated guidelines offer a comprehensive, evidence-based framework for the management and follow-up of adults with CeD. By integrating recent scientific advances with pragmatic, patient-centred recommendations, they seek to optimise clinical outcomes, quality of life, and long-term health in individuals with CeD.

1 | Introduction

The 2025 European Society for the Study of Coeliac Disease (ESSCD) guidelines on coeliac disease (CeD) are presented in two complementary parts. Part 1 focuses on the diagnostic approach [1], whilst part 2- the present document-addresses disease management, follow-up, and the approach to patients with persistent or recurrent symptoms despite gluten-free diet (GFD) and refractory CeD (RCD). Together, these updates provide an integrated framework encompassing the full spectrum of adult CeD care.

These guidelines present an update to the 2019 ESSCD guidelines on the Management of CeD and Other Gluten-Related Disorders [2]. Since their publication, emerging evidence has prompted substantial refinements in dietary management, structured follow-up, and the care of patients with complex or treatment-refractory disease. Key developments include:

1. **Dietary Management:** Maintenance of a lifelong strict GFD remains central to treatment, with new emphasis on nutritional adequacy, metabolic health, prevention of deficiencies and aspects of food labelling. The safety of gluten-free oats is clarified, and a low-FODMAP dietary approach is proposed for patients with persistent gastrointestinal symptoms despite histological remission.
2. **Follow-Up & Monitoring:** The updated guidance recognises two possible follow-up strategies—a *tailored model* adapted to patient-specific needs and risk profiles, and a *standardised model* with fixed assessment intervals. Both approaches prioritise multidisciplinary evaluation, including telemedicine integration and systematic assessment of adherence, psychosocial wellbeing, and quality of life (QoL).
3. **Patient Support Groups** are recognized as an integral part of long-term management.
4. **Transition from Paediatric to Adult Care:** A “coeliac passport” is recommended to facilitate smooth transition

and ensuring continuity of care, summarising key medical and dietary information.

5. **Management of RCD:** Updated recommendations define diagnostic criteria and therapeutic strategies, including the potential use of targeted immunosuppressive therapies such as JAK inhibitors, cladribine or fludarabine, and autologous haematopoietic stem-cell transplantation for selected patients.
6. **Newly addressed areas:** Management of confirmed exocrine pancreatic insufficiency in CeD, stratified bone-health monitoring, psychosocial support, pregnancy management, risk-based family screening and the potential use of gluten immunogenic peptides (GIPs) as an adjunct tool for monitoring adherence to the GFD.

Collectively, these guideline revisions reflect a paradigm shift towards personalised, proactive, and multidisciplinary care for adults with CeD. The overarching goals are to improve disease control, enhance QoL, and reduce the risk of long-term complications through evidence-based, patient-centred management.

2 | Summary of Recommendations

2.1 | Methodology

We followed the methodology for assessing the quality of evidence and risk of bias as described in Part 1 of these ESSCD guidelines [1].

Whilst maintaining the same approach to grading the overall certainty of evidence according to the Grading of Recommendations, Assessment, Development, and Evaluation (GRADE) methodology, <https://www.gradeworkinggroup.org/>, we applied the following tools depending on study design: the Cochrane Risk of Bias tool (RoB 2) for randomized controlled trials (RCTs), ROBINS-I for non-randomized studies of interventions,

QUADAS-2 for diagnostic accuracy studies, AMSTAR-2 for systematic reviews, and the NIH Quality Assessment Tool for observational cohort and cross-sectional studies.

3 | Questions, Recommendations and Evidence: Management of Adults With Coeliac Disease

The management of CeD aims to relieve symptoms, promote mucosal healing, restore nutritional status, prevent complications, and optimize quality of life. The cornerstone of treatment is a strict, lifelong GFD, supported by expert dietetic input and ongoing adherence monitoring, whilst emerging dietary approaches, such as low-FODMAP strategies, may be considered in selected cases. Comprehensive care also includes screening for comorbidities, supporting psychosocial well-being, monitoring bone mineral density, providing vaccinations when indicated, and addressing associated conditions, including autoimmune disorders, metabolic syndrome, and extra-intestinal manifestations. Tailored strategies are required for special populations, evolving clinical scenarios such as CeD-like enteropathy during treatment with immune checkpoint inhibitors, and patients with co-existing conditions.

An overview of the key recommendations and statements addressing adult CeD management, follow-up strategies, and complex disease courses is summarised in Table 1. The clinical questions underpinning these recommendations, together with the supporting evidence, are detailed in Supplementary File 1 (PICOs and evidence tables), whilst the corresponding Evidence-to-Decision frameworks are presented in Supplementary File 2. Collectively, these resources provide a framework for holistic, multidisciplinary care that addresses both the biological and psychosocial impact of CeD.

3.1 | Dietary Management: The Gluten-Free Diet

3.1.1 | The Fundamentals of Dietary Management

3.1.1.1 | Q. What Is the Treatment for CeD?. Recommendation: The treatment for CeD is a lifelong adherence to a GFD. Strict adherence is essential for controlling symptoms, improving QoL, and reducing the risk of long-term complications.

Certainty of evidence (CoE):moderate; GR: strong; Agreement: 100%

3.1.1.2 | Q. Is There an Established Threshold for Safe Gluten Intake in Patients With CeD?. Recommendation: The generally accepted daily threshold of gluten intake for individuals with CeD is no more than 10 mg of gluten per day, as higher intakes over time may induce mucosal injury in some individuals.

CoE: moderate; GR: strong; Agreement: 100%

3.1.1.3 | Q. Are Oats Safe for Patients With CeD?. Recommendation: Only certified gluten-free oats are safe and should be recommended for people with CeD and can be

included from diagnosis as part of a well-balanced GFD. A small subset may develop intolerance related to avenin-specific CD4+ T cells.

CoE: moderate; GR: strong; Agreement: 100%

Summary of evidence: The only effective treatment for CeD is lifelong, strict adherence to a GFD, requiring complete exclusion of wheat, rye, and barley. Consistent evidence from large observational cohorts and systematic reviews involving more than 30,000 individuals shows that a GFD leads to rapid and sustained symptom improvement, serological normalization, and mucosal healing in most patients [3–5].

Symptomatic relief typically occurs within months, although 10%–20% of adults may experience persistent symptoms despite apparent adherence [6, 7]. Normalization of IgA anti-TG2 antibodies is common correlates fairly well with disease control, but complete histological healing is observed less frequently in adults than in children [4, 5]. Strict adherence improves nutritional status, reduces deficiency-related complications, and is associated with better QoL, although many patients experience social, practical, and psychological burdens in daily life and healthcare settings [6, 8–10].

Professional dietary counselling significantly enhances adherence and patient well-being, and minimizing inadvertent gluten exposure—particularly through cross-contamination in domestic, social, travel, and hospital environments—remains an essential component of long-term management [11]. In hospital settings, unintended gluten exposure remains common; safe management requires clear communication of dietary needs, access to gluten-free meals, and involvement of hospital dietetic services to ensure appropriate food provision and avoidance of contamination [12].

Because eliminating gluten entirely is difficult, *international thresholds* have been established to define ‘safe’ gluten exposure and to regulate gluten-free labelling. The relationship between gluten intake, symptoms, mucosal injury, and long-term outcomes in CeD remains incompletely defined. A well-designed 3-month micro-challenge study showed that 50 mg of gluten per day caused mucosal damage in several participants, whereas 10 mg per day was not consistently associated with histological deterioration over the study period [13]. Based on these and other available data, the Codex Alimentarius commission defined ‘gluten-free’ foods as containing no more than 20 mg gluten/kg (20 ppm), a threshold derived using conservative assumptions and clinical evidence indicating that most individuals with CeD tolerate daily gluten intakes below 10 mg without histological damage. Assuming a daily consumption of approximately 500 g of solid food, this corresponds to a maximum gluten exposure of up to ~10 mg/day. This threshold has been adopted into European law (Commission Implementing Regulation (EU) No 828/2014) [14]. Importantly, the 20 ppm gluten-free threshold is currently not under reconsideration [15].

A persistent challenge is the widespread use of ‘may contain gluten’ warnings when unintentional cross-contamination during production cannot be excluded, complicating risk

TABLE 1 | Overview of recommendations and statements regarding management of CeD in adults, follow-up, and complex disease courses.

Statement/Recommendation		Certainty of evidence	Grade of recommendation	Agreement %
3. Management of adults with CeD				
3.1. Dietary management: thg gluten-free diet (GFD)				
3.1.1. The fundamentals of dietary management				
3.1.1.1. Q. What is the treatment for CeD?	The treatment for CeD is a lifelong adherence to a GFD. Strict adherence is essential for controlling symptoms, improving quality of life, and reducing the risk of long-term complications.	Moderate	Strong	100
3.1.1.2. Q. Is there an established threshold for safe gluten intake in patients with CeD?	The generally accepted daily threshold of gluten intake for individuals with CeD is no more than 10 mg of gluten per day, as higher intakes over time may induce mucosal injury in some individuals.	Moderate	Strong	100
3.1.1.3. Q. Are oats safe for patients with CeD?	Only certified gluten-free oats are safe and should be recommended for people with CeD and can be included from diagnosis as part of a well-balanced GFD. A small subset may develop intolerance related to avenin-specific CD4+ T cells.	Moderate	Strong	100
3.1.2. Role of the dietitian at diagnosis and follow-up				
3.1.2.1. Q. What is the role of the dietitian at diagnosis?	At diagnosis, the dietitian should provide education on the lifelong GFD, perform a baseline nutritional assessment, address deficiencies, and offer personalised dietary guidance. Early coordination with the healthcare team and planned follow-up are essential to support treatment initiation.	Low	Strong	100
3.1.2.2. Q. What is the role of the dietitian in the follow-up of CeD?	1. During follow-up, a CeD-specialised dietitian should monitor nutritional status, reassess deficiencies, and support ongoing adherence to a strict and balanced GFD. 2. Dietetic review is essential for detecting inadvertent gluten exposure and helping evaluate persistent or recurrent symptoms.	Low	Strong	100
3.1.3. Nutritional assessment and management				
3.1.3.1. Q. How to assess the nutritional status at diagnosis of CeD in adults?	Assessment of nutritional status at diagnosis of CeD in adults should combine clinical evaluation, anthropometric measurements, dietary assessment, and targeted laboratory testing to identify malnutrition and micronutrient deficiencies.	Low	Strong	100
3.1.3.2. Q. What are the recommendations for a balanced GFD in order to improve the nutritional status, overall health, and well-being?	For patients with CeD, a balanced GFD should be based on naturally gluten-free foods whilst recognising the risk of gluten contamination, particularly in grain products. Vigilant attention to food labelling is important. The diet should include dairy or fortified alternatives, ensure adequate macronutrient intake, and address common nutritional deficiencies, including supplementation when required.	Low	Strong	100

(Continues)

TABLE 1 | (Continued)

	Statement/Recommendation	Certainty of evidence	Grade of recommendation	Agreement %
3.1.3.3. Q. Can deficiencies of micro- or macro-nutrient arise as a result of the GFD?	When a GFD is poorly balanced, it may lead to macro- and micronutrient deficiencies. Thusly, nutritional monitoring and dietetic support are recommended.	<i>Low</i>	<i>Strong</i>	100
3.1.4. Assessment of gluten-free Diet adherence				
3.1.4.1. Q. How can adherence to a GFD be assessed in adults with CeD?	- Adherence to a GFD in adults with CeD can be assessed using a combination of clinical evaluation, serology, and structured dietary review by a specialized dietitian. - For an objective measure of recent gluten intake—particularly in cases of persistent symptoms or uncertain adherence—detection of GIPs in stool or urine can be considered. - Duodenal biopsy remains the definitive method for assessing mucosal healing but is not routinely required for adherence monitoring alone.	<i>Low</i>	<i>Strong</i>	100
3.1.4.2. Q. What are the factors associated with lower rates of adherence to GFD?	Adherence to a GFD varies widely and is reduced by factors such as younger age, lower socioeconomic status, eating outside the home, absence of symptoms, and limited knowledge.	<i>Low</i>	<i>Strong</i>	95
3.1.5. Other dietary interventions				
3.1.5.1. Q. What is the role of temporary lactose restriction in adults with CeD?	Temporary lactose restriction can play a role in adults with CeD who experience persistent gastrointestinal symptoms at diagnosis due to secondary lactase deficiency. Such intolerance is usually transient and resolves as the intestinal mucosa heals on a GFD.	<i>Low</i>	<i>Conditional</i>	100
3.1.5.2. Q. For adult patients with CeD with persistent GI symptoms, is there a role for a low-FODMAP diet?	In adults with CeD who have persistent GI symptoms despite a strict GFD and confirmed histological healing, low-FODMAP diet may be considered, provided that other potential causes of symptoms are excluded before starting this additional dietary restriction.	<i>Moderate</i>	<i>Conditional</i>	100
3.2. Multidisciplinary support				
3.2.1. Q. Does psychosocial support improve well-being, adherence, and QoL?	Psychological assessment and support may be beneficial for a subset of adults with CeD, particularly those experiencing distress or coping difficulties, as it can contribute to improved well-being, dietary adherence, and QoL.	<i>Low</i>	<i>Conditional</i>	95
3.2.2. Q. How can the transition of care from paediatric to adult services be conducted efficiently?	The guidelines panel suggests a formal transfer of medical care of adolescent with CeD. To facilitate the transfer, a transition letter from paediatricians or a coeliac passport is required, detailing criteria for CeD diagnosis, follow-up, serology, growth data, possible comorbidities, and dietary adherence.	<i>Low</i>	<i>Strong</i>	100

(Continues)

TABLE 1 | (Continued)

	Statement/Recommendation	Certainty of evidence	Grade of recommendation	Agreement %
3.2.3. Q. What is the role of patient support groups in the management of adults with CeD?	Patient support groups play a valuable role in the management of adults with CeD by providing education, practical guidance, emotional support, and advocacy, all of which can improve dietary adherence and QoL.	<i>Not applicable (NA)</i>	<i>Ungraded good practise statement</i>	100
3.3. Management of associated conditions & special scenarios				
3.3.1.1. Q. How should thyroid function be assessed in adults with CeD?	Thyroid function should be assessed at the time of CeD diagnosis using a TSH test, with automatic reflex free T4 testing if TSH is abnormal. Consider repeating thyroid function tests periodically, especially in patients with symptoms or risk factors for thyroid disease.	<i>Low</i>	<i>Strong</i>	100
3.3.1.2. Q. Are there additional dietary recommendations for patients with CeD patients with concomitant type1 diabetes mellitus?	Patients with CeD and concomitant T1DM should follow a dietary plan that integrates gluten-free requirements with the carbohydrate-counting strategies essential for T1DM management. Emphasis on gluten-free carbohydrate sources with a lower glycaemic index may help support more stable glycaemic control.	<i>Low</i>	<i>Strong</i>	100
3.3.2. Bone mineral density in adults with CeD				
3.3.2.1. Q. Should adults with CeD undergo assessment of bone mineral density, and which individuals require a DXA scan?	Adults with CeD are at increased risk of reduced bone mineral density (BMD) and related fractures. Assessing BMD enables timely interventions to support bone health and prevent osteoporosis, including correction of nutritional deficiencies, lifestyle measures, and optimisation of the GFD. A baseline DXA scan is recommended after 1 year on a GFD for adults with CeD who have additional risk factors for low BMD, including delayed diagnosis, severe malabsorption or marked weight loss at presentation, a history of fragility fractures, other independent osteoporosis risk factors, or down syndrome. For adults without these additional risk factors, routine DXA scanning is not mandatory. However, it may be considered in those approaching midlife (> 35–40 years), where low bone mass is more common. FRAX can support individualised fracture-risk assessment and decision-making.	<i>Low</i>	<i>Strong</i>	89
3.3.3. Immune dysfunction and vaccination				
3.3.3.1. Q. Which patients with CeD require pneumococcal vaccination?	Pneumococcal vaccination is recommended for adults with CeD with evidence of functional asplenia, concurrent autoimmune disorders or RCD-II, as well as for patients over 65 years.	<i>NA</i>	<i>Ungraded good practise statement</i>	100
3.3.4. Extra-intestinal manifestations				
See Section 3.3.4	We recommend that significant extra-intestinal manifestations of CeD be managed by a multidisciplinary team. This should include a	<i>NA</i>	<i>Ungraded good practise statement</i>	100

(Continues)

TABLE 1 | (Continued)

	Statement/Recommendation	Certainty of evidence	Grade of recommendation	Agreement %
	gastroenterologist and dietitian to oversee the GFD and intestinal health, alongside a specialist with expertise in the specific manifestation (i.e., neurologist, dermatologist, dentist) for targeted diagnosis and management.			
3.3.5. Concurrent GI and metabolic issues				
3.3.5.1. Q. Would CeD patients benefit from pancreatic enzyme supplementation?	In adult patients with CeD, pancreatic enzyme replacement therapy (PERT) may be indicated when exocrine pancreatic insufficiency is confirmed. PERT can improve digestive function, relieve symptoms such as steatorrhoea and bloating, enhance nutrient absorption, and improve QoL.	Moderate	Conditional	89
3.3.5.2. Q. Do probiotics improve symptoms and QoL when added to a GFD?	Current evidence is insufficient to support or oppose the use of probiotics alongside a GFD for improving symptoms and QoL in patients with CeD. Further research may determine their effectiveness.	NA	Ungraded good practise statement	95
3.3.5.3. Q. Are adults with CeD on a GFD at higher risk of developing metabolic syndrome?	Adults with CeD on a GFD have a higher risk of developing metabolic syndrome compared to individuals with CeD before dietary therapy. A balanced GFD, active lifestyle, and regular monitoring for metabolic risk factors—including obesity, hypertension, dyslipidaemia, and insulin resistance—are recommended.	Low	Strong	89
3.3.6. Special scenarios				
3.3.6.1. Q. Do pregnant women with CeD require specialized care?	Although evidence is limited, coordinated preconception and perinatal care may help optimize nutritional status in women with CeD and support a healthy environment for early foetal development. Postpartum care should continue to emphasize maintaining the GFD.	NA	Ungraded good practise statement	95
3.3.6.2. Q. What are the key management considerations for patients diagnosed with CeD at an older age?	In patients diagnosed with CeD at an older age, non-classical symptoms are common, often causing diagnostic delays. These patients have an increased risk of complications, including osteoporosis and malignancy. Although response to a GFD may be slower, it provides substantial clinical benefit and should be strongly considered.	NA	Ungraded good practise statement	100
3.3.6.3. Q. How to screen family members of patients with CeD?	1. HLA-DQ2/8 genotyping is recommended as the initial screening step primarily in children of patients with CeD —where it can prevent repeated testing—while anti-TG2 serology remains the most cost-effective and widely available initial test for adults and lower-risk relatives. 2. Follow-up: For first-degree relatives who are seronegative at initial assessment, periodic antibody follow-up (e.g., every 4–5 years) may	Moderate	Conditional	95

(Continues)

TABLE 1 | (Continued)

	Statement/Recommendation	Certainty of evidence	Grade of recommendation	Agreement %
	be considered based on individual risk factors and new symptoms.			
3.3.6.4. Q. How to manage adults presenting with the rare severe presentation ‘coeliac crisis’?	Although rare in adults, coeliac crisis—often a manifestation of refractory CeD type 2—requires urgent admission for multidisciplinary care to address dehydration, electrolyte/nutritional deficits, and precipitating factors, whilst promptly initiating a strict GFD.	NA	Ungraded good practise statement	95
3.3.6.5. Q. What is the recommended management for patients who develop CeD-like enteropathy during treatment with immune checkpoint inhibitors?	In patients receiving immune checkpoint inhibitors (ICI) who develop diarrhoea, coeliac serology should be obtained. If serology is positive, histological confirmation of CeD should be sought. Conversely, if a CeD-like enteropathy is identified histologically in a patient on ICI, coeliac serology should be performed to confirm the diagnosis. First-line management is a strict GFD when CeD is confirmed. Systemic immunosuppression or modification of immunotherapy should be reserved for severe presentations or cases not responding to a GFD, with decisions guided by multidisciplinary discussion.	Low	Conditional	95
3.4. Pharmacological therapies				
3.4.1. Q. What is the current status of non-dietary drug treatments for CeD?	Non-dietary pharmacological treatments for CeD hold promise for the future but remain experimental and are currently available only within clinical trials.	Moderate	Strong	100
4. Long-term follow-up and monitoring				
4.1. Q. Why is long-term follow-up recommended for adults with CeD?	Long-term follow-up is recommended to monitor dietary adherence, detect complications and comorbidities, and provide ongoing nutritional and psychosocial support, which helps ensure sustained disease control and optimised long-term outcomes.	NA	Ungraded good practise statement	95
4.2. Q. What is the most appropriate healthcare setting for the long-term follow-up of adults with CeD, and how should responsibilities be shared between primary and secondary care?	Long-term follow-up of adults with CeD is best coordinated through hospital-based outpatient clinics or specialized coeliac centres, where feasible. For stable patients, follow-up may be conducted in primary care, provided there is clear access to specialist support. Digital tools can be utilised to support remote monitoring and dietary adherence.	NA	Ungraded good practise statement	95
4.3. Q. Should follow-up of adults with CeD be conducted at fixed-intervals or tailored to the individual through a patient-centred approach?	Both fixed-interval and individualised strategies for follow-up of adults with CeD have advantages; however, follow-up should be individualised and patient-centred, taking into account disease activity, treatment adherence, comorbidities, and individual needs.	NA	Ungraded good practise statement	100
	The role of IgA anti-TG2 in follow-up is to identify ongoing gluten exposure; a positive	Moderate	Strong	95

(Continues)

TABLE 1 | (Continued)

	Statement/Recommendation	Certainty of evidence	Grade of recommendation	Agreement %
4.4. Q. What is the role of coeliac serology in follow-up of adult patients with CeD?	IgA anti-TG2 result in patients with CeD on a GFD suggests potential poor dietary adherence or gluten contamination, whilst a negative result does not confirm strict adherence or the absence of gluten exposure and it is not a reliable marker of villous atrophy.			
4.5. Q. Is a follow-up duodenal biopsy necessary in adults with CeD?	A follow-up duodenal biopsy is not routinely necessary for all adults with CeD. It is recommended in a personalized manner, guided by factors such as age at diagnosis, symptom severity, serological status, and clinical response to the GFD. A biopsy should be considered if symptoms persist or worsen.	Low	Conditional	95
5. Complicated CeD				
5.1. Kinetics of clinical, serologic and histologic response to a GFD in adults with CeD				
5.1.1. Q. When is a clinical response to the diet to be expected in adults with CeD?	Clinical response to a GFD in adults with CeD shows substantial inter-individual variability, with symptom improvement generally expected within 4 weeks to 4–5 months after diet initiation.	Low	Strong	100
5.1.2. Q. When is a serological response to the diet to be expected in adults with CeD?	A decline in IgA anti-TG2 can be observed as early as 2–4 weeks after starting a GFD, and in the majority, titres normalize within approximately 12 months. There is, however, strong inter-individual variation in this response.	Moderate	Strong	100
5.1.3. Q. When is a histological response to the GFD to be expected in adult CeD patients?	Healing of the duodenal mucosa is generally expected around 1 year after starting a GFD. However, the proportion of patients achieving full histological recovery ranges widely, from approximately 50%–83% after 1–5 years.	Low	Strong	100
5.2. Delayed or incomplete response to a GFD				
5.2.1. Q. What are causes of delayed or incomplete response to a gluten-free diet in adult CeD patients?	An incomplete response to a GFD is often due to ongoing gluten exposure but may also indicate slow-responsive disease, RCD, initial misdiagnosis, or concurrent conditions. Persistent symptoms or villous atrophy after $\geq$ 12 months requires systematic evaluation.	Moderate	Strong	100
5.2.2. Q. How to evaluate an adult patient with CeD having persistent symptoms despite GFD?	For a symptomatic adult with CeD despite a GFD, first verify the original diagnosis and assess for gluten exposure. If adherence is confirmed, proceed with follow-up histology and evaluate for alternative or overlapping conditions—such as functional disorders, other GI diseases, refractory CeD, or malignancy—using a multidisciplinary approach.	Moderate	Strong	100
5.3. Refractory coeliac disease				
5.3.1. Q. What is the definition of RCD?	RCD is defined by the persistence or recurrence of symptoms and villous atrophy after at least 12 months on a strict GFD, in the	Moderate	Strong	100

(Continues)

TABLE 1 | (Continued)

	Statement/Recommendation	Certainty of evidence	Grade of recommendation	Agreement %
	absence of other causes. RCD can be either primary or secondary. Depending on the proportion of aberrant T cells, RCD is further subdivided into: RCD-I and RCD-II.			
5.3.2. Diagnosis of RCD				
5.3.2.1. Q. What is the diagnostic approach to a patient with suspected RCD?	The diagnostic approach involves a systematic workup to confirm the initial CeD diagnosis, ensure strict dietary adherence, and exclude alternative causes of symptoms or an overt EATL. Essential investigations include serology, duodenal biopsies (for histology, IEL flow cytometry, and T-cell receptor clonality analysis), enteroscopy, and cross-sectional imaging. If a strong clinical suspicion for RCD persists, further evaluation and management should be conducted in or in consultation with a tertiary centre experienced in managing RCD.	Moderate	Strong	100
5.3.2.2. Q. Which molecular diagnostic technology is the gold standard for diagnosis of RCD-II?	Accurate subtyping of RCD requires comprehensive immunophenotyping of small bowel lymphocytes to reliably diagnose or exclude RCD-II. Flow cytometric analysis of isolated IELs after immunostaining is superior to immunohistochemistry for detecting aberrant lymphocyte populations. Additional diagnostic value is provided by PCR-based T-cell receptor clonality assessment and by sequencing of genes in the JAK/STAT pathway to identify relevant somatic mutations.	Moderate	Strong	100
5.3.3. Treatment of RCD				
5.3.3.1. Q. Is there an evidence-based treatment for RCD-I and RCD-II?	No evidence-based medical treatments supported by controlled trials exist for either RCD-I or RCD-II; current management is based on expert consensus, case series, and observational data.	Low	Conditional	100
5.3.3.2. Q. What is the treatment for patients with RCD-I?	Based on retrospective and longitudinal evidence, open-capsule budesonide is considered first-line therapy for RCD-I. Conventional immunosuppressant, such as azathioprine, may be added in selected cases. When used, azathioprine should be re-evaluated for possible discontinuation after 2–3 years of clinical, histological and immunophenotypic stability.	Low	Conditional	100
5.3.3.3. Q. What is the treatment for patients with RCD-II?	Mild-to-moderate RCD-II may be treated with open-capsule budesonide. In selected patients, cladribine (or fludarabine), with or without autologous haematopoietic stem cell transplantation (auto-HSCT), or JAK inhibitors may be considered on an individual basis.	Low	Conditional	100
6. Coeliac disease and malignancy: Associations and risk of malignant transformation		Low	Strong	95

(Continues)

TABLE 1 | (Continued)

	Statement/Recommendation	Certainty of evidence	Grade of recommendation	Agreement %
6.1. Q. What is the risk of malignant complications in coeliac disease (non-refractory CeD)?	The risk of malignant complications in non-refractory CeD is only slightly increased, remains low in absolute terms, and decreases with long-term adherence to a strict GFD. Routine malignancy screening beyond general population recommendations is not indicated in the absence of clinical suspicion.			
6.2. Q. What is the risk of malignant transformation in RCD-II?	RCD-II is associated with a substantial risk of malignant transformation, most commonly progression to EATL, which develops in up to half of affected patients and is the major determinant of the markedly reduced 5-year survival seen in this subtype.	Low	Strong	100

Abbreviations: auto-HSCT, Autologous Hematopoietic Stem Cell Transplantation; BMD, Bone Mineral Density; CeD, Coeliac Disease; CoE, Certainty of Evidence; DXA, Dual Energy Absorptiometry Measurement; EATL, Enteropathy-associated T cell Lymphoma; FODMAPs, Fermentable oligo di-mono-saccharides and polyols; FRAX, Fracture Risk Assessment Tool; GFD, Gluten-Free Diet; GI, Gastrointestinal; HLA, Human Leucocyte Antigen; ICI, immune checkpoint inhibitors; IEL, Intraepithelial Lymphocytes; IgA anti-EMA, anti-Endomysial antibodies; JAK/STAT, Janus kinase/signal transducer and activator of transcription pathway; NA, Not Applicable; PCR, polymerase chain reaction; PERT, pancreatic enzyme replacement therapy; QoL, Quality of Life; RCD-I, Refractory coeliac disease type 1; RCD-II, Refractory coeliac disease type 2; T1DM, Type 1 Diabetes Mellitus; T4, Thyroxine; TG2, Tissue Transglutaminase 2; TSH, Thyroid stimulating hormone.

assessment for patients. Given this uncertainty, most centres pragmatically advise patients to avoid these products to eliminate the possibility of sporadic high-level gluten exposure.

Pharmaceutical excipients must also comply with gluten-labelling rules to prevent inadvertent exposure [16–19].

Emerging evidence indicates that sensitivity to gluten varies amongst individuals and that even very low-level exposure may provoke subtle immune activation without accompanying histological abnormalities [20], challenging the reliance on mucosal healing as the sole marker of dietary safety. Moreover, it is unclear whether histological remission fully reflects long-term risk, as complications such as lymphomagenesis may stem from persistent cytokine activity and mucosal immune dysregulation that are not captured by standard histological assessment [21–23]. Further research incorporating immunologic, molecular, and clinical outcomes is needed to better define what constitutes ‘safe’ gluten exposure.

However, until newer evidence is validated and incorporated into regulatory frameworks, the ≤ 10 mg/day threshold and the 20 ppm labelling standard remain the most evidence-based and operationally feasible definitions of ‘gluten-free’.

The safety of oats in the GFD has been a longstanding topic of debate. Standard oats are commonly contaminated with gluten and are unsafe for people with CeD [24–26]. Certified gluten-free oats, however, are nutritionally valuable—providing fibre, vitamins, minerals, and antioxidants—and have been shown to improve dietary variety, GFD acceptability, and long-term gastrointestinal, psychological, and dermatological outcomes [24–26].

Historical concerns about avenin sensitivity [27, 28]. have largely been mitigated by recent studies demonstrating that

purified avenin can induce only transient, dose-dependent immune activation in a small minority of adults, without causing enteropathy or serological deterioration [24, 25]. True avenin sensitivity appears to be rare.

Current evidence therefore supports the inclusion of certified gluten-free oats from diagnosis, supported by clear labelling and patient education, although timing may be individualised based on patient preference, symptom burden, or local practise. In patients with symptoms suggestive of avenin intolerance, medical review and dietetic support are recommended; a short, structured trial of oat exclusion may be considered, with subsequent reassessment, to avoid unnecessary long-term restriction if symptoms do not clearly improve [29].

3.1.2 | Role of the Dietitian at Diagnosis and Follow-Up

3.1.2.1 | Q. What Is the Role of the Dietitian at Diagnosis of CeD?. *Recommendation:* At diagnosis, the dietitian should provide education on the lifelong GFD, perform a baseline nutritional assessment, address deficiencies, and offer personalised dietary guidance. Early coordination with the healthcare team and planned follow-up are essential to support treatment initiation.

CoE: low; GR: strong; Agreement: 100%

3.1.2.2 | Q. What Is the Role of the Dietitian in the Follow-Up of CeD?. *Recommendation:*

1. During follow-up, a CeD-specialised dietitian should monitor nutritional status, reassess deficiencies, and support ongoing adherence to a strict and balanced GFD.

2. Dietetic review is essential for detecting inadvertent gluten exposure and helping evaluate persistent or recurrent symptoms.

CoE: low; GR: strong; Agreement: 100%

Summary of evidence: Given the complexity and lifelong nature of the GFD, referral to an experienced dietitian is essential at diagnosis and throughout follow-up. Dietitians provide personalised counselling that considers the patient's social, cultural, and economic context [30, 31].

At diagnosis: The initial consultation should reaffirm the CeD diagnosis, review comorbidities, and emphasise the need for a strict lifelong GFD. Core components of early education include: clarifying gluten-containing grains and the distinction between contaminated and certified gluten-free oats; [7, 32–35] practical training on label reading and identification of naturally gluten-free foods, suitable substitutes, and processed foods not requiring a gluten-free label; [36] strategies for safe eating outside the home and during travel; [11, 37–39] introduction to patient support organisations; [40] and a baseline nutritional assessment.

During follow-up: Dietitians support ongoing management by monitoring nutritional adequacy, reassessing deficiencies, and providing tailored guidance to maintain a strict but balanced GFD. Education should reinforce healthy dietary patterns, weight maintenance, and avoidance of disordered eating or excessive hypervigilance [41–43].

Structured dietetic assessment is central to evaluating adherence and identifying causes of persistent or recurrent symptoms. Specialist dietitians detect inadvertent gluten exposure or dietary imbalance more reliably than tools such as the Coeliac Dietary Adherence Test (CDAT), showing higher sensitivity and specificity (64% and 80%) for detecting ongoing villous atrophy [44–47].

Despite clear benefits, access to coeliac-specialist dietetic services remains inconsistent, with shortages in dedicated clinics, staffing, and consultation time [48, 49]. Improved investment, defined service standards, and better integration of dietetic care into routine management are needed to ensure equitable long-term support for adults with CeD [50, 51].

3.1.3 | Nutritional Assessment and Management

3.1.3.1 | Q. How to Assess the Nutritional Status at Diagnosis of CeD in Adults?. *Recommendation:* Assessment of nutritional status at diagnosis of CeD in adults should combine clinical evaluation, anthropometric measurements, dietary assessment, and targeted laboratory testing to identify malnutrition and micronutrient deficiencies.

CoE: low; GR: strong; Agreement: 100%

3.1.3.2 | Q. What Are the Recommendations for a Balanced GFD in Order to Improve the Nutritional Status, Overall Health, and Well-Being?. *Recommendation:* For patients with CeD, a balanced GFD should be based on

naturally gluten-free foods whilst recognising the risk of gluten contamination, particularly in grain products. Vigilant attention to food labelling is important. The diet should include dairy or fortified alternatives, ensure adequate macronutrient intake, and address common nutritional deficiencies, including supplementation when required.

CoE: low; GR: strong; Agreement: 100%

3.1.3.3 | Q. Can Deficiencies of Micro- or Macro-Nutrient Arise as a Result of the GFD?. *Recommendation:* When a GFD is poorly balanced, it may lead to macro- and micronutrient deficiencies. Thusly, nutritional monitoring and dietetic support are recommended.

CoE: low; GR: strong; Agreement: 100%

Summary of evidence: Nutritional deficiencies are common in adults with CeD at diagnosis, even in those with normal weight or mild symptoms, due to malabsorption from mucosal damage and inadequate intake [52–54]. Iron deficiency affects 7%–80% of newly diagnosed patients, and 2%–5% of individuals with iron-deficiency anaemia are found to have CeD [53, 55]. Fat malabsorption can reduce levels of fat-soluble vitamins—especially vitamin D, which may be further lowered when dairy is restricted for lactose intolerance—and may also lead to deficiencies of vitamins A, E, and K [56, 57]. Additional abnormalities include low albumin, haemoglobin, and trace elements such as zinc and copper; the latter is notable for potentially causing haematological abnormalities and irreversible neurological deficits [54].

Assessment of nutritional status at diagnosis and during follow-up is essential to monitor correction of deficiencies and ensure dietary adequacy [58]. Key components include:

1. History and physical assessment for signs and symptoms suggestive of deficiencies, and screening for other autoimmune disorders.
2. Weight history including BMI, and where appropriate, waist-to-hip ratio and hand-grip strength [59].
3. Laboratory evaluation: haemoglobin, iron studies, fat-soluble vitamins, B12, folate, minerals, electrolytes, and liver function tests [57].
4. Dietary assessment: food and beverage intake, evaluation of nutrients commonly deficient in CeD (iron, calcium, vitamin D, B12, and fibre) [60].

A balanced GFD is central to correcting deficiencies and promoting overall health. Recommended strategies include: [61–64].

- Naturally gluten-free foods: Gluten-free grains (e.g., quinoa, brown rice, millet, amaranth, buckwheat, gluten-free oats), fruits and vegetables, lean proteins, dairy or fortified alternatives, and healthy fats form the foundation of a balanced gluten-free diet. Whilst naturally gluten-free foods are encouraged, it should be recognised that there is a heightened risk of gluten contamination in some food categories.

Evidence indicates that contamination occurs predominantly in oats, other grains, flours, and processed products, whereas fresh fruits and vegetables, unprocessed meats, and unprocessed dairy products are rarely implicated [65, 66]. Accordingly, appropriate sourcing, careful attention to food labelling, and preferential use of certified gluten-free products—particularly for staple grain-based foods—are important to minimise inadvertent gluten exposure.

- Gluten-free substitutes: Breads, pastas, and flours facilitate culturally relevant meals and social participation. Patients should preferentially select fortified products, whilst monitoring for higher saturated fat, sugar, and energy content [67]. Processed foods may be subject to gluten contamination during manufacturing and should therefore be selected with appropriate caution.
- Supplementation tailored to individual needs.
- Education and support: Ongoing support from a specialist dietitian and engagement with patient support groups [44, 68].

Despite clinical improvement on a GFD, nutritional inadequacies may arise from the diet itself [69–78]. Many gluten-free substitutes are nutritionally suboptimal, lower in protein, fibre, iron, vitamin B12, folate, and vitamin D, and higher in saturated fat, sodium, and added sugars [77, 79–82]. A systematic review and meta-analysis showed increased risk of multiple micronutrient deficiencies in treated CeD [78]. Heavy reliance on these products can perpetuate deficiencies, contribute to constipation reduce cardiometabolic benefits, and increase energy intake.

Body-weight changes after initiating a GFD vary with baseline status, and adequate diet quality does not appear to increase long-term obesity risk [83–85].

Overall, a well-balanced GFD—emphasising naturally gluten-free foods, judicious use of fortified substitutes, supplementation when needed, and structured dietetic support—is essential to prevent nutritional deficiencies and support long-term health and wellbeing in adults with CeD [77, 86].

3.1.4 | Assessment of GFD Adherence

3.1.4.1 | Q. How can Adherence to a GFD Be Assessed in Adults With CeD?. *Recommendations:*

- Adherence to a GFD in adults with CeD can be assessed using a combination of clinical evaluation, serology, and structured dietary review by a specialized dietitian.
- For an objective measure of recent gluten intake—particularly in cases of persistent symptoms or uncertain adherence—detection of gluten immunogenic peptides (GIPs) in stool or urine can be considered.
- Duodenal biopsy remains the definitive method for assessing mucosal healing but is not routinely required for adherence monitoring alone.

CoE: low; GR: strong; Agreement: 100%

3.1.4.2 | Q. What Are the Factors Associated With Lower Rates of Adherence to GFD?. *Statement:* Adherence to a GFD varies widely and is reduced by factors such as younger age, lower socioeconomic status, eating outside the home, absence of symptoms, and limited knowledge.

CoE: low; GR: strong; Agreement: 95%

Summary of evidence: Assessment of adherence to a GFD in adults with CeD should employ a combination of methods, including clinical evaluation, serology, structured dietary review by a specialist dietitian, and, when appropriate, direct detection of gluten ingestion. Table 2 provides an overview of methods for assessing adherence to the GFD.

Factors Affecting Adherence to GFD:

Adherence to a GFD varies widely, with reported rates ranging from 42% to 91%, depending on assessment method [100]. Lower adherence is associated with younger age at diagnosis, particularly adolescence, lower socioeconomic status, local food cultures, frequent eating outside the home, absence of symptoms, and limited disease knowledge or motivation [101, 102].

Gluten-free foods remain more expensive than standard equivalents, creating a financial barrier, particularly in lower-income households, which is linked to poorer adherence [103].

Targeted support—such as prioritizing dietetic resources for at-risk individuals, providing additional consultations, improving health literacy, and addressing perceived dietary burden—can enhance adherence and long-term outcomes [104].

3.1.5 | Other Dietary Interventions

3.1.5.1 | Q. What Is the Role of Temporary Lactose Restriction in Adults With CeD?. *Recommendation:* Temporary lactose restriction can play a role in adults with CeD who experience persistent gastrointestinal symptoms at diagnosis due to secondary lactase deficiency. Such intolerance is usually transient and resolves as the intestinal mucosa heals on a GFD.

CoE: low; GR: conditional; Agreement: 100%

3.1.5.2 | Q. For Adult Patients With CeD With Persistent Gastrointestinal Symptoms, Is There a Role for a Low-FODMAP Diet?. *Recommendation:* In adults with CeD who have persistent GI symptoms despite a strict GFD and confirmed histological healing, low-FODMAP diet may be considered, provided that other potential causes of symptoms are excluded before starting this additional dietary restriction.

CoE: moderate; GR: conditional; Agreement: 100%

Summary of evidence: Persistent GI symptoms in adult patients with CeD are common, even in those adhering strictly to a GFD. Whilst inadequate dietary adherence and CeD-

TABLE 2 | Overview of methods for assessing adherence to the gluten-free diet.

Method	Details
Dietary assessment and adherence questionnaires	Validated tools evaluate self-reported compliance and understanding of the GFD. They are simple, inexpensive, and widely available, but are subjective and may miss inadvertent gluten exposure [87, 88]. structured dietetic assessment is often more sensitive than questionnaires or serology alone in detecting ongoing dietary transgressions and guiding interventions [87, 89].
CeD serology (e. g., IgA anti-TG2)	Reflects ongoing autoimmune response to gluten. Serology is routinely available and useful for long-term follow-up, but slow normalisation and occasional gluten exposure may not be detected, and negative results do not guarantee mucosal recovery. [90]
Testing for GIPs in stool or urine	GIPs are gluten fragments detectable for up to 4 days in stool and 48 h in urine. When available, GIP testing provides a moderately sensitive and specific real-time measure of recent gluten ingestion and is particularly useful when symptoms persist or adherence is uncertain. Nevertheless, GIPs do not provide a fully reliable measure of inadvertent gluten exposure [91–93].
Other biomarkers	Intestinal fatty acid-binding protein (I-FABP) reflects recent enterocyte injury and correlates with villous atrophy; however, its use remains investigational pending validation in larger studies [94–97]. Faecal calprotectin has inconsistent utility in CeD monitoring [98, 99].
Duodenal biopsy	Remains the gold standard for assessing mucosal healing but is not routinely required for adherence monitoring. See Section 4.5.

Abbreviations: CeD, Coeliac disease; GFD, Gluten-free diet; GIPs, Gluten Immunogenic peptides; I-FABP, Intestinal fatty acid-binding protein; TG2, Transglutaminase 2.

related complications—such as refractory CeD (RCD) or ongoing mucosal inflammation—are important considerations, a substantial proportion of patients may experience symptoms due to coexisting functional GI disorders, particularly irritable bowel syndrome (IBS) [105–108].

Secondary lactose intolerance is frequent at CeD diagnosis due to villous atrophy and reduced lactase activity, affecting up to 50%–70% of untreated patients [109]. Symptoms usually improve with mucosal healing on a GFD. Temporary lactose restriction may be considered in symptomatic patients, with gradual reintroduction once tolerance is restored [110]. Dietitian guidance is essential to ensure adequate calcium and vitamin D intake.

In patients with persistent IBS-type symptoms despite strict GFD adherence and confirmed histological healing, a structured low-fermentable oligo-, di-, mono-saccharides and polyols (low-FODMAP) diet may reduce bloating, abdominal pain, and altered bowel habits [105, 106, 111–113].

A systematic review and meta-analysis found that, when implemented for a minimum of 4 weeks, a low-FODMAP diet was significantly more effective than a standard GFD alone in reducing GI symptoms, particularly bloating, abdominal pain, and altered bowel habits [113].

Before initiating a low-FODMAP diet, other potential causes of persistent symptoms should be systematically excluded, including secondary lactose intolerance, fructose malabsorption, or other food intolerances. Recognition and targeted management of these conditions—through structured dietary assessment, breath testing when appropriate, and specialist dietetic input—can improve symptom control, QoL, and dietary adequacy whilst avoiding unnecessary broader FODMAP dietary exclusions.

The low-FODMAP diet is complex and highly restrictive and without appropriate guidance, there is a risk of nutritional inadequacy, disordered eating behaviours, and increased food-related anxiety. Therefore, it is strongly recommended that any implementation of the low-FODMAP diet in patients with CeD be undertaken only after excluding other potential causes of symptoms—including histological confirmation of remission of gluten-related inflammation—and under the supervision of a dietitian with expertise in both gluten-free and low-FODMAP dietary protocols. Individualized dietary reintroduction and symptom tracking are also essential components of this approach to avoid unnecessary long-term restrictions and hypervigilance [105].

3.2 | Multidisciplinary Support

3.2.1 | Q. Does Psychosocial Support in Patients With CeD Improve Well-Being, Dietary Adherence, and QoL?

Recommendation: Psychological assessment and support may be beneficial for a subset of adults with CeD, particularly those experiencing distress or coping difficulties, as it can contribute to improved well-being, dietary adherence, and QoL.

CoE: low; GR: conditional; Agreement: 95%

Summary of evidence: Adults with CeD do not always adjust easily to the GFD [114]. Beyond the physical implications of the disease, adherence to the GFD can have profound psychological and behavioural consequences. Many individuals experience lasting changes in mood, food-related behaviours, attitudes towards eating, and even their sense of identity [115]. The need for lifelong dietary restrictions may lead to feelings of frustration, social isolation, and emotional distress, particularly in

situations where access to gluten-free options is limited or where individuals feel excluded from shared meals and cultural traditions [77, 86].

Psychological distress in CeD is common, with strong evidence of increased anxiety and depression post-diagnosis, which are both associated with reduced QoL and resilience [71, 116, 117]. Moreover, there is strong evidence of an increased risk of eating disorders in patients with CeD [118, 119]. The strict requirements of a GFD, constant vigilance around food, and anxiety about inadvertent gluten exposure may contribute to restrictive or maladaptive eating behaviours. The risk is higher in women and in those diagnosed during adolescence or early adulthood. Early recognition and multidisciplinary management—including psychological support alongside dietary counselling—are essential to improve adherence, nutritional status, and overall well-being [120–123].

In patients with CeD with affective disorders, psychological support seems to be able to reduce depression and to increase GFD compliance [124]. Furthermore, psychological wellbeing is a predictor of adherence to the GFD [125], so that it appears important to provide psychological support to patients with CeD showing signs of distress in order to ensure good QoL [117, 124, 126, 127].

Whilst current evidence indicates that psychological support can be beneficial—and may even be necessary—for achieving optimal outcomes, this applies only to a subset of CeD patients and should be reserved for those with the psychological sequelae described above.

3.2.2 | How can the Transition of Care for Children or Adolescents With CeD to Adult Care Be Conducted Efficiently?

Recommendation: The guidelines panel suggests a formal transfer of medical care of adolescent with CeD. To facilitate the transfer, a transition letter from paediatricians or a coeliac passport is required, detailing criteria for CeD diagnosis, follow-up, serology, growth data, possible comorbidities, and dietary adherence.

CoE: low; GR: strong; Agreement: 100%

Summary of evidence: Structured transition from paediatric to adult care is recommended to ensure continuity, prevent gaps in follow-up, and support self-management in adolescents with CeD. Expert consensus statements advocate a planned, individualised transition process incorporating preparatory education, assessment of transition readiness, and a formal transfer of essential clinical information through a transition letter or ‘coeliac passport’ [128, 129].

Observational studies suggest improved attendance and self-management with multidisciplinary support and structured transition programmes, although the evidence is heterogeneous and largely non-interventional [130].

Data linking structured transition to long-term outcomes are limited, and current recommendations are mainly consensus-based and extrapolated from other chronic disease models [131].

Practical tools such as transition checklists and coeliac passports are feasible in clinical practise, but further research is needed to identify which components most effectively improve adherence and long-term outcomes [132, 133].

Coeliac Passport—Checklist: [129]

- Main clinical presentation at diagnosis
- Coeliac disease diagnostic criteria (Serology and histology at diagnosis)
- Growth and anthropometric data
- Important lab findings
- Current medications
- Nutritional supplements
- Dietary adherence and education status
- Relevant comorbidities
- Serology and important lab at transition

3.2.3 | Q. What Is the Role of Patient Support Groups in the Management of Adults With CeD?

Recommendation: Patient support groups play a valuable role in the management of adults with CeD by providing education, practical guidance, emotional support, and advocacy, all of which can improve dietary adherence and QoL.

CoE: Not applicable (NA); GR: ungraded good clinical practise statement (UGPS); Agreement: 100%

Summary of evidence: The management of CeD extends far beyond the simple removal of gluten from the diet. It is a life-long journey with substantial practical, emotional, and social challenges. Patient support groups play an indispensable and multi-faceted role in this journey. They provide the essential education, emotional scaffolding, and collective advocacy that empower individuals to manage their condition effectively, improve their dietary adherence, and enhance their overall QoL. As such, healthcare professionals should actively encourage newly diagnosed and existing CeD patients to engage with a reputable support group as a standard component of comprehensive care [134, 135].

3.3 | Management of Associated Conditions and Special Scenarios

3.3.1 | Associated Autoimmune Diseases

3.3.1.1 | Q. How Should Thyroid Function Be Assessed in Adults With CeD? *Recommendation:* Thyroid function should be assessed at the time of CeD diagnosis using a TSH

test, with automatic reflex free T4 testing if TSH is abnormal. Consider repeating thyroid function tests periodically, especially in patients with symptoms or risk factors for thyroid disease.

CoE: low; GR: strong; Agreement: 100%

Summary of evidence: Autoimmune thyroid disorders occur more frequently in individuals with CeD than in the general population, with reported prevalence rates ranging from 2% to 10% for overt thyroid disease and up to 20%–30% for thyroid autoantibody positivity [136, 137]. Untreated thyroid dysfunction can worsen symptoms such as fatigue and weight changes, complicating assessment of response to a GFD.

It is recommended that thyroid function, including serum TSH with reflex free T4 if abnormal, be assessed at CeD diagnosis [129, 138]. Periodic reassessment is strongly advised, especially for symptomatic patients or those with a personal or family history of autoimmune disease.

Although thyroid autoantibodies are frequently positive in CeD, routine antibody testing is not required, as positivity alone does not reliably predict progression to overt thyroid disease. However, selective testing, such as, antibodies against thyroid peroxidase (anti-TPO) and thyroglobulin (anti-TG) may be useful in individuals with abnormal thyroid function tests, symptoms suggestive of thyroid disease, or strong autoimmune predisposition [139]. Any abnormal findings warrant further evaluation, ideally in collaboration with an endocrinologist.

This proactive approach to monitoring is critical, as untreated thyroid dysfunction can exacerbate symptoms like fatigue and weight changes, potentially obscuring the clinical response to a GFD [140]. Early detection and management, ideally with endocrinology input, support overall metabolic stability and patient wellbeing.

3.3.1.2 | Q. Are Additional Dietary Considerations Needed for Patients With CeD Who Also Have Type 1 Diabetes Mellitus?. Recommendation: Patients with CeD and concomitant type 1 diabetes mellitus (T1DM) should follow a dietary plan that integrates gluten-free requirements with the carbohydrate-counting strategies essential for T1DM management. Emphasis on gluten-free carbohydrate sources with a lower glycaemic index may help support more stable glycaemic control.

CoE: Low; GR: strong; Agreement: 100%

Summary of evidence: Patients with CeD and T1DM require specific and coordinated dietary support to address the needs of both conditions [141]. Evidence, particularly in younger patients, indicates that adherence to a GFD supports normal growth and stable BMI without adverse effects on HbA1c or insulin requirements [142, 143]. However, evidence on the impact of a GFD on T1DM-specific outcomes is mixed and varies by population. Meta-analyses and larger trials show no consistent improvement in HbA1c, although smaller studies

suggest potential benefits in selected subgroups, such as preservation of residual beta-cell function in newly diagnosed children [144–146].

Overall, a GFD does not appear to negatively affect growth, with studies confirming stable BMI and height outcomes [147, 148]. The relationship between GFD adherence and QoL is complex, with reports of both improved and reduced QoL, highlighting the psychosocial burden and need for strong support [149, 150]. Patients with coexisting CeD and T1DM may have increased micro- and macrovascular risk, and emerging evidence suggests that GFD adherence may be protective, though data remain limited [151, 152]. Further research is needed to clarify the long-term impact of GFD therapy on diabetes-related complications [143].

Key management considerations include:

- Accurate carbohydrate counting, selection of lower-glycaemic index gluten-free foods, and ensuring nutritional adequacy [153].
- Regular monitoring of glycaemic control and nutritional status is essential, ideally within a multidisciplinary care model involving gastroenterology, endocrinology, and dietitians [154]. The involvement of dietitians with expertise in both CeD and T1DM is highly desirable but may not be universally accessible. This recommendation sets an optimal standard of care, though implementation may depend on available local resources.
- Adherence to a GFD is often suboptimal in this population, underscoring the importance of strengthened follow-up and targeted dietary support [155–157].

3.3.2 | Bone Mineral Density in Adults With CeD

3.3.2.1 | Q. Should Adults With CeD Undergo Assessment of Bone Mineral Density, and Which Individuals Require a DXA Scan?. Recommendation: Adults with CeD are at increased risk of reduced bone mineral density (BMD) and related fractures. Assessing BMD enables timely interventions to support bone health and prevent osteoporosis, including correction of nutritional deficiencies, lifestyle measures, and optimisation of the GFD.

A baseline DXA scan is recommended after 1 year on a GFD for adults with CeD who have additional risk factors for low BMD, including delayed diagnosis, severe malabsorption or marked weight loss at presentation, a history of fragility fractures, other independent osteoporosis risk factors, or Down syndrome.

For adults without these additional risk factors, routine baseline DXA scanning is not mandatory. However, it may be considered in those approaching midlife (> 35–40 years), where low bone mass is more common. FRAX can support individualised fracture-risk assessment and decision-making.

CoE: low; GR: strong; Agreement: 89%

Summary of evidence: A decrease in bone mineral density (BMD) is reported in over 50% of adults with newly diagnosed CeD, and osteoporosis may be the sole presentation or develop later in the disease course [158, 159]. Osteoporosis and osteopenia have been reported in approximately 14% and 39% of patients, respectively, with an increased risk of osteoporotic fractures [160, 161]. CeD-related osteoporosis is associated with underweight, age over 45 years, severity of intestinal damage, and possibly male sex [160, 162, 163]. BMD often improves during the first year of a GFD, although recovery is variable and incomplete in many patients, and fracture risk remains increased compared with the general population [164, 165]. The fracture risk in CeD patients varies from 1.3 to 10-fold more than that in the general population [166–169].

Assessment of bone health should include evaluation of clinical risk factors and biochemical testing, including serum calcium, albumin, alkaline phosphatase, and 25-hydroxyvitamin D, with further investigations in those with low BMD to exclude secondary causes such as osteomalacia [170–173]. DXA and vertebral fracture assessment are useful for fracture risk prediction and monitoring, with minimal radiation exposure [171, 174]. Exposure from a DXA scan is comparable to only a few days of natural background radiation [175].

Although evidence on optimal timing is limited, baseline DXA may be considered at diagnosis or after 1 year of GFD in adults at high risk, including those with delayed diagnosis, malabsorption, significant weight loss, prior fragility fractures, or other osteoporosis risk factors. FRAX may help identify low-risk patients who would benefit from DXA, although further validation in CeD is needed given potential confounding from disease duration, adherence to the GFD, time to resolution of malabsorption, and presence of RCD [176].

Management of osteopenia includes strict adherence to a GFD, correction of vitamin D and calcium deficiency, and lifestyle measures such as weight-bearing exercise, smoking cessation, and moderation of alcohol intake. Pragmatically, many adults with CeD may benefit from ensuring a daily intake of approximately 1000 IU of vitamin D and achieving a calcium intake of around 1000 mg/day (preferably through diet, with supplementation when dietary intake is insufficient), in line with general bone-health recommendations. Repeat DXA should be considered based on individual risk profile.

Drug therapy should be reserved for patients with progressive bone loss or high fracture risk, following standard osteoporosis guidelines and individual risk assessment. Importantly, a low T-score alone—particularly in younger adults—should not automatically trigger pharmacotherapy; conversely, some older adults with osteopenia may warrant treatment based on overall fracture risk. Treatment strategies may differ between males and females, and premenopausal women require special consideration [177, 178]. In individuals with persistent diarrhoea, parenteral medication may be necessary [179].

Specialist referral is recommended when bone loss is unexplained, comorbid endocrine or musculoskeletal disorders are present, or management decisions are uncertain.

Key Points:

- Low bone mineral density is common in adults with CeD, and fracture risk is increased.
- Assess clinical risk factors and check calcium, vitamin D, and related biochemical markers.
- DXA is indicated in high-risk adults and may be considered in those > 35–40 years depending on individual risk.
- FRAX can help guide DXA decisions in low-risk patients, though further validation in CeD is needed.
- Management includes strict GFD, adequate calcium/vitamin D, and regular weight-bearing/resistance exercise.
- Correct vitamin D and calcium deficiency to avoid osteomalacia before considering osteoporosis therapy.
- Pharmacotherapy should follow country-specific guidelines and individual fracture risk—not T-score alone.
- Refer to an osteoporosis specialist when bone loss is unexplained or comorbidities complicate management.

3.3.3 | Immune Dysfunction and Vaccination

3.3.3.1 | Q. Which Patients With CeD Require Pneumococcal Vaccination? *Recommendation:* Pneumococcal vaccination is recommended for adults with CeD with evidence of functional asplenia, concurrent autoimmune disorders or RCD-II, as well as for patients over 65 years.

CoE: NA; GR: UGPS; Agreement: 100%

Summary of evidence: Functional asplenia or hyposplenism is diagnosed by the presence of Howell-Jolly bodies and pitted erythrocytes in the peripheral blood smear or radiological conformation of splenic atrophy.

Functional asplenia and/or radiological evidence of splenic atrophy is observed in 19%–43% of uncomplicated CeD cases, in 59% of those with associated autoimmune disorders, and in over 80% of patients with RCD-II or enteropathy-associated T-cell lymphoma (EATL) [180–183]. The duration of gluten exposure before diagnosis and older age at diagnosis are key prognostic factors for developing functional asplenia [184].

Because of hyposplenism, patients with CeD have a higher risk of sepsis, particularly from encapsulated bacteria such as *Streptococcus pneumoniae*, compared with the general population [185, 186]. Up to 61% of patients with CeD—including

those adhering to a GFD and those without Howell–Jolly bodies or pitted erythrocytes—show functional defects in IgM memory B cells [187]. Although a GFD can reverse functional asplenia, it does not restore structural splenic atrophy [188, 189].

Adherence to vaccination guidelines markedly reduces the incidence of severe infections and contributes to improved overall health and QoL [190–193].

3.3.4 | Management of Specific Extra-Intestinal Manifestations

Recommendation: We recommend that significant extra-intestinal manifestations of CeD be managed by a multidisciplinary team. This should include a gastroenterologist and dietitian to oversee the GFD and intestinal health, alongside a specialist with expertise in the specific manifestation (i.e., neurologist, dermatologist, dentist) for targeted diagnosis and management.

CoE: NA; GR: UGPS; Agreement: 100%

Summary of Evidence: A multidisciplinary approach is the standard of care for specific extra-intestinal manifestations of CeD, as these conditions require specialized diagnostic and therapeutic expertise beyond the scope of gastroenterology alone. The GFD remains the foundational, shared treatment goal, but collaboration is essential for optimal patient outcomes. The management of specific extra-Intestinal manifestations is summarised in Table 3.

3.3.5 | Concurrent GI and Metabolic Issues

3.3.5.1 | Q. Would CeD Patients Benefit From Pancreatic Enzyme Supplementation?. *Recommendation:* In adult patients with CeD, pancreatic enzyme replacement therapy (PERT) may be indicated when exocrine pancreatic insufficiency is confirmed. PERT can improve digestive function, relieve symptoms such as steatorrhea and bloating, enhance nutrient absorption, and improve QoL.

TABLE 3 | Management of specific extra-intestinal manifestations.

3.3.4.1. Neuro-Coeliac
The management of neurological manifestations (e.g., gluten ataxia, peripheral neuropathy) in CeD necessitates a strict GFD, involving a multidisciplinary team (gastroenterology, neurology, dietetics). Neurological assessment to define deficits, exclude alternative causes, monitor progression, and manage symptoms. Early establishment of a gluten-related aetiology is important, as recovery may be only partial [194, 195].
3.3.4.2. Dermatitis herpetiformis (DH) and other dermatological associations
DH: Lifelong strict GFD (cornerstone therapy); dermatology-led diagnosis (skin biopsy) and short-term pharmacologic treatment (e.g., dapsone) [196–198]. multidisciplinary care with gastroenterology to assess enteropathy, monitor treatment-related adverse effects, and reinforce GFD adherence [196]. Other dermatological associations: Higher prevalence in CeD (e.g. psoriasis, atopic dermatitis, alopecia areata, chronic urticaria); not primarily gluten-dependent, though partial improvement with GFD reported in some patients [199, 200]. Management is primarily dermatology-led, with gastroenterology involvement when CeD is suspected or confirmed.
3.3.4.3. Oro-Dental care
Recurrent aphthous stomatitis and other oral lesions may occur in adult CeD [201]. Dental involvement to identify suggestive oral signs, manage oral lesions, and reinforce GFD adherence. Address oral complications related to nutrient deficiencies and associated autoimmune conditions [202, 203].
3.3.4.4. Chronic fatigue
Fatigue is a common extra-intestinal manifestation of CeD. Management includes strict GFD adherence, assessment and correction of micronutrient deficiencies (e.g. iron, folate, B12), and evaluation for comorbid conditions (e.g. thyroid disease, depression). Psychological support may be beneficial to address behavioural and lifestyle factors. [204, 205]
3.3.4.5. Coeliac hepatitis/Autoimmune hepatitis
Coeliac hepatitis presents with abnormal liver tests and/or histology that resolve with a strict GFD after exclusion of other causes [206, 207]. Coexisting autoimmune liver diseases (e.g. AIH, PBC, PSC) are not GFD-responsive and require appropriate hepatology-led evaluation. Coexistent AIH in CeD requires a GFD alongside corticosteroids and, when required, longer-term immunosuppressive therapy. Shared care ensures that GFD adherence is optimized, the liver disease is accurately characterized, and timely treatment is provided to improve long-term hepatic and systemic outcomes [207, 208].
3.3.4.6. IgA nephropathy
Association with CeD remains uncertain [209, 210]. Nephrology-led management is recommended, with gastroenterology involvement when CeD is confirmed or strongly suspected. A GFD may be considered in confirmed CeD, with possible improvement in renal parameters reported in some patients.

Abbreviations: AIH, autoimmune hepatitis; CeD, Coeliac disease; DH, Dermatitis herpetiformis; GFD, gluten-free diet; PBC, primary biliary cholangitis; PSC, primary sclerosing cholangitis.

CoE: moderate; GR: conditional; Agreement: 89%

Summary of evidence: Exocrine pancreatic insufficiency (EPI) is a recognized cause of persistent GI symptoms in patients with CeD, particularly those who do not respond fully to a GFD [211]. In newly diagnosed CeD, EPI prevalence can reach 26%, likely from enterocyte damage and pancreatic hypostimulation [212]. However, in those patients with persistent GI symptoms despite adherence to a GFD, the prevalence of EPI remains high (28%) [212].

The pathophysiology of PEI in CeD is complex. Proposed mechanisms include mucosal damage and a reduction in the absorbing surface due to duodenal intestinal villi atrophy in CeD, which reduce the secretion of the gastrointestinal hormone CCK-pancreozymin and thusly prejudice intraluminal fat digestion because the secretory and motor functions of the target organs, the pancreas and the gallbladder, are impaired [213].

When EPI is confirmed, pancreatic enzyme replacement therapy (PERT) significantly improve digestive function. PERT alleviates symptoms, particularly steatorrhoea and bloating, enhances nutrient absorption, and improve QoL [214]. European PEI guidelines recommend considering PEI testing in CeD patients with significant malnutrition at diagnosis or persistent symptoms despite a GFD, although supporting evidence remains limited, highlighting an important knowledge gap [215].

3.3.5.2 | Q. In Patients With CeD, Do Probiotics Improve Symptoms and QoL When Added to a GFD?. *Statement:* Current evidence is insufficient to support or oppose the use of probiotics alongside a GFD for improving symptoms and QoL in patients with CeD. Further research may determine their effectiveness.

CoE: NA; GR: UGPS; Agreement: 95%

Summary of evidence: In general, there is no consensus on the role of various probiotics, and there is still controversy about the safety of probiotics. A systematic review and meta-analysis found that probiotics significantly improved GI symptoms in patients with CeD. However, there was no significant improvement in QoL due to insufficient data [216].

Another meta-analysis reported that probiotics increased the abundance of beneficial bacteria and noted that probiotics might alleviate GI symptoms and improve immune response in CeD [217]. Small sample sizes, varying probiotic strains, variable periods of GFD, differing methodologies and absence of well-designed, large-scale studies often contribute to the inconsistency of the results.

3.3.5.3 | Q. Are Adults With CeD on a GFD at Higher Risk of Developing Metabolic Syndrome?. *Recommendation:* adults with CeD on a GFD have a higher risk of developing metabolic syndrome compared to individuals with CeD before dietary therapy. A balanced GFD, active lifestyle, and regular monitoring for metabolic risk factors—including obesity, hypertension, dyslipidaemia, and insulin resistance—are recommended.

CoE: low; GR: strong; Agreement: 100%

Summary of evidence: Evidence suggests that patients with CeD are not at increased risk of metabolic syndrome at diagnosis compared with the general population. In fact, the prevalence of metabolic syndrome in newly diagnosed, treatment-naïve CeD patients is substantially lower (approximately 3%–4%) than in the general adult population, where reported metabolic syndrome prevalence ranges from 26% in Canada to 38%–42% in the United States, and 24%–33% across Europe, depending on population age and diagnostic criteria. Similarly, and metabolic dysfunction-associated fatty liver disease (MASLD) is uncommon at CeD diagnosis, with reported rates around 1.7% [218–220].

However, after initiation of a GFD, both metabolic syndrome and MASLD become more prevalent. A study demonstrated that the prevalence of metabolic syndrome and MASLD increased after GFD [218]. A systematic review of 11 studies ($n = 2578$) revealed that patients with CeD have a 18.2% prevalence of MASLD and 4% prevalence of metabolic syndrome at diagnosis [219]. These prevalence rates are actually lower than those reported in the general population in Western countries, but increase to 28.2% and 21.3%, respectively, after initiating a GFD, approaching the prevalence in the general population [221].

The increase in prevalence of metabolic syndrome following GFD initiation is thought to reflect improved nutrient absorption, weight gain, and changes in body composition, which occur as intestinal healing progresses. Regular clinical evaluation, metabolic monitoring, dietary counselling, and encouragement of physical activity are essential to mitigate these risks [207, 219, 222–225].

3.3.6 | Special Scenarios

3.3.6.1 | Q. Do Pregnant Women With CeD Require Specialized Care?. *Recommendation:* Although evidence is limited, coordinated preconception and perinatal care may help optimize nutritional status in women with CeD and support a healthy environment for early foetal development. Postpartum care should continue to emphasize maintaining the GFD.

CoE: NA; GR: UGPS; Agreement: 95%

Summary of evidence: Available data, although limited, suggest that women with CeD may be at increased risk of adverse pregnancy outcomes. A systemic review reported that women with CeD are at significantly higher risk of developing preterm birth, intrauterine growth retardation, stillbirth, low birth weight, and being small for gestational age [226]. Malnutrition and the autoimmune mechanism are considered two possible causes [227].

Maternal CeD may be a risk factor for deficiency of nutrients during pregnancy [228]. In women with CeD planning pregnancy, strict adherence to a GFD is essential, together with resolution of disease-related symptoms and adequate supplementation of folate and other vitamins, to optimise nutritional

status and create a favourable environment for preimplantation, early foetal development, and placental formation [229]. Additionally, during pregnancy, care is needed to optimize nutrient needed of both the mother and the infant to protect them from complications [230]. Adherence to a GFD has been associated with improved reproductive outcomes [230–232].

3.3.6.2 | Q. What Are the Key Management Considerations for Patients Diagnosed With CeD at an Older Age?

Recommendation: In patients diagnosed with CeD at an older age, non-classical symptoms are common, often causing diagnostic delays. These patients have an increased risk of complications, including osteoporosis and malignancy. Although response to a GFD may be slower, it provides substantial clinical benefit and should be strongly considered.

CoE: NA; GR: UGPS; Agreement: 100%

Summary of evidence: CeD in the elderly (typically > 65 years) frequently presents with non-classical features such as iron-deficiency anaemia, fatigue, and bloating, rather than classic diarrhoea, often leading to a significant diagnostic delay [233–237]. This often leads to a significant diagnostic delay in diagnosis.

The diagnostic criteria for CeD in the elderly are the same as for other adults. The severity of villous atrophy at diagnosis is generally comparable to that in younger adults. Clinicians should have a low threshold for testing for CeD in elderly individuals with suggestive symptoms or associated conditions.

Patients diagnosed with CeD at an older age have a higher burden of specific complications:

- Metabolic Bone Disease: A high prevalence of osteopenia and osteoporosis, necessitating BMD assessment.
- Malignancy: A significantly increased risk of EATL and other malignancies.
- Functional Hyposplenism: This is more common in elderly CeD patients, with implications for vaccination.
- Other Autoimmune Diseases: A higher prevalence of associated autoimmune conditions is often seen.

A strict, lifelong GFD is the primary treatment. Whilst elderly patients may experience a slower or incomplete clinical and histological response due to long disease duration and practical challenges, the initiation of a GFD leads to significant symptomatic improvement, enhanced QoL, and correction of nutrient deficiencies. Therefore, diagnosis and dietary treatment are strongly recommended and beneficial, regardless of age.

3.3.6.3 | Q. How to Screen Family Members of Patients With CeD Patients?. *Recommendations:*

1. HLA-DQ2/8 genotyping is recommended as the initial screening step primarily in children of patients with CeD—where it can prevent repeated testing—while anti-TG2

serology remains the most cost-effective and widely available initial test for adults and lower-risk relatives.

2. Follow-up: For first-degree relatives who are seronegative at initial assessment, periodic antibody follow-up (e.g., every 4–5 years) may be considered based on individual risk factors and new symptoms.

CoE: moderate; GR: conditional; Agreement: 95%

Summary of evidence: The recommendations for screening family members of patients with CeD, including cost-effectiveness across family relative groups, are summarised in Table 4.

First-degree relatives (FDRs) of CeD patients have a 5%–10% disease risk, highlighting a need for targeted screening [238–240]. Genetic susceptibility, primarily conferred by HLA-DQ2 and/or HLA-DQ8 haplotypes, plays a central role in disease development. HLA genotyping is therefore a useful first-line risk stratification tool, particularly in children, as the absence of HLA-DQ2/8 effectively excludes CeD [241]. Rare cases linked to alleles like HLA-DQ7.5 carry a residual risk below 1% [242, 243].

Amongst genetically susceptible individuals, HLA-DQ2 homozygosity is associated with a substantially increased risk of developing CeD and with earlier disease onset [244]. Prospective cohort studies and recently developed risk prediction models suggest that combining HLA genotype with age, sex, and serological data may help individualise decisions regarding the timing and frequency of serological re-screening in at-risk relatives. These emerging data indicate that such personalised approaches may be preferable to uniform re-screening intervals [245].

Twin studies further support a strong genetic contribution, with concordance rates of approximately 70% in monozygotic twins compared with around 9% in dizygotic twins within 5 years of follow-up [246]. These findings justify periodic monitoring of genetically susceptible relatives.

In children who are FDRs of patients with CeD, initial HLA genotyping can avoid unnecessary long-term follow-up in those who are HLA-DQ2/8 negative. DQ2/8-positive children should undergo periodic serological screening, as disease presentation is frequently asymptomatic [241].

In adult relatives of patients with CeD, serological testing—specifically IgA anti-TG2 with total serum IgA—is recommended as the primary screening approach [247]. Early identification may prevent complications such as nutritional deficiencies, metabolic bone disease, and the development of associated autoimmune conditions.

Second-degree relatives of patients with CeD have a lower, but still increased, risk compared with the general population [239, 244]. Whilst universal screening of second-degree relatives is not recommended, serological testing should be considered in the presence of suggestive symptoms, associated autoimmune conditions (e.g., T1DM or autoimmune thyroid disease), or

TABLE 4 | Summary of the screening methods across family relative groups together with cost-effectiveness considerations.

Family relative group	Recommended approach	Cost-effectiveness rationale
Children (FDR)	<i>HLA-DQ2/8 genotyping</i> is preferred when it is accessible and affordable, as a first-line risk stratification tool. HLA-negative children do not require further follow-up.	The pre-test probability is <i>high</i> . The upfront cost of HLA testing is offset by eliminating decades of repeated serology in HLA-negative children, making it cost-saving over the long term.
Adults (FDR)	<i>Serology</i> is preferred in most adult FDRs due to lower cost and broad availability. <i>HLA-DQ2/8 genotyping</i> is reserved for selected situations, such as equivocal serology, IgA deficiency, individuals already on a GFD, or complex family histories.	The pre-test probability of CeD is moderately elevated. Adults have already passed much of the highest-risk period; repeated long-term screening is less likely. Serology is cheaper and widely available.
Second-degree relatives	<i>No universal screening is recommended.</i> Serological testing (IgA anti-TG2 with total IgA) should be considered selectively, particularly in the presence of symptoms suggestive of CeD or associated autoimmune diseases (e.g., T1DM, autoimmune thyroid disease).	The pre-test probability of CeD is low. Universal screening is not cost-effective due to low disease prevalence; HLA testing adds little value and increases cost.
Families with multiple CeD cases	<i>Screening should follow age- and genotype-based principles</i> , rather than family history alone. In children, HLA-DQ2/8 genotyping may help stratify individual risk and guide follow-up. In adults, serology remains the first-line screening test.	Recent large cohort studies indicate that multiple affected family members do not constitute an independent risk factor once HLA genotype is accounted for. Cost-effectiveness depends on individual genetic risk and anticipated need for future screening, rather than family history alone.

Abbreviations: CeD, Coeliac disease; FDR, First-degree relatives; GFD, Gluten-free diet; HLA, Human Leucocyte antigen; T1DM, Type 1 diabetes mellitus.

other clinical indications [239, 248]. Recent large cohort studies indicate that the presence of multiple affected family members alone does not constitute an independent risk factor once HLA genotype is taken into account, and should therefore not be used as a sole criterion for screening [244, 245].

Finally, consider serological follow-up every 4–5 years for initially negative FDRs with persistent symptoms or strong family history, accounting for possible later-onset disease.

3.3.6.4 | Q. How to Manage Adults Presenting With the Rare Severe Presentation ‘Coeliac Crisis’? *Recommendation:* Although rare in adults, coeliac crisis—often a manifestation of refractory CeD type 2—requires urgent admission for multidisciplinary care to address dehydration, electrolyte/nutritional deficits, and precipitating factors, whilst promptly initiating a strict GFD.

CoE: NA; GR: UGPS; Agreement: 95%

Summary of Evidence: In adults, CeD may rarely present as a severe condition known as a ‘coeliac crisis’, which can be life-threatening if not promptly recognized and managed [249]. This is usually in the context of refractory CeD type 2.

Clinically, patients have a severe malabsorption with electrolyte disturbances, metabolic acidosis, dehydration, hypoalbuminemia and anaemia [250, 251]. A systematic review showed that amongst 42 adults with CeD crisis, the median age was 50 years (range 23–83), with a 2:1 female to male ratio. In the majority of cases (88%), the coeliac crisis most often presents as the initial

manifestation of CeD, whilst the remaining were previously diagnosed CeD cases reporting non-adherence to a GFD [250]. Precipitating factors (such as: trauma, surgery, infections) are present in about 25% of cases [250].

Management is multidisciplinary and includes in-hospital supportive care, with intravenous fluid administration, correction of electrolyte imbalances and nutritional deficiencies [251, 252]. The GFD should be started promptly [249, 250].

3.3.6.5 | Q. What Is the Recommended Management for Patients Who Develop CeD-like Enteropathy in Association With Immune Checkpoint Inhibitors? *Recommendation:* In patients receiving immune checkpoint inhibitors (ICI) who develop diarrhoea, coeliac serology should be obtained. If serology is positive, histological confirmation of CeD should be sought. Conversely, if a CeD-like enteropathy is identified histologically in a patient on ICI, coeliac serology should be performed to confirm the diagnosis.

First-line management is a strict GFD when CeD is confirmed. Systemic immunosuppression or modification of immunotherapy should be reserved for severe presentations or cases not responding to a GFD, with decisions guided by multidisciplinary discussion.

CoE: low; GR: conditional; Agreement: 95%

Summary of Evidence: ICI-CeD is a rare immune-related adverse event from immunotherapy, supported mainly by case reports and small studies [253–255]. ICIs can unmask or trigger CeD as

a rare immune-related adverse event (irAE). Patients typically present with diarrhoea, weight loss, and malabsorption, with histology showing villous atrophy similar to classic CeD. Whilst serology is often positive, seronegative cases require duodenal biopsy and exclusion of other causes [253].

Most patients respond significantly to a strict GFD, often avoiding systemic steroids. This response helps distinguish it from other ICI-related enteropathies (e.g., colitis), which typically require immunosuppression [253, 256]. Accurate diagnosis is crucial to prevent unnecessary interruption of cancer therapy.

Due to sparse, low-certainty evidence, firm recommendations are limited. However, a GFD is consistently effective first-line therapy for both seropositive and carefully evaluated seronegative cases. Immunosuppression should be reserved for dietary failure or suspected overlapping immune-related toxicity [254, 257].

3.4 | Pharmacological Therapies

3.4.1 | Q. What Is the Current Status of Non-dietary Medical Treatments for CeD?

Statement: Non-dietary pharmacological treatments for CeD hold promise for the future but remain experimental and are currently available only within clinical trials.

CoE: moderate; GR: strong; Agreement: 100%

Summary of evidence: Since adherence to a GFD can be difficult and some patients have persistent symptoms or mucosal damage despite a GFD, there is growing interest in non-dietary medical treatments for CeD.

Several therapeutic approaches target various aspects of CeD pathophysiology are under investigation [258, 259]. However, broader validation through phase III trials is still necessary.

Clinical trials often reveal discrepancies between symptoms, serological markers, and mucosal biopsy results, highlighting the need for further research. Additionally, many results and potential pitfalls remain unpublished. Existing studies also have methodological limitations, particularly regarding patient selection. Most trials have enrolled patients with CeD with GI symptoms and duodenal mucosal damage, often linked to poor GFD adherence, which may persist throughout the trial [260–262].

Enzyme-based over-the-counter supplements that claim to degrade gluten are not approved therapies for CeD and should not replace a strict GFD; their efficacy and safety have not been established in rigorous clinical trials [263, 264].

In conclusion, non-dietary treatments for CeD hold promise for the future but remain experimental and are currently available only within clinical trials.

4 | Long-Term Follow-Up and Monitoring of CeD in Adults

4.1 | Q. Why Is Long-Term Follow-Up Recommended for Adults With CeD?

Recommendation: Long-term follow-up is recommended to monitor dietary adherence, detect complications and comorbidities, and provide ongoing nutritional and psychosocial support, which helps ensure sustained disease control and optimised long-term outcomes.

CoE: NA; GR: UGPS; Agreement: 95%

Summary of evidence: For patients with CeD, a lifelong GFD is not a voluntary lifestyle choice, it is a necessity and simply avoiding gluten-containing products after diagnosis is not enough. Follow-up is relevant in the long-term, given that persistent symptoms and mucosal changes occur in 20%–40% of adult patients with CeD [219, 265, 266].

Strict GFD is expected to reduce symptoms and normalize biomarkers (biochemical, serological and histological) that were abnormal at diagnosis. However, neither symptom improvement nor biochemical or serological testing may reliably predict CeD activity or histological recovery [23, 267].

Dietary adherence improves with regular follow-up in a specialist coeliac clinic, where structured dietetic support is a key component of care [268–270]. Patients should be encouraged to join national coeliac societies or other disease-specific patient support groups.

Follow-up presents a valuable opportunity to proactively detect and manage associated autoimmune disorders, address complications like bone disease, and recognize warning signs that may suggest the development of RCD or malignancies [269].

Key endpoints at follow-up include the resolution of symptoms and evidence of disease remission. Mucosal healing—defined as normalization of villous architecture, absence of intraepithelial lymphocytosis, and resolution of crypt hyperplasia (Marsh 0–I)—is an important therapeutic goal, as it is associated with improved long-term outcomes. However, histological reassessment is not required in all patients and should be individualized based on clinical status, serological response, and risk factors for persistent disease activity [271]. By maintaining a strict GFD and regular follow-up with healthcare providers, most patients can achieve sustained health and good QoL [272].

4.2 | Q. What Is the Most Appropriate Healthcare Setting for the Long-Term Follow-Up of Adults With CeD, and How Should Responsibilities Be Shared Between Primary and Secondary Care?

Recommendation: Long-term follow-up of adults with CeD is best coordinated through hospital-based outpatient clinics or

specialized coeliac centres, where feasible. For stable patients, follow-up may be conducted in primary care, provided there is clear access to specialist support. Digital tools can be utilised to support remote monitoring and dietary adherence.

CoE: NA; GR: UGPS; Agreement: 95%

Summary of evidence: No universal consensus exists on who should provide long-term care for adults with CeD [269]. Creating international standards is difficult due to differing healthcare systems, resources, and access.

Follow-up has traditionally been based in hospital clinics or specialized centres. However, these centres often lack the capacity to meet the growing demand associated with the increasing prevalence of CeD.

For clinically stable, patient with good dietary adherent, primary care can manage follow-up if specialist and dietitian support is available [68, 273]. Dietitian-led care with physician oversight is effective and cost-efficient [68, 273]. Telemedicine and digital tools can aid monitoring and adherence without overburdening hospitals [274, 275].

In several European countries, primary care commonly handles long-term follow-up, supported by higher patient volumes. Successful decentralization requires adequate provider training and easy access to specialist consultation. The hospital team must clearly communicate to primary care providers the management plan, including key follow-up points, recognition of red flags for complications, and criteria for referral back to specialist care [268].

4.3 | Q. Should Follow-Up of Adults With CeD Be Conducted at Fixed-Intervals or Tailored to the Individual Through a Patient-Centred Approach?

Recommendation: Both fixed-interval and individualised strategies for follow-up of adults with CeD have advantages; however, follow-up should be individualised and patient-centred, taking into account disease activity, treatment adherence, comorbidities, and individual needs.

CoE: NA; GR: UGPS; Agreement: 100%

Summary of evidence: The optimal interval between follow-up visits for adults with CeD remains unclear and has not been systematically studied [269]. Whilst a strictly structured follow-up at fixed intervals may not always be feasible in routine clinical practise, it provides a valuable framework to standardize care and ensure consistent monitoring. Conversely, an individualized, patient-centred approach—considering factors such as the patient's risk profile, serological response, dietary adherence, symptom evolution, and personal preferences—is both reasonable and practical. Both strategies have their merits and may be applied flexibly based on clinical context.

A structured follow-up framework as outlined in Table 5 may be implemented.

Tailored follow-up is necessary to address individual needs and comorbidities, ensuring optimized health outcomes [4, 269]. Key factors for successful implantation of this strategy are outlined in Table 6.

4.4 | Q. What Is the Role of Coeliac Serology in Follow-Up of Adult Patients With CeD?

Statement: The role of IgA anti-TG2 in follow-up is to identify ongoing gluten exposure; a positive IgA anti-TG2 result in patients with CeD on a GFD suggests potential poor dietary adherence or gluten contamination, whilst a negative result does not confirm strict adherence or the absence of gluten exposure and it is not a reliable marker of villous atrophy.

CoE: moderate; GR: strong; Agreement: 95%

Summary of evidence: CeD-specific antibodies, particularly IgA anti-TG2, typically decline within months of starting a GFD and often normalize within the first year [281–284].

Persistently positive, or not decreasing, IgA anti-TG2 levels are predictive of some degree of gluten intake. The sensitivity of IgA anti-TG2 for the detection of diet transgressions evaluated by patient self-reports or dietitian-led assessments was low (52%–57%) [282, 285]. This illustrates that negative IgA anti-TG2 levels should not be considered a marker of strict dietary compliance, nor do they reflect histological recovery [23, 267, 286]. IgA anti-EMA and IgG anti-DGP perform similarly to IgA anti-TG2 detection in this setting [281]. With regard to patients with CeD and IgA deficiency, IgG anti-TG2 levels also decline slowly over time on a GFD but fail to reach normalization in up to 80% of cases despite long-term strict diet adherence, limiting its utility as a standalone marker of compliance [287, 288].

4.5 | Q. Is a Follow-Up Duodenal Biopsy Necessary in Adults With CeD?

Recommendation: A follow-up duodenal biopsy is not routinely necessary for all adults with CeD. It is recommended in a personalized manner, guided by factors such as age at diagnosis, symptom severity, serological status, and clinical response to the GFD. A biopsy should be considered if symptoms persist or worsen.

CoE: low; GR: conditional; Agreement: 95%

Summary of evidence: In adults, neither symptoms nor serology is reliable to predict small-bowel damage. The need for routine follow-up duodenal biopsies to monitor CeD is debatable, as there is insufficient evidence that this practise impacts clinical outcomes [267, 289]. Studies on mucosal healing after a GFD show that the process can be slow or incomplete, with some studies reporting healing in only 30% of patients after 3–5 years [46, 290].

Currently, there are no studies indicating an absolute necessity for performing routine follow-up biopsy for all patients.

TABLE 5 | Suggested follow-up scheme for adults with CeD.

Contact (timing, provider, mode) ^a	Assessments and interventions
At diagnosis (physician and dietitian), face-to-face consultation	- Physical examination including body mass index (BMI) - Coeliac serology (if not previously obtained at diagnosis)^b - Routine tests (complete blood count, iron status, folate, B12, thyroid function tests, liver enzymes, albumin, calcium, phosphate, vitamin D, blood glucose) - Education on CeD and dietary counselling by a CeD dietitian - Discuss family screening - Advise membership of a coeliac national society or support group - Check the indication for bone densitometry and decide on future timing
At 4–6 months (physician and dietitian), face-to-face consultation	- Assess symptoms and coping strategies - Repeat routine tests (if previously abnormal) - Dietary review - Offer referral to a psychologist (in case of evident coping issues)
At 12 months (physician and dietitian), face-to-face, telephone, video consultation or via digital app (if available)	- Assess symptoms - Physical examination (as indicated) - Coeliac serology - Repeat routine tests (if previously abnormal) - Dietary review
At 24 months (physician or dietitian), by telephone, video consultation or digital app (if available), thereafter every 1–2 years, depending on dietary adherence, level of education, and patient confidence in self-management	- Assess symptoms - Dietary review (optional) - Coeliac serology (if negative seroconversion is already achieved then it is optional) - Thyroid function tests - Other tests as clinically indicated (e.g., deficiencies, testing for GIPs when indicated and if available)

Abbreviations: BMI, body mass index; CeD, Coeliac disease; GIPs, Gluten Immunogenic Peptides.

^aMethods of contact depend on local resources.

^bSerology does not need to be routinely repeated within the first year of a gluten-free diet.

However, there is a need for distinguishing asymptomatic patients with negative serology from symptomatic patients who need repeated biopsies to rule out RCD or malignancy [291]. There are data suggesting a more personalized follow-up, wherein the follow-up biopsy is conducted after a few years and only for a selected group based on age, initial disease severity and response to the GFD [292].

The indication for a follow-up biopsy in patients with CeD adhering to a strict GFD should be discussed with the patient, taking into consideration their preferences and choices. Common indications for a follow-up biopsy include:

- Persisting or worsening symptoms and/or biochemical or laboratory evidence of malabsorption.
- Development of new red flag symptoms raising suspicion of complications.
- In adults diagnosed with CeD after the age of 45 or those with initially severe presentation, a follow-up biopsy after 1–2 years on a GFD may be reasonable to assess mucosal healing.
- In seronegative CeD, follow-up biopsies are essential for accurate diagnosis and monitoring recovery.

TABLE 6 | Key Factors Influencing tailored follow-up in Adults with CeD.

Factor	Consideration
1. Symptom severity and presentation	Patients with persistent symptoms or nutritional deficiencies may require more frequent follow-up and additional testing. Asymptomatic individuals may need fewer visits but still require periodic assessments to ensure adherence to a GFD and to monitor for silent complications.
2. Serological testing and intestinal healing	Patients with persistently elevated CeD antibodies may require more frequent monitoring to evaluate adherence or identify ongoing gluten exposure and may benefit from dietary counselling or psychological support to help maintain a strict GFD. Those who have achieved serological remission and demonstrated intestinal healing may benefit from less frequent follow-ups.
3. Age and comorbidities	Older adults or those with coexisting conditions may require integrated management of non-CeD-specific health risks, including cardiovascular disease. Whilst CeD has been associated with a modestly increased relative cardiovascular risk in some studies, absolute risk in treated disease is low and is largely driven by traditional cardiovascular risk factors and dietary quality [276, 277]. Accordingly, cardiovascular health should be addressed as part of holistic long-term follow-up, focussing on optimisation of nutrition, promotion of physical activity, and management of conventional risk factors in line with general population recommendations. Younger patients without comorbidities may benefit from less frequent visits focussed on education and prevention. Monitoring for the development of comorbidities during follow-up remains important.
4. Dietary quality	An unhealthy and nutritionally unbalanced GFD may be harmful, leading to micronutrient deficiencies, undesirable weight gain, and the development of metabolic syndrome, MAFLD, and cardiovascular complications [218, 219]. Current evidence does not demonstrate a clear, direct effect of a GFD per se on reducing cardiovascular outcomes [278, 279]. The impact of a GFD on cardiovascular risk factors is heterogeneous and does not consistently translate into improved cardiovascular outcomes [278, 279]. Dietary counselling should therefore emphasise not only gluten exclusion, but also overall dietary quality, nutritional adequacy, and a balanced dietary pattern, alongside encouragement of regular physical activity.
5. Coexistence of autoimmune conditions	The presence of autoimmune conditions (e.g., autoimmune thyroid disease, T1DM, Addison's disease, vitiligo, alopecia, hypogonadism, chronic autoimmune gastritis, systemic lupus erythematosus) requires integrated management and input from relevant specialists such as endocrinologists [138, 248, 280].
6. Individual preferences	Preferences regarding visit frequency and level of support should be considered. Some patients may prefer regular follow-up for reassurance, whilst others may opt for a more independent approach.

Abbreviations: CeD, Coeliac disease; GFD, gluten-free diet; MAFLD, metabolic dysfunction-associated fatty liver disease; T1DM, type 1 diabetes mellitus.

- A biopsy may be performed at the patient's request for reassurance that mucosal healing has been achieved.

5 | Complicated CeD

5.1 | Kinetics of Clinical, Serologic and Histologic Response to a GFD in Adults With CeD

5.1.1 | Q. When Is a Clinical Response to the Diet to Be Expected in Adults With CeD?

Statement: Clinical response to a GFD in adults with CeD shows substantial inter-individual variability, with symptom improvement generally expected within 4 weeks to 4–5 months after diet initiation.

CoE: low; GR: strong; Agreement: 100%

Summary of evidence: Evidence describing the symptomatic course after initiating a GFD in adults with CeD is limited, as few studies have prospectively evaluated the kinetics of clinical

response. Available prospective data largely support clinical experience. Symptomatic improvement shows marked inter-individual variability: some adults experience relief within 4 weeks of starting a GFD, whereas others may require 4–5 months before a clear response becomes evident [284]. In a cohort evaluated at the time of follow-up biopsy—performed after highly variable intervals averaging 2–3 years—82% reported good symptomatic improvement, independent of histological recovery [293]. This highlights that clinical response does not reliably predict mucosal healing. Additionally, a minority of patients may show a delayed symptomatic response 'late responders', sometimes occurring more than 1 year after starting the GFD [294, 295].

5.1.2 | Q. When Is a Serological Response to the Diet to Be Expected in Adults With CeD?

Statement: A decline in IgA anti-TG2 can be observed as early as 2–4 weeks after starting a GFD, and in the majority, titres normalize within approximately 12 months. There is, however, strong inter-individual variation in this response.

CoE: moderate; GR: strong; Agreement: 100%

Summary of evidence: Once a GFD is initiated, a reduction in IgA anti-TG2 titres is expected. Some prospective studies have investigated the time course of IgA anti-TG2 normalization, but the limited follow-up in most studies precludes precise conclusions. A decline in IgA anti-TG2 can be observed as early as 2–4 weeks after starting a GFD, and in many patients, titres usually normalize within approximately 12 months [281, 284]. Considerable inter-individual variability exists, however, in the time required to reach normal levels. In one study, 47% of adults with CeD still had nominally positive IgA anti-TG2 titres 1 year after starting a GFD, although titres were significantly lower than at baseline [164].

5.1.3 | Q. When Is a Histological Response to the GFD to Be Expected in Adult CeD Patients?

Statement: Healing of the duodenal mucosa is generally expected around 1 year after starting a GFD. However, the proportion of patients achieving full histological recovery ranges widely, from approximately 50%–83% after 1–5 years.

CoE: low; GR: Strong; Agreement: 100%

Summary of evidence: Histological healing of the duodenal mucosa represents a key milestone in the successful management of CeD, as it underpins restoration of normal intestinal absorption. Whilst Marsh criteria are commonly used in clinical practise to assess the severity of mucosal lesions, they are semi-quantitative and, in research contexts, less precise than measurements of the villous-to-crypt (Vh:Cd) ratio. A Vh:Cd ratio above 2.0–2.5 is generally considered indicative of mucosal recovery [296].

Available studies show considerable variability in the timing and proportion of patients achieving histological healing. In the Risnes study, nearly all patients had recovered a Vh:Cd ratio ≥ 2.0 1 year after starting a GFD [284]. Another study reported that 83% of patients reached a Vh:Cd ratio ≥ 2.0 after an average of 4 years on a GFD [297]. In contrast, Lebwohl et al. observed that only 50% of adults achieved mucosal healing within 6 months–1 year of GFD initiation, increasing to 62% at 1–2 years [265]. Wahab and colleagues reported 65% mucosal recovery after 2 years and 85.3% after 2–5 years, highlighting the existence of a subset of ‘slow responders’ [266]. Data from the US suggest even slower recovery in some cohorts, with only 34% of patients showing full mucosal healing at 2 years and 66% at 5 years after starting a GFD [293].

Overall, these findings underscore both the variability in histological recovery amongst adults with CeD and the importance of long-term follow-up, as a significant proportion of patients may require several years to achieve full mucosal healing.

5.2 | Delayed or Incomplete Response to a GFD

5.2.1 | What Are the Causes of Delayed or Incomplete Response to a GFD in Adults With CeD?

Statement: An incomplete response to a GFD is often due to ongoing gluten exposure but may also indicate slow-responsive disease, RCD, initial misdiagnosis, or concurrent conditions. Persistent symptoms or villous atrophy after ≥ 12 months requires systematic evaluation.

CoE: moderate; GR: strong; Agreement: 100%

5.2.2 | Q. How to Evaluate an Adult Patient With CeD Having Persistent Symptoms Despite GFD?

Recommendation: For a symptomatic adult with CeD despite a GFD, first verify the original diagnosis and assess for gluten exposure. If adherence is confirmed, proceed with follow-up histology and evaluate for alternative or overlapping conditions—such as functional disorders, other GI diseases, refractory CeD, or malignancy—using a multidisciplinary approach.

CoE: moderate; GR: strong; Agreement: 100%

Summary of evidence: Up to 30%–40% of adults with CeD may have persistent symptoms or villous atrophy despite a GFD, although rates vary by definitions, follow-up duration, and clinical setting [265, 298, 299]. The optimal timing for assessing response remains uncertain and is typically based on expert opinion (12–24 months) [2, 265, 300].

Based on a combination of clinical, serological, histopathological, and immunological features, patients who continue to experience symptoms and/or show incomplete mucosal healing despite presumed adherence to a strict GFD are classified into one of the following categories:

1. Continued gluten exposure: Gluten exposure, whether intentional and inadvertent, accounts for up to 80% of persistent symptoms [299, 301, 302]. Intentional gluten intake is reported in up to 40% of adult patients, [303] and it is multifactorial, and may depend on poor self-efficacy, lower symptom burden, scarce knowledge of CeD, social-economic barriers and low educational level [265].
2. Delayed or slow-responsive CeD: Some patients show slow clinical, serological, or histological recovery despite confirmed adherence. Improvement occurs gradually and may take months to years, especially in those with longstanding disease or severe baseline mucosal damage [266].
3. RCD: See Section 5.3 for detailed discussion.

4. Misdiagnosis of CeD. See part 1 of the guidelines, section Q.IV.4. What Is the Approach to Villous Atrophy in the Absence of CeD-Specific Serology? [1].
5. Other conditions unrelated to gluten exposure. these include:
 - Associated or concomitant disorders—other food intolerances (e.g., lactose, fructose), microscopic colitis, inflammatory bowel disease [304], bile acid diarrhoea [305], exocrine pancreatic insufficiency, IBS, medication effects [306, 307], or other autoimmune diseases.
 - Malignant complications—EATL or small-bowel adenocarcinoma [308, 309].
 - Functional or psychosomatic disorders—functional gastrointestinal symptoms, difficulty adapting to a chronic diagnosis, or health-related anxiety, particularly in younger patients.

Structured, stepwise approach to evaluate persistent symptoms despite a GFD includes:

1. *Re-assessment of the original CeD diagnosis* to confirm it was appropriately established before GFD initiation [310, 311].
2. *Exclusion of ongoing gluten exposure* through expert dietary review and, where appropriate, GIPs testing; [91, 312–315] if excluded, alternative gastrointestinal, functional, or psychological causes should be considered [308, 309]. Empirical approaches such as a low-FODMAP diet or pancreatic enzymes may be used when clinically indicated [113, 212].
3. *Assessment of persistent villous atrophy* should be performed with follow-up biopsy, as mucosal healing may be delayed despite symptom resolution, particularly in older adults [265].
4. *Evaluate for and exclude RCD and malignant complications* in patients with persistent villous atrophy persists despite confirmed dietary adherence. This evaluation should be conducted in specialised centres (Figure 1).

5.3 | Refractory Coeliac Disease

5.3.1 | Definition of RCD

5.3.1.1 | Q. What Is the Definition of RCD?. Statement: RCD is defined by the persistence or recurrence of symptoms and villous atrophy after at least 12 months on a strict GFD, in the absence of other causes. RCD can be either primary or secondary. Depending on the proportion of aberrant T cells, RCD is further subdivided into: RCD-I and RCD-II.

CoE: moderate; GR: strong; Agreement: 100%

Summary of evidence: RCD is defined by the persistence or recurrence of symptoms and villous atrophy after at least 12 months on a strict GFD, despite negative coeliac serology. This 12-month duration accounts for the fact that some patients may be slow-responders, extending beyond the 6-month period used in earlier studies [316–318].

From the clinical point of view, RCD can be either primary, when there is no clinical or histological response to a strict GFD from the outset of disease, or secondary, when symptoms and villous atrophy recur after an initial period of response to the GFD, once ongoing gluten exposure has been reliably excluded.

RCD is a rare complication of CeD, affecting approximately 0.3%–2% of adult patients, with most population-based studies suggesting a prevalence closer to 1% or less [319]. RCD-I accounts for the majority of cases, whilst RCD-II represents a minority (around 20%–30% of RCD cases) and is exceedingly rare in the general coeliac population. In referral and tertiary care centres, the reported prevalence of RCD—particularly type II—has been estimated at 1%–4% of all CeD cases; however, this likely reflects referral bias and overestimates true community prevalence [320, 321].

RCD is further subdivided into:

- **RCD-I:** The immunophenotype of IELs is normal. Thusly, it requires the observation of the clinical course of the patient to differentiate RCD-I from slow-responsive CeD. PCR-based amplification of the variable region of the T-cell receptor (TCR) gene in duodenal mucosal biopsies (molecular pathology) reveals a poly- or oligoclonal T-cell population.
- **RCD-II:** Is defined by the presence of an aberrant IEL population characterised by loss of surface CD3 and CD8, with expression of intracytoplasmic CD3. Flow cytometry typically demonstrates $\geq 20\%$ aberrant IELs, and clonality is confirmed by T-cell receptor (TCR) gene rearrangement analysis [322, 323]. RCD-II is considered a pre-lymphomatous state with a high risk of transforming into EATL, accounting for its poor prognosis. Complications such as ulcerative jejunitis and subsequent small bowel stenoses are also common in this subtype [324].

The classification of RCD into types 1 and 2 is based on immunological criteria and may not fully encompass the spectrum of clinical presentations. Incorporating both immunological and clinical criteria into the classification system enhances diagnostic accuracy [322, 325]. The two RCD subtypes represent biologically and prognostically distinct entities. RCD-I is typically characterised by less severe malabsorptive symptoms and relatively preserved nutritional status. In contrast, RCD-II is associated with more severe malabsorption, frequent hypoalbuminaemia, and ulceration or stenosis of the small bowel. These differences underpin the markedly divergent clinical outcomes, with RCD-I generally following a benign course with excellent long-term survival, whereas RCD-II carries a high risk of progression to EATL and a poor prognosis.

Notably, a subset of patients presents with clonal or aberrant IELs in the absence of significant clinical symptoms, biochemical abnormalities, or histological damage, raising uncertainty about their classification as RCD-II. These cases are challenging to categorize and may not warrant immediate aggressive therapy; instead, they require careful monitoring. Importantly, the clonal T cell population in such individuals may remain stable over time or even regress during follow-up. Recognizing this intermediate subset has meaningful clinical implications,

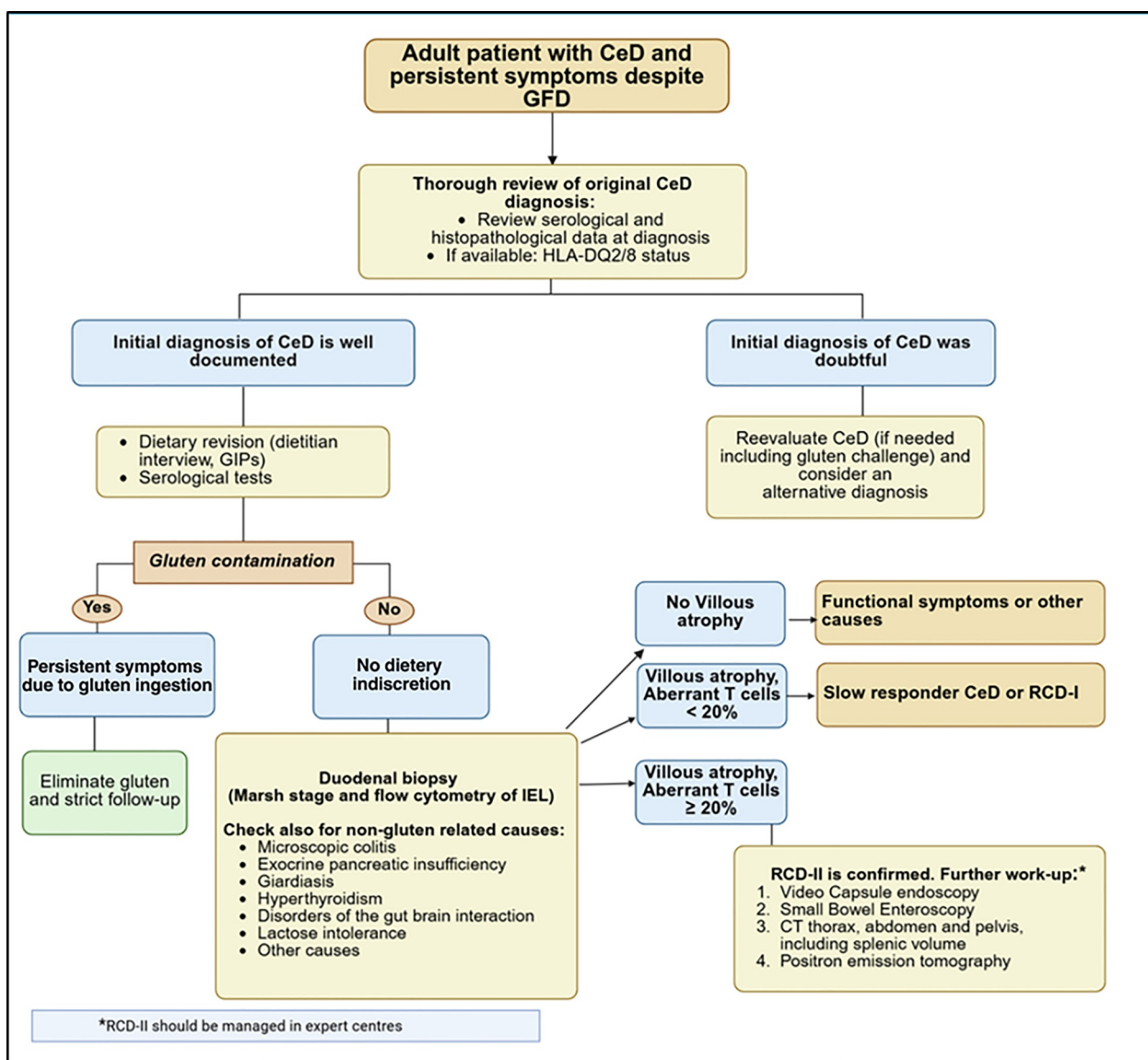


FIGURE 1 | Summarises the approach to adult patients with CeD having persistent symptoms despite GFD. CeD, Coeliac disease; CT scan, Computed tomography; GIPs, Gluten Immunogenic Peptides; GFD, Gluten-Free Diet; HLA-DQ2/8, Human leucocyte antigen; IEL, Intraepithelial lymphocytes; RCD, Refractory Coeliac Disease, RCD-I, refractory coeliac disease type 1; RCD-II, refractory coeliac disease type 2.

potentially preventing overtreatment and supporting more individualized management strategies. Further research is essential to validate these observations and refine existing classification frameworks [322, 326].

5.3.2 | Diagnosis of RCD

5.3.2.1 | Q. What Is the Diagnostic Approach to a Patient With Suspected RCD?. *Recommendation:* The diagnostic approach involves a systematic workup to confirm the initial CeD diagnosis, ensure strict dietary adherence, and exclude alternative causes of symptoms or an overt EATL. Essential investigations include serology, duodenal biopsies (for histology, IEL flow cytometry, and T-cell receptor clonality analysis), enteroscopy, and cross-sectional imaging.

If a strong clinical suspicion for RCD persists, further evaluation and management should be conducted in or in consultation with a tertiary centre experienced in managing RCD.

CoE: moderate; GR: strong; Agreement: 100%

5.3.2.2 | Q. Which Molecular Diagnostic Technology Is the Gold Standard for Diagnosis of RCD-II?. *Recommendation:* Accurate subtyping of RCD requires comprehensive immunophenotyping of small bowel lymphocytes to reliably diagnose or exclude RCD-II. Flow cytometric analysis of isolated IELs after immunostaining is superior to immunohistochemistry for detecting aberrant lymphocyte populations. Additional diagnostic value is provided by PCR-based T-cell receptor clonality assessment and by sequencing of genes in the JAK/STAT pathway to identify relevant somatic mutations.

CoE: moderate; GR: strong; Agreement: 100%

Summary of evidence: The diagnostic workup of RCD is summarised in Table 7:

5.3.3 | Treatment of RCD

5.3.3.1 | Q. Is There an Evidence-Based Treatment for RCD-I and RCD-II? *Recommendation:* No evidence-based medical treatments supported by controlled trials exist for either RCD-I or RCD-II; current management is based on expert consensus, case series, and observational data.

CoE: low; GR: strong; Agreement: 100%

5.3.3.2 | Q. What Is the Treatment for Patients With RCD-II? *Recommendation:* Based on retrospective and longitudinal evidence, open-capsule budesonide is considered first-line therapy for RCD-I. Conventional immunosuppressants, such as thiopurines, may be added in selected cases. When used, thiopurines should be re-evaluated for possible discontinuation after 2–3 years of clinical, histological and immunophenotypic stability.

CoE: low; GR: strong; Agreement: 100%

5.3.3.3 | Q. What Is the Treatment for Patients With RCD-II? *Recommendation:* Mild-to-moderate RCD-II may be treated with open-capsule budesonide. In selected patients, cladribine (or fludarabine), with or without autologous haematopoietic stem cell transplantation (auto-HSCT), or JAK inhibitors may be considered on an individual basis.

CoE: low; GR: strong; Agreement: 100%

Summary of evidence: Prospectively designed, randomized controlled trials (RCTs) in RCD are exceedingly scarce. To date, the only prospective controlled study evaluated anti-IL-15 therapy (AMG 714) and did not meet its primary endpoint, which was a reduction in aberrant IELs. However, secondary endpoints—such as clinical improvement in chronic diarrhoea and a reduction in T-cell clonality—were achieved [350].

All other available studies are retrospective and typically longitudinal in design. Their conclusions rely on interpreting clinical, histological, and immunological changes after initiating a medical intervention. Because these studies lack appropriate control groups (e.g., placebo or equivalent comparator), it is not possible to determine with certainty whether the observed outcomes are attributable to the intervention itself [342, 346, 356–362].

Treatment of RCD-I:

In an open-label study including 43 patients with RCD-I, budesonide administered as an open-capsule, induced clinical response and histologic improvement in approximately 90% of patients with RCD-I [356]. This treatment is based on the hypothesis that opening the gelatine capsule followed by grinding the drug will allow the release of budesonide from polymer

matrix, providing more immediate action in the proximal small-bowel. Another retrospective study concluded that open-capsule budesonide was well-tolerated and associated with improvements in enteropathy (83%) and symptoms (90%) in CeD patients with persistent symptoms and RCD [357].

Escalation therapy to immunosuppressants is not standardized and is also not supported by firm data from clinical trials. Nevertheless, it can be considered in cases of steroid refractoriness or dependency on higher steroid doses. In RCD-I, the addition of thiopurines, such as, azathioprine or 6-mercaptopurine to steroids was effective in inducing a clinical response in some cases [316, 360, 363]. When used, thiopurines should be re-evaluated for possible discontinuation after 2–3 years of clinical, histological and immunophenotypic stability. The options for pharmacological treatment of RCD-I are summarized in Table 8.

Treatment of RCD-II:

Summary of evidence: Several therapies have been attempted for RCD-II, including cladribine (2-chlorodeoxyadenosine (2-CdA) [358, 360], open-capsule budesonide [356], 6-thioguanine [364], alemtuzumab [316], infliximab [365, 366], auto-HSCT [330, 342, 346, 359, 367, 368], and mesenchymal stem cell infusion [369]. Whilst some treatments showed positive results, there is still a risk of progression to EATL [360, 368, 370].

In a study, including 13 RCD-II, budesonide administered as an open-capsule, induced clinical response and histologic improvement in 77% and 55% of cases, respectively [356]. Moreover, in 7 patients, the clonal TCR rearrangement observed at diagnosis was not detected in the follow-up biopsies and the number of abnormal IELs were reduced.

The use of azathioprine in patients with RCD-II is not currently recommended due to the potential risk of increasing the likelihood of developing EATL [318, 361]. Treatment with biologics, such as infliximab, may induce responses, but only a few cases have been reported [371].

In an open-label study, including 13 patients with RCD-II unresponsive to cladribine and treated with auto-HSCT, most showed clinical improvement, with a 4-year survival rate of 66% [346, 359].

The anti-IL-15 monoclonal antibody AMG714 was tested in a phase-2 trial but did not meet the primary endpoint of reducing aberrant IELs [350]. However, it did improve diarrhoea and reduce T-cell receptor clonality.

Due to the presence of JAK/STAT pathway mutations in RCD-II, tofacitinib, a JAK 1/3 inhibitor, may offer promising therapeutic potential [372]. In a small study, tofacitinib induced clinical and histological improvement in six patients with RCD-II but did not reduce abnormal IELs [362]. Further research is needed to confirm the safety and effectiveness of this approach, with the goal of reducing inflammation and limiting the risk of EATL [361]. The options for pharmacological treatment of RCD-II are summarized in Table 8.

TABLE 7 | The diagnostic workup of refractory coeliac disease.

Domain	Key points
1. Clinical assessment	Common manifestations: Chronic diarrhoea, weight loss, abdominal pain, and malnutrition. RCD typically presents in adults, often after a long-standing CeD diagnosis, and may include systemic features such as fever or signs of protein-losing enteropathy [323, 327]. In severe cases with widespread bowel involvement, a 'functional' short bowel syndrome may occur, complicated by D-lactic acidosis in the context of carbohydrate malabsorption or bacterial overgrowth [328, 329]. Clinical suspicion should also be raised in newly diagnosed CeD with severe symptoms and extensive villous atrophy. Rarely, extra-intestinal localisation of aberrant T cells (skin, sinuses, lungs, liver, CNS) may cause atypical presentations [330–332].
2. Endoscopy and histology	Endoscopic features may resemble active CeD; adequate duodenal biopsies must be collected in formalin and for IEL immunophenotyping. Exclude collagenous sprue, characterised by thickened subepithelial collagen bands [333–335]. Mucosal ulcers raise suspicion for RCD-II. VCE is useful for assessing extent of mucosal damage and guiding device-assisted enteroscopy to rule out EATL [333–338].
3. Immunophenotyping of IELs	Essential to differentiate RCD-I from RCD-II. RCD-I shows normal cytotoxic T-lymphocytes phenotype (CD103+, sCD3+, CD8+). RCD-II shows aberrant IELs with intracellular CD3 but lacking surface CD3 and CD8 [339]. These aberrant T cells represent a clonal, pre-malignant population with potential progression to EATL. Flow cytometry is regarded as the most sensitive and specific technique for detecting these aberrant IEL populations, as it allows precise differentiation between cytoplasmic and membranous CD3 expression and quantifies the proportion of aberrant cells [340]. It is also valuable for excluding other indolent small-bowel lymphoproliferative disorders [322, 323, 341]. However, the availability of flow cytometry with expertise in IEL phenotyping remains limited to specialised centres, and sample transport and tissue handling requirements further constrain its routine use. Immunohistochemistry (IHC), in contrast, is more widely available and can provide reliable information when performed by experienced laboratories. IHC allows assessment of CD3 and CD8 expression patterns and, when combined with TCR gene rearrangement studies, can support the diagnosis in most cases. Although less quantitative than flow cytometry, IHC remains a practical first-line approach for many institutions, reserving flow cytometry for cases where IHC findings are inconclusive or where RCD-II is strongly suspected [322]. Overall, both techniques contribute to accurate classification of RCD subtypes. Flow cytometry remains the reference standard for identifying aberrant IELs, but immunohistochemistry—with or without TCR clonality testing—offers a feasible and informative alternative in settings where flow cytometry is unavailable.
4. T-cell clonality analysis	Assesses TCR rearrangements. RCD-II shows monoclonal/oligoclonal populations; RCD-I remains polyclonal [327, 339, 342]. If TCR- γ is absent, TCR- δ may be detected [316, 343]. TCR analysis allows diagnosis of extra-intestinal locations of RCD-II such as skin, lung, brain or in peripheral blood by identification of the same TCR rearrangement found in duodenum [344–346].
5. Genetic analysis	Screening for germline mutations and constitutive (primary) immune disorders aids in differentiating RCD-I from autoimmune enteropathy [347].
6. Somatic mutation analysis	Somatic mutations may guide therapy. JAK1–STAT3 gain-of-function mutations found in up to 80% of RCD-II cases and 90% of EATL cases [348–350]. Other mutations (TNFAIP3/A20, TET2, KMT2D) may explain poor response to anti-IL-15 therapy [348, 350, 351].
7. Radiological imaging	Findings may include marked small-bowel wall thickening, strictures, ulceration or ulcerative jejunitis, mass-like changes, and atypical or necrotic mesenteric lymph nodes. Reduced spleen volume may also be present. PET can assist in excluding overt EATL and detecting metabolically active complications [183, 352–354].
8. Nutritional assessment	Wasting and hypoalbuminaemia are suggestive of RCD-II or EATL [322, 355].
9. Exclusion of EATL	All diagnostic modalities should be used to exclude EATL before confirming RCD.

Abbreviations: CeD, coeliac disease; CNS, central nervous system; EATL, enteropathy-associated T-cell lymphoma; IEL, intraepithelial lymphocytes; IHC, immunohistochemistry; PET, positron emission tomography; RCD, refractory coeliac disease; sCD3, surface CD3; TCR, T-cell receptor (TCR- γ , gamma, TCR- δ delta); VCE, video capsule endoscopy.

TABLE 8 | Options for pharmacological treatment in RCD-I and RCD-II.^a

RCD-I	
Nutritional support	Enteral or parenteral nutrition, as needed.
Open-capsule budesonide	9 mg daily for 6 weeks, followed by 6 mg daily for an additional 6 weeks, then 3 mg daily thereafter. 9 mg dose: open two budesonide capsules and grind the contents; swallow with plenty of water. Swallow the third capsule whole. 6 and 3 mg doses: Open the capsule(s) and grind the contents; swallow with plenty of water.
Oral prednisone	0.5–1 mg/kg body weight may be needed for few weeks in cases of no response to budesonide. Following a response, thiopurines, such as azathioprine or 6-thioguanine may be initiated and continued for up to 2 years. Careful monitoring is required to detect potential complications.
Thiopurines, such as azathioprine	Azathioprine 2.5 mg/kg body weight to maintain remission after tapering of steroids. Re-evaluated for possible discontinuation after 2–3 years of clinical histological and immunophenotypic stability.
RCD-II	
Nutritional support	Enteral or parenteral nutrition, as needed.
Open capsule budesonide	Dosage and capsule handling are described in the RCD-I section. Seriously ill patients might require alternative treatment including parenteral steroids.
Cladribine (2-CdA)	A dose of 0.15 mg/kg/day administered intravenously for 5 consecutive days, repeated in 1–3 treatment courses at intervals of 6 months.
Autologous haematopoietic stem cell transplantation (Auto-HSCT)	A conditioning regimen with fludarabine and melphalan followed by Auto-HSCT.
JAK inhibitors, such as, tofacitinib	10 mg twice daily for 12 weeks, followed by a reduced maintenance dose of 5 mg twice daily.

Abbreviations: Auto-HSCT, autologous haematopoietic stem cell transplantation; 2-CdA, 2-chlorodeoxyadenosine; EATL, enteropathy-associated T-cell lymphoma; JAK, Janus kinase; RCD, refractory coeliac disease; RCD-I, refractory coeliac disease type 1; RCD-II, refractory coeliac disease type 2.

^aImportant notes: (1) Dosages are indicative and should be individualised based on clinical response, tolerability, and comorbidities. (2) Patients with RCD-II should be managed in specialised referral centres with expertise in complicated coeliac disease, including dedicated gastroenterology, immunology, and haematology services. Whenever possible, treatment within clinical trials is strongly recommended. (3) Enteropathy-associated T-cell lymphoma (EATL) must be confidently excluded before initiating immunosuppressive or cytotoxic therapy. (4) In a subset of patients with RCD-II complicated by ulcerative jejunitis and significant small-bowel stenosis, surgical resection prior to pharmacological therapy may improve survival [324].

5.3.4 | Prognosis of RCD

Summary of evidence: The prognosis of RCD is influenced by multiple factors, including disease subtype, nutritional status, and development of complications such as ulcerative jejunitis, EATL, and small bowel adenocarcinoma [308]. The most important prognostic determinant is the distinction between RCD-I and RCD-II. RCD-I generally follows a benign course with a 5-year survival rate exceeding 90%, whereas RCD-II carries a significantly higher risk of progression to EATL and has a 5-year survival of approximately 44%–58% [342, 373].

Other poor prognostic indicators in RCD include severe malabsorption, weight loss, hypoalbuminemia, and persistent inflammation despite treatment [355, 373]. Nutritional deficits in patients with EATL are usually profound, driven by severe malabsorption and systemic disease burden, and are associated with poorer tolerance to chemotherapy and worse overall survival rates [342, 373, 374].

Additionally, patients with RCD who develop ulcerative jejunitis, strictures, or infections have a worse prognosis [342].

6 | Coeliac Disease and Malignancy: Associations and Risk of Malignant Transformation

6.1 | Q. What Is the Risk of Malignant Complications in Coeliac Disease (Non-refractory CeD)?

Statement: The risk of malignant complications in non-refractory CeD is only slightly increased, remains low in absolute terms, and decreases with long-term adherence to a strict GFD. Routine malignancy screening beyond general population recommendations is not indicated in the absence of clinical suspicion.

CoE: low; GR: strong; Agreement: 95%

6.2 | Q. What Is the Risk of Malignant Transformation in RCD-II?

Statement: RCD-II is associated with a substantial risk of malignant transformation, most commonly progression to EATL,

which develops in up to half of affected patients and is the major determinant of the markedly reduced 5-year survival seen in this subtype.

CoE: low; GR: strong; Agreement: 100%

Summary of Evidence: Patients with CeD who are responsive to GFD and do not develop refractory disease have a modestly increased risk of certain malignancies compared with the general population; however, the absolute risk remains low [308]. Large population-based studies demonstrate a small increase in overall cancer risk, particularly during the first years following diagnosis, likely reflecting prolonged untreated disease prior to diagnosis rather than the effect of treated CeD [375, 376].

Specific malignancies reported at increased relative risk include lymphoproliferative malignancies, small bowel adenocarcinoma, and oesophageal cancer, but these remain rare events in absolute terms [308]. Importantly, the risk of malignancy appears to decline over time with sustained adherence to a strict GFD and may approach that of the general population during long-term follow-up. Development of EATL in patients without RCD is exceedingly uncommon. Accordingly, in patients with non-refractory CeD who are clinically stable and adherent to a GFD, routine cancer surveillance beyond age- and risk-appropriate population screening is not recommended. Long-term follow-up should focus on assessment of dietary adherence, symptom review, nutritional status, and vigilance for alarm features.

In contrast, RCD—particularly RCD-II—carries a substantially increased risk of malignant transformation, most notably progression to EATL. RCD-II is widely regarded as a pre-lymphomatous condition [342]. The cumulative risk of EATL in RCD-II is high, and malignant transformation may occur despite dietary adherence. The risk of malignant transformation in RCD-I is considerably lower, though still higher than in uncomplicated CeD.

Patients with RCD should therefore undergo close clinical follow-up, including regular assessment for symptoms suggestive of lymphoma (e.g., persistent abdominal pain, unexplained weight loss, fevers, bowel obstruction), and appropriate use of imaging, endoscopy, and histological reassessment where indicated.

In patients who progress to EATL, prognosis is particularly poor. EATL often presents late, with nonspecific symptoms or complications such as bowel perforation, bleeding, or obstruction [374, 377]. Median survival after diagnosis is less than 1 year, even with treatment. The survival in EATL is negatively affected by: profound malnutrition and catabolic state, poor performance status at diagnosis and delayed or incomplete response to therapy [342].

Two types of EATL are recognised. Type I EATL accounts for approximately 80%–90% of cases and is exclusively associated with pre-existing CeD, often following RCD-II. Type II EATL (de novo) is usually not associated with pre-existing CeD and may occur independently of CeD [378].

Management of EATL typically involves multi-agent chemotherapy, but outcomes are limited by chemotherapy toxicity, particularly in patients with malnutrition. Recent protocols incorporate pre-emptive debulking surgery with resection of the affected bowel segment, and auto-HSCT as consolidation therapy in selected fit patients, which may improve survival [379]. Combination chemotherapy followed by auto-HSCT has shown improved progression-free survival in highly selected patients, with 2-year survival rates up to 60%–70% in some series. However, many patients are ineligible for intensive regimens due to poor functional status or advanced disease at presentation.

Early identification of RCD-II and close surveillance for transformation to EATL is critical. A multidisciplinary approach involving gastroenterologists, dietitians, haematologists/oncologists, and pathologists is essential to optimize diagnosis, supportive care, and treatment planning [380]. Advances in targeted therapies offer hope for improving survival rates and QoL in affected individuals.

Small bowel adenocarcinoma, although less common than EATL, is a recognized and serious complication that also confers a poor prognosis [308]. Small bowel adenocarcinoma often presents with obstructive symptoms, bleeding, or perforation, typically in the jejunum. Diagnosis is usually made via capsule endoscopy, CT/MRI enterography, or enteroscopy with biopsy. Treatment involves surgical resection, which is the only curative option [381–383].

7 | Conclusions, Limitations of These Guidelines and Future Perspectives

These 2025 ESSCD guidelines, presented in two parts, update the 2019 recommendations on the diagnosis and management of CeD in adults [2], incorporating new evidence from multidisciplinary expert consensus using the AGREE II and GRADE methodologies. Part 1, which has been already published, refines diagnostic strategies, including serological testing, biopsy protocols, and the no-biopsy approach for high-titre IgA anti-TG2 cases [1].

The current Part 2 of the ESSCD guidelines provides a comprehensive, evidence-based framework for the management and follow-up of adult CeD. This update transforms the standard of care from a sole emphasis on dietary exclusion to a more sophisticated, personalised, proactive, and multidisciplinary paradigm. Central to these advancements are refined strategies for complex disease courses such as RCD, a strengthened focus on the metabolic health and nutritional quality of the GFD, and the formal integration of innovative care models leveraging telemedicine and dietitian-led services.

The guidelines specifically address dietary management—including the GFD, safe consumption of oats, and the use of a low-FODMAP diet for persistent symptoms—alongside the management of associated and documented exocrine pancreatic insufficiency. Furthermore, it introduces critical new topics: expanded strategies for nutritional and psychosocial monitoring, metabolic risk management, the use of digital

communication tools, and updated recommendations for family screening and pregnancy care. It also details systematic approaches to patients with persistent symptoms despite GFD and RCD, clarifying the RCD-I/II subtypes and the associated risk of EATL.

Despite the significant progress in the management of adults with CeD, several challenges and knowledge gaps remain. The certainty of evidence supporting guideline recommendations is limited, largely because many clinical situations have not been studied prospectively or in controlled settings. As a result, we often lack the opportunity to propose algorithms supported by robust evidence. Key limitations include the absence of a unified framework for long-term follow-up and the lack of standardized approaches for managing patients with persistent symptoms. Consensus is also lacking on optimal strategies for monitoring and maintaining bone mineral health specifically in CeD. There is a clear need for high-quality data on the indications, timing, and frequency of DXA testing in adults with CeD, rather than relying on extrapolation from other disease populations.

In addition, whilst non-dietary pharmacological therapies represent a promising and much-needed avenue for treatment, their efficacy and safety require validation through rigorously designed clinical trials. Addressing these gaps will be critical to improving evidence-based, individualized care and long-term outcomes for adults with CeD.

Furthermore, the practical implementation of these recommendations faces hurdles, especially in low-resource settings where equitable access to specialised diagnostics, dietetic expertise, the local or national policies and novel treatments may be constrained.

Looking ahead, future efforts must focus on bridging these gaps through high-quality basic science and robust clinical trials. A critical priority is the development of pragmatic, context-sensitive guidance that can be adapted to diverse healthcare systems, ensuring that all patients with CeD, regardless of location, can benefit from advancing standards of care. Wide-spread dissemination and flexible application of these guidelines are essential to elevate the QoL and long-term health outcomes for adults living with this condition globally.

Author Contributions

The ESsCD board (A.A., A.P., L.E., C.G., N.T., R.A., K.L., L.M.S., M.S.) organised the working groups and designed the preliminary list of topics to be covered. C.C., I.R., and H.O. conducted the assessment of the evidence and applied the GRADE approach. All authors (A.A., F.Z., G.M., A.S., N.T., F.B., L.E., A.P., C.G., R.A., A.B., D.S., C.S., C.M., G.B., K.L., L.M.S., M.S.) systematically reviewed the literature and draughted the statements and recommendations and provided GRADE evaluations. All authors and members of the guidelines working group voted on the statements and recommendations. The subgroups then draughted the initial manuscript, which was reviewed, revised and approved by all members of the guidelines working group. Subsequently, it was made available to all members for final comments prior to submission for publication.

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Conflicts of Interest

Before appointment to the panel, individuals disclosed financial and nonfinancial interests. No industry or government affiliations influenced this guideline. *Fabiana Zingone* has received speaker fees from Werfen, EG Stada Group, Fresenius Kabi, Kedrion, Janssen, Pfizer, Takeda, Unifarco, Malesci, and Galapagos; and has consulted for Galapagos, Takeda, and Tillotts. *Ludvig M. Sollid* has served as a consultant in the past three years for Falk, GSK, Precigen ActoBio, Sanofi, Takeda, and Topas Therapeutics. *Knut Lundin* has had confidentiality agreements, consultancy roles, or speaker honorariums with Allero, Alimentiv, Anokion, Amyra, Chugai, GenXBioscience, Falk, Takeda, Topas, and Tillotts. *David S. Sanders* has received an educational grant from Dr Schaer, serves as a board member of Nemysis, and has received consulting fees from Tillotts and Takeda. *Michael Schumann* has had confidentiality agreements, consultancy roles, or speaker honorariums with Falk, Takeda, Topas, Dr. Schär and Tillotts. All other authors declared no conflict of interest.

Disclaimer

These guidelines have been developed with reasonable care and with the best of knowledge available to the authors at the time of preparation. They are intended to assist healthcare professionals and allied healthcare professionals as an educational tool to provide information that may support them in providing care to patients. Patients or other community members using these guidelines shall do so only after consultation with a health professional and shall not mistake these guidelines as professional medical advice. These guidelines must not substitute seeking professional medical and health advice from a health professional. These guidelines may not apply to all situations and should be interpreted in the light of specific clinical situations and resource availability.

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Data Availability Statement

The data that supports the findings of this study are available in the supplementary material of this article.

References

1. A. Al-Toma, F. Zingone, F. Branchi, et al., "European Society for the Study of Coeliac Disease 2025 Updated Guidelines on the Diagnosis and

- Management of Coeliac Disease in Adults. Part 1: Diagnostic Approach," *United European Gastroenterology Journal* 13, no. 10 (2025): 1–32, <https://doi.org/10.1002/UEG2.70119>.
2. A. Al-Toma, U. Volta, R. Auricchio, et al., "European Society for the Study of Coeliac Disease (ESsCD) Guideline for Coeliac Disease and Other Gluten-Related Disorders," *United European Gastroenterology Journal* 7, no. 5 (2019): 583–613, <https://doi.org/10.1177/2050640619844125>.
3. J. A. Murray, T. Watson, B. Clearman, and F. Mitros, "Effect of a Gluten-Free Diet on Gastrointestinal Symptoms in Celiac Disease," *American Journal of Clinical Nutrition* 79, no. 4 (2004): 669–673, <https://doi.org/10.1093/ajcn/79.4.669>.
4. H. Pekki, K. Kaukinen, T. Ilus, et al., "Long-Term Follow-Up in Adults With Coeliac Disease: Predictors and Effect on Health Outcomes," *Digestive and Liver Disease* 50, no. 11 (2018): 1189–1194, <https://doi.org/10.1016/J.DLD.2018.05.015>.
5. D. V. Balaban, I. Enache, M. Balaban, et al., "Outcomes in Adults With Celiac Disease Following a Gluten-Free Diet," *Journal of Clinical Medicine* 14, no. 14 (2025): 5144, <https://doi.org/10.3390/JCM14145144>.
6. C. Dochat, N. Afari, R. M. Satherley, S. Coburn, and J. F. McBeth, "Celiac Disease Symptom Profiles and Their Relationship to Gluten-Free Diet Adherence, Mental Health, and Quality of Life," *BMC Gastroenterology* 24, no. 1 (2024): 9, <https://doi.org/10.1186/S12876-023-03101-X>.
7. S. La Vieille, O. M. Pulido, M. Abbott, T. B. Koerner, and S. Godefroy, "Celiac Disease and Gluten-Free Oats: A Canadian Position Based on a Literature Review," *Chinese Journal of Gastroenterology and Hepatology* 2016 (2016): 1–10, <https://doi.org/10.1155/2016/1870305>.
8. R. Enaud, C. Tetard, R. Dupuis, et al., "Compliance With Gluten Free Diet Is Associated With Better Quality of Life in Celiac Disease," *Nutrients* 14, no. 6 (2022): 1210, <https://doi.org/10.3390/NU14061210>.
9. N. Trott, W. Holland, O. Hoffmann, et al., "Food Related Quality of Life and Associations With Demographic and Clinical Characteristics in People With Coeliac Disease," *Journal of Human Nutrition and Dietetics* 38, no. 3 (2025): e70051, <https://doi.org/10.1111/JHN.70051>.
10. A. Kowalczyk and F. Moor, "A Meta-Synthesis Exploring Daily Experiences of Adults With Coeliac Disease in Adhering to a Gluten-Free Diet," *Journal of Human Nutrition and Dietetics* 38, no. 2 (2025): e70043, <https://doi.org/10.1111/JHN.70043>.
11. D. Studerus, E. I. L. G. Hampe, D. Fahrner, M. Wilhelmi, and S. R. Vavricka, "Cross-Contamination With Gluten by Using Kitchen Utensils: Fact or Fiction?," *Journal of Food Protection* 81, no. 10 (2018): 1679–1684, <https://doi.org/10.4315/0362-028X.JFP-17-383>.
12. Accessing Gluten Free Food in Hospital - Coeliac UK. Accessed April 9, 2023, https://www.coeliac.org.uk/information-and-support/living-gluten-free/the-gluten-free-diet/hospital-visits/?utm_source=chatgpt.com&&type=rfst&set=true#cookie-widget.
13. C. Catassi, E. Fabiani, G. Iacono, et al., "A Prospective, Double-Blind, Placebo-Controlled Trial to Establish a Safe Gluten Threshold for Patients With Celiac Disease," *American Journal of Clinical Nutrition* 85, no. 1 (2007): 160–166, <https://doi.org/10.1093/ajcn/85.1.160>.
14. Implementing Regulation - 828/2014 - EN - EUR-Lex. Accessed October 25, 2024, https://eur-lex.europa.eu/eli/reg_impl/2014/828/oj.
15. Ad Hoc Joint FAO/WHO Expert Consultation on Risk Assessment of Food Allergens – Reference Dose(S) for Cereals Containing Gluten or Gluten. Summary and Conclusions. Accessed December 11, 2025, <https://openknowledge.fao.org/items/0a455421-491b-4c44-a786-83c51452bdef>.
16. I. Lizano-Díez, E. L. Mariño, and P. Modamio, "Gluten in Pharmaceutical Products: A Scoping Review," *Systematic Reviews* 10, no. 1 (2021): 218, <https://doi.org/10.1186/S13643-021-01772-9>.
17. A. V. Shah, A. T. M. M. Serajuddin, and R. A. Mangione, "Making all Medications Gluten Free," *Journal of Pharmaceutical Sciences* 107, no. 5 (2018): 1263–1268, <https://doi.org/10.1016/j.xphs.2017.12.021>.
18. V. Phan, J. Aurora, S. Gondi, and L. Ceglie, "An Overlooked Medication-Induced Celiac Flare Complicating Treatment of Osteoporosis," *AACE Clin Case Reports* 10, no. 2 (2024): 60–62, <https://doi.org/10.1016/J.AACE.2024.01.001>.
19. Annex of the European Commission guideline "Excipients in the Labelling and Package Leaflet of Medicinal Products for Human Use": (EMA/CHMP/302620/2017). Accessed, October 25, 2024, https://www.ema.europa.eu/en/documents/scientific-guideline/questions-and-answers-wheat-starch-containing-gluten-used-excipient-medicinal-products-human-use_en.pdf.
20. Gluten Threshold Study - Wesley Research Institute. Accessed November 2, 2025, <https://www.wesleyresearch.org.au/clinical-trial/gluten-threshold-study/>.
21. J. Stamnaes, D. Stray, M. Stensland, et al., "Well-Treated Celiac Patients Low-Level Mucosal Inflammation Predicts Response to 14-Day Gluten Challenge," *Advanced Science* 8, no. 4 (2021): 2003526, <https://doi.org/10.1002/ADVS.202003526>.
22. M. Brottveit, A.-C. R. Beitnes, S. Tollefsen, et al., "Mucosal Cytokine Response After Short-Term Gluten Challenge in Celiac Disease and Non-Celiac Gluten Sensitivity," *American Journal of Gastroenterology* 108, no. 5 (2013): 842–850, <https://doi.org/10.1038/ajg.2013.91>.
23. N. Patel, D. A. Leffler, A. Al-Toma, et al., "Clinical Data Do Not Reliably Predict Duodenal Histology at Follow-up in Celiac Disease: A 13 Center Correlative Study," *American Journal of Surgical Pathology* 48, no. 2 (2024): 212–220, <https://doi.org/10.1097/PAS.0000000000002150>.
24. K. Aaltonen, P. Laurikka, H. Huhtala, M. Mäki, K. Kaukinen, and K. Kurppa, "The Long-Term Consumption of Oats in Celiac Disease Patients Is Safe: A Large Cross-Sectional Study," *Nutrients* 9, no. 6 (2017): 611, <https://doi.org/10.3390/NU9060611>.
25. A. Alakoski, K. Hervonen, E. Mansikka, et al., "The Long-Term Safety and Quality of Life Effects of Oats in Dermatitis Herpetiformis," *Nutrients* 12, no. 4 (2020): 1060, <https://doi.org/10.3390/NU12041060>.
26. A. R. Lee, M. Dennis, J. Lebovits, et al., "Dietary Assessments in Individuals Living With Coeliac Disease: Key Considerations," *Journal of Human Nutrition and Dietetics* 38, no. 1 (2025): e13380, <https://doi.org/10.1111/JHN.13380>.
27. M. Silano, R. Di Benedetto, F. Maiale, et al., "Avenins From Different Cultivars of Oats Elicit Response by Coeliac Peripheral Lymphocytes," *Scandinavian Journal of Gastroenterology* 42, no. 11 (2007): 1302–1305, <https://doi.org/10.1080/00365520701420750>.
28. G. Tanner, A. Juhász, C. G. Florides, et al., "Preparation and Characterization of Avenin-Enriched Oat Protein by Chill Precipitation for Feeding Trials in Celiac Disease," *Frontiers in Nutrition* 6 (2019): 162, <https://doi.org/10.3389/FNUT.2019.00162>.
29. M. I. Pinto-Sánchez, N. Causada-Calo, P. Bercik, et al., "Safety of Adding Oats to a Gluten-Free Diet for Patients With Celiac Disease: Systematic Review and Meta-Analysis of Clinical and Observational Studies," *Gastroenterology* 153, no. 2 (2017): 395–409.e3, <https://doi.org/10.1053/j.gastro.2017.04.009>.
30. J. A. See, K. Kaukinen, G. K. Makharia, P. R. Gibson, and J. A. Murray, "Practical Insights Into Gluten-Free Diets," *Nature Reviews Gastroenterology & Hepatology* 12, no. 10 (2015): 580–591, <https://doi.org/10.1038/NRGASTRO.2015.156>.
31. A. R. Lee, "Review Article: Dietary Management of Coeliac Disease," supplement, *Alimentary Pharmacology & Therapeutics* 56, no. S1 (2022): S38–S48, <https://doi.org/10.1111/APT.16974>.
32. F. Thies, L. F. Masson, P. Boffetta, and P. Kris-Etherton, "Oats and Bowel Disease: A Systematic Literature Review," supplement, *British*

- Journal of Nutrition* 112, no. Suppl 2 (2014): S31–S43, <https://doi.org/10.1017/S0007114514002293>.
33. L. Gilissen, I. Van der Meer, and M. Smulders, “Why Oats Are Safe and Healthy for Celiac Disease Patients,” *Medical Science* 4, no. 4 (2016): 21, <https://doi.org/10.3390/MEDSCI4040021>.
 34. I. S. Cohen, A. S. Day, and R. Shaoul, “Gluten in Celiac Disease—More or Less?,” *Rambam Maimonides Medical Journal* 10, no. 1 (2019), <https://doi.org/10.5041/RMMJ.10360>.
 35. J. M. Rodríguez, V. Estévez, K. Bascuñán, J. Ayala, and M. Araya, “Commercial Oats in Gluten-Free Diet: A Persistent Risk for Celiac Patients,” *Frontiers in Nutrition* 9 (2022): 986282, <https://doi.org/10.3389/FNUT.2022.986282>.
 36. D. Wild, G. G. Robins, V. J. Burley, and P. D. Howdle, “Evidence of High Sugar Intake, and Low Fibre and Mineral Intake, in the Gluten-Free Diet,” *Alimentary Pharmacology & Therapeutics* 32, no. 4 (2010): 573–581, <https://doi.org/10.1111/J.1365-2036.2010.04386.X>.
 37. O. Vincentini, M. Izzo, F. Maiale, E. Gonnelli, S. Neuhold, and M. Silano, “Risk of Cross-Contact for Gluten-Free Pizzas in Shared-Production Restaurants in Relation to Oven Cooking Procedures,” *Journal of Food Protection* 79, no. 9 (2016): 1642–1646, <https://doi.org/10.4315/0362-028X.JFP-15-538>.
 38. B. D. McDonald and S. S. Kupfer, “Can We Cross off Common Kitchen Practices as Causes of Gluten Cross-Contact?,” *Gastroenterology* 158, no. 1 (2020): 51–53, <https://doi.org/10.1053/J.GASTRO.2019.11.011>.
 39. N. Korth, S. L. Taylor, J. L. Clarke, and M. L. Downs, “Gluten Cross-Contact in Restaurant-Scale Pasta Cooking,” *Journal of Food Protection* 84, no. 12 (2021): 2159–2162, <https://doi.org/10.4315/JFP-21-230>.
 40. AOECs - Association of European Coeliac Societies. Accessed, October 25, 2024, <https://www.aoecs.org/>.
 41. R. L. Wolf, B. Lebwohl, A. R. Lee, et al., “Hypervigilance to a Gluten-Free Diet and Decreased Quality of Life in Teenagers and Adults With Celiac Disease,” *Digestive Diseases and Sciences* 63, no. 6 (2018): 1438–1448, <https://doi.org/10.1007/S10620-018-4936-4>.
 42. R. M. Satherley, S. Higgs, and R. Howard, “Disordered Eating Patterns in Coeliac Disease: A Framework Analysis,” *Journal of Human Nutrition and Dietetics* 30, no. 6 (2017): 724–736, <https://doi.org/10.1111/jhn.12475>.
 43. N. Trott, S. A. Raju, A. Rej, et al., “Long-Term follow-up in Patients With Coeliac Disease in the Pandemic-Era: A View From Sheffield the NHS England National Centre for Adult Coeliac Disease,” *Gastroenterol Hepatol from bed to bench* 16, no. 2 (2023): 158–166, <https://doi.org/10.22037/GHFBB.V16I2.2637>.
 44. K. Gładys, J. Dardzińska, M. Guzek, K. Adrych, Z. Kochan, and S. Małgorzewicz, “Expanded Role of a Dietitian in Monitoring a Gluten-Free Diet in Patients With Celiac Disease: Implications for Clinical Practice,” *Nutrients* 13, no. 6 (2021): 1859, <https://doi.org/10.3390/NU13061859>.
 45. K. Gładys, J. Dardzińska, M. Guzek, K. Adrych, and S. Małgorzewicz, “Celiac Dietary Adherence Test and Standardized Dietician Evaluation in Assessment of Adherence to a Gluten-Free Diet in Patients With Celiac Disease,” *Nutrients* 12, no. 8 (2020): 1–10, <https://doi.org/10.3390/nu12082300>.
 46. L. M. Sharkey, G. Corbett, E. Currie, J. Lee, N. Sweeney, and J. M. Woodward, “Optimising Delivery of Care in Coeliac Disease - Comparison of the Benefits of Repeat Biopsy and Serological Follow-Up,” *Alimentary Pharmacology and Therapeutics* 38, no. 10 (2013): 1278–1291, <https://doi.org/10.1111/apt.12510>.
 47. F. Biagi, C. Vattiato, S. Agazzi, et al., “A Second Duodenal Biopsy is Necessary in the Follow-up of Adult Coeliac Patients,” *Annals of Medicine* 46, no. 6 (2014): 430–433, <https://doi.org/10.3109/07853890.2014.913378>.
 48. A. Rej, R. L. Buckle, C. C. Shaw, et al., “National Survey Evaluating the Provision of Gastroenterology Dietetic Services in England,” *Frontline Gastroenterology* 12, no. 5 (2020): 380–384, <https://doi.org/10.1136/FLGASTRO-2020-101493>.
 49. Y. M. Jeanes, S. Kallos, H. Muhammad, and S. Reeves, “Who Gets an Annual Review for Coeliac Disease? Patients With Lower Health Literacy and Lower Dietary Adherence Consider Them Important,” *Journal of Human Nutrition and Dietetics* 37, no. 4 (2024): 1022–1031, <https://doi.org/10.1111/JHN.13314>.
 50. N. Venkateswaran, B. Claxton, D. Locke, et al., “Referral for Dietary Intervention in Celiac Disease Is Low Among Gastroenterologists and Primary Care Providers,” *Digestive Diseases* 41, no. 2 (2023): 343–352, <https://doi.org/10.1159/000525398>.
 51. P. Crespo-Escobar, M. Vázquez-Polo, M. van der Hofstadt, et al., “Knowledge Gaps in Gluten-Free Diet Awareness Among Patients and Healthcare Professionals: A Call for Enhanced Nutritional Education,” *Nutrition* 16, no. 15 (2024): 2512, <https://doi.org/10.3390/NU16152512>.
 52. J. M. Kreutz, M. P. M. Adriaanse, E. M. C. van der Ploeg, and A. C. E. Vreugdenhil, “Narrative Review: Nutrient Deficiencies in Adults and Children With Treated and Untreated Celiac Disease,” *Nutrients* 12, no. 2 (2020): 500, <https://doi.org/10.3390/NU12020500>.
 53. N. J. Wierdsma, M. A. E. van Bokhorst-de van der Schueren, M. Berkenpas, et al., “Vitamin and Mineral Deficiencies are Highly Prevalent in Newly Diagnosed Celiac Disease Patients,” *Nutrients* 5, no. 10 (2013): 3975e92, <https://doi.org/10.3390/NU5103975>.
 54. A. C. Bledsoe, K. S. King, J. J. Larson, et al., “Micronutrient Deficiencies Are Common in Contemporary Celiac Disease Despite Lack of Overt Malabsorption Symptoms,” *Mayo Clinic Proceedings* 94, no. 7 (2019): 1253–1260, <https://doi.org/10.1016/J.MAYOCP.2018.11.036>.
 55. S. Lamjadli, I. Oujamaa, I. Souli, et al., “Micronutrient Deficiencies in Patients With Celiac Disease: A Systematic Review and Meta-Analysis,” *International Journal of Immunopathology & Pharmacology* 39 (2025): 03946320241313426, <https://doi.org/10.1177/03946320241313426>.
 56. C. Mosca, F. Thorsteinsdottir, B. Abrahamsen, J. J. Rumessen, and M. N. Händel, “Newly Diagnosed Celiac Disease and Bone Health in Young Adults: A Systematic Literature Review,” *Calcified Tissue International* 110, no. 6 (2022): 641–648, <https://doi.org/10.1007/s00223-021-00938-w>.
 57. K. Nuermaimaiti, T. Li, N. Li, et al., “Vitamin and Trace Elements Imbalance are Very Common in Adult Patients With Newly Diagnosed Celiac Disease,” *Scientific Reports* 15, no. 1 (2025): 28315, <https://doi.org/10.1038/S41598-025-12631-1>.
 58. M. I. Pinto-Sanchez, J. J. Blom, P. R. Gibson, and D. Armstrong, “Nutrition Assessment and Management in Celiac Disease,” *Gastroenterology* 167, no. 1 (2024): 116–131.e1, <https://doi.org/10.1053/j.gastro.2024.02.049>.
 59. A. Ukkola, M. Mäki, K. Kurppa, et al., “Changes in Body Mass Index on a Gluten-Free Diet in Coeliac Disease: A Nationwide Study,” *European Journal of Internal Medicine* 23, no. 4 (2012): 384–388, <https://doi.org/10.1016/j.ejim.2011.12.012>.
 60. C. Hallert, M. Svensson, J. Tholstrup, and B. Hultberg, “Clinical Trial: B Vitamins Improve Health in Patients With Coeliac Disease Living on a Gluten-free Diet,” *Alimentary Pharmacology & Therapeutics* 29, no. 8 (2009): 811–816, <https://doi.org/10.1111/j.1365-2036.2009.03945.x>.
 61. N. Úbeda, M. P. González, M. Achón, et al., “Nutritional Composition of Breakfast in Children and Adolescents With and Without Celiac Disease in Spain—Role of Gluten-Free Commercial Products,” *Nutrients* 15, no. 10 (2023): 2368, <https://doi.org/10.3390/NU15102368>.
 62. T. G. Theethira, M. Dennis, and D. A. Leffler, “Nutritional Consequences of Celiac Disease and the Gluten-Free Diet,” *Expert Review of Gastroenterology & Hepatology* 8, no. 2 (2014): 123–129, <https://doi.org/10.1586/17474124.2014.876360>.

63. T. G. Theethira and M. Dennis, "Celiac Disease and the Gluten-free Diet: Consequences and Recommendations for Improvement," *Digestive Diseases* 33, no. 2 (2015): 175–182, <https://doi.org/10.1159/000369504>.
64. C. C. Payette, C. Desjardins, E. Lalanne, M. Marquis, and M. Perreault, "Exploring Challenges Faced by Adults Living With Celiac Disease: A Food Literacy Perspective," *Journal of Human Nutrition and Dietetics* 38, no. 2 (2025): e70057, <https://doi.org/10.1111/JHN.70057>.
65. M. Guennouni, B. Admou, N. El Khoudri, et al., "Gluten Contamination in Labelled Gluten-Free, Naturally Gluten-Free and Meals in Food Services in Low-Middle- and High-Income Countries: A Systematic Review and Meta-Analysis," *British Journal of Nutrition* 127, no. 10 (2022): 1528–1542, <https://doi.org/10.1017/S0007114521002488>.
66. H. Y. Öcal and H. G. Özel, "Naturally Gluten-Free Flours are Commonly Contaminated, While Commercially Produced Gluten-Free Flours are Relatively Safe: A Market-Based Study in Turkey," *Frontiers in Nutrition* 12 (2025): 1707584, <https://doi.org/10.3389/fnut.2025.1707584>.
67. Y. Zhang, M. Xu, X. Zhang, Y. Hu, and G. Luan, "Application of Zein in Gluten-Free Foods: A Comprehensive Review," *Food Research International* 160 (2022): 111722, <https://doi.org/10.1016/j.foodres.2022.111722>.
68. C. Costas-Batlle, N. Trott, Y. Jeanes, L. Seamark, and C. Gardiner, "A Dietitian-Led Coeliac Service Helps to Identify and Reduce Involuntary Gluten Ingestion With Subsequent Reduction in the Frequency of Repeat Endoscopies," *Journal of Human Nutrition and Dietetics* 36, no. 5 (2023): 1751–1759, <https://doi.org/10.1111/JHN.13206>.
69. K. Mårild, J. Söderling, S. R. Bozorg, et al., "Costs and Use of Health Care in Patients With Celiac Disease: A Population-Based Longitudinal Study," *American Journal of Gastroenterology* 115, no. 8 (2020): 1253–1263, <https://doi.org/10.14309/AJG.0000000000000652>.
70. C. Sategna-Guidetti, S. B. Grosso, S. B. Grosso, et al., "The Effects of 1-Year Gluten Withdrawal on Bone Mass, Bone Metabolism and Nutritional Status in Newly-Diagnosed Adult Coeliac Disease Patients," *Alimentary Pharmacology & Therapeutics* 14, no. 1 (2000): 35–43, <https://doi.org/10.1046/j.1365-2036.2000.00671.x>.
71. A. Schieppatti, S. Maimaris, S. Randazzo, et al., "Resilience in Adult Coeliac Patients on a Gluten-Free Diet: A Cross-Sectional Multicentre Italian," *Study* 16, no. 16 (2024): 2595, <https://doi.org/10.3390/NU16162595>.
72. N. Peleg, Y. Niv, R. Dickman, et al., "The Effects of Gluten-Free Diet on Body Mass Indexes in Adults With Celiac Disease: A Systematic Review and Meta-Analysis of Observational Studies," *Journal of Clinical Gastroenterology* 58, no. 10 (March 2024): 989–997, <https://doi.org/10.1097/MCG.0000000000001998>.
73. C. Xin, R. Imanifard, M. Jarahzadeh, P. Rohani, P. Velu, and M. H. Sohoul, "Impact of Gluten-Free Diet on Anthropometric Indicators in Individuals With and Without Celiac Disease: A Systematic Review and Meta-Analysis," *Clinical Therapeutics* 45, no. 12 (2023): e243–e251, <https://doi.org/10.1016/j.clinthera.2023.09.018>.
74. T. A. Kabbani, A. Goldberg, C. P. Kelly, et al., "Body Mass Index and the Risk of Obesity in Coeliac Disease Treated With the Gluten-Free Diet," *Alimentary Pharmacology & Therapeutics* 35, no. 6 (2012): 723–729, <https://doi.org/10.1111/j.1365-2036.2012.05001.x>.
75. S. Mora, G. Barera, S. Beccio, et al., "A Prospective, Longitudinal Study of the Long-Term Effect of Treatment on Bone Density in Children With Celiac Disease," *Jornal de Pediatria* 139, no. 4 (2001): 516–521, <https://doi.org/10.1067/mpd.2001.116298>.
76. K. Papamichael, E. Kokkinakis, E. J. Archavlis, et al., "S2044 Effect of a Gluten Free Diet on Bone Mineral Density in Patients With Celiac Disease," *Gastroenterology* 138, no. 5 (2010): S308, [https://doi.org/10.1016/S0016-5085\(10\)61416-3](https://doi.org/10.1016/S0016-5085(10)61416-3).
77. M. Marciniak, A. Szymczak-Tomczak, D. Mahadea, P. Eder, A. Dobrowolska, and I. Krela-Kaźmierczak, "Multidimensional Disadvantages of a Gluten-Free Diet in Celiac Disease: A Narrative Review," *Nutrients* 13, no. 2 (2021): 1–15, <https://doi.org/10.3390/NU13020643>.
78. L. A. Russell, P. Alliston, D. Armstrong, E. F. Verdu, P. Moayyedi, and M. I. Pinto-Sanchez, "Micronutrient Deficiencies Associated With a Gluten-Free Diet in Patients With Celiac Disease and Non-Celiac Gluten or Wheat Sensitivity: A Systematic Review and Meta-Analysis," *Journal of Clinical Medicine* 14, no. 14 (2025): 4848, <https://doi.org/10.3390/JCM14144848>.
79. B. Allen and C. Orfila, "The Availability and Nutritional Adequacy of Gluten-Free Bread and Pasta," *Nutrients* 10, no. 10 (2018): 1370, <https://doi.org/10.3390/NU10101370>.
80. V. Melini and F. Melini, "Gluten-Free Diet: Gaps and Needs for a Healthier Diet," *Nutrients* 11, no. 1 (2019): 170, <https://doi.org/10.3390/NU11010170>.
81. A. Cardo, I. Churrua, A. Lasa, et al., "Nutritional Imbalances in Adult Celiac Patients Following a Gluten-Free Diet," *Nutrients* 13, no. 8 (2021): 2877, <https://doi.org/10.3390/NU13082877>.
82. W. Mehtab, S. Agarwal, H. Agarwal, et al., "Gluten-Free Foods are Expensive and Nutritionally Imbalanced Than Their Gluten-Containing Counterparts," *Indian Journal of Gastroenterology* 43, no. 3 (2024): 668–678, <https://doi.org/10.1007/S12664-024-01519-Z>.
83. A. C. Parada, C. Méndez, and C. Aguirre, "Excess Weight and Gastrointestinal Symptoms in Chilean Celiac Patients at the Time of Diagnosis," *Revista Española de Enfermedades Digestivas* 111, no. 5 (2019): 384–387, <https://doi.org/10.17235/reed.2019.5251/2017>.
84. W. Dickey and N. Kearney, "Overweight in Celiac Disease: Prevalence, Clinical Characteristics, and Effect of a Gluten-Free Diet," *American Journal of Gastroenterology* 101, no. 10 (2006): 2356–2359, <https://doi.org/10.1111/J.1572-0241.2006.00750.X>.
85. M. Barone, A. Iannone, F. Cristofori, et al., "Risk of Obesity During a Gluten-Free Diet in Pediatric and Adult Patients With Celiac Disease: A Systematic Review With Meta-Analysis," *Nutrition Reviews* 81, no. 3 (2023): 252–266, <https://doi.org/10.1093/NUTRIT/NUAC052>.
86. C. Canova, I. Rosato, I. Marsilio, et al., "Quality of Life and Psychological Disorders in Coeliac Disease: A Prospective Multicentre Study," *Nutrients* 13, no. 9 (2021): 3233, <https://doi.org/10.3390/NU13093233>.
87. D. A. Leffler, M. Dennis, J. B. Edwards George, et al., "A Simple Validated Gluten-Free Diet Adherence Survey for Adults With Celiac Disease," *Clinical Gastroenterology and Hepatology* 7, no. 5 (2009): 530–536.e2, <https://doi.org/10.1016/J.CGH.2008.12.032>.
88. L. Elli, D. Leffler, C. Cellier, et al., "Guidelines for Best Practices in Monitoring Established Coeliac Disease in Adult Patients," *Nature Reviews Gastroenterology & Hepatology* 21, no. 3 (2024): 198–215, <https://doi.org/10.1038/S41575-023-00872-2>.
89. A. Atsawarungrangkit, J. A. Silvester, D. Weiten, et al., "Development of the Dietitian Integrated Evaluation Tool for Gluten-Free Diets (DIET-GFD)," *Nutrition* 78 (2020): 110819, <https://doi.org/10.1016/J.NUT.2020.110819>.
90. J. Syage, A. Ramos, V. Loskutov, et al., "Dynamics of Serologic Change to Gluten in Celiac Disease Patients," *Nutrition* 15, no. 24 (2023): 5083, <https://doi.org/10.3390/NU15245083>.
91. V. Lombardo, A. Scricciolo, A. Costantino, et al., "Evaluation of a Single Determination of Gluten Immunogenic Peptides in Urine From Unaware Celiac Patients to Monitor Gluten-Free Diet Adherence," *Nutrients* 15, no. 5 (2023): 1259, <https://doi.org/10.3390/NU15051259>.
92. B. Porcelli, F. Ferretti, F. Cinci, et al., "Fecal Gluten Immunogenic Peptides as Indicators of Dietary Compliance in Celiac Patients,"

- Minerva Gastroenterologica e Dietologica* 66, no. 3 (2020): 201–207, <https://doi.org/10.23736/S1121-421X.20.02662-8>.
93. L. Coto, I. Mendia, C. Sousa, J. C. Bai, and A. Cebolla, “Determination of Gluten Immunogenic Peptides for the Management of the Treatment Adherence of Celiac Disease: A Systematic Review,” *World Journal of Gastroenterology* 27, no. 37 (2021): 6306–6321, <https://doi.org/10.3748/WJG.V27.I37.6306>.
 94. M. Logan, M. MacKinder, C. M. Clark, et al., “Intestinal Fatty Acid Binding Protein is a Disease Biomarker in Paediatric Coeliac Disease and Crohn’s Disease,” *BMC Gastroenterology* 22, no. 1 (2022): 260, <https://doi.org/10.1186/S12876-022-02334-6>.
 95. M. P. M. Adriaanse, A. Mubarak, R. G. Riedl, et al., “Progress Towards Non-invasive Diagnosis and Follow-Up of Celiac Disease in Children; A Prospective Multicentre Study to the Usefulness of Plasma I-FABP,” *Scientific Reports* 7, no. 1 (2017): 8671, <https://doi.org/10.1038/S41598-017-07242-4>.
 96. M. P. M. Adriaanse, D. A. Leffler, C. P. Kelly, et al., “Serum I-FABP Detects Gluten Responsiveness in Adult Celiac Disease Patients on a Short-Term Gluten Challenge,” *American Journal of Gastroenterology* 111, no. 7 (2016): 1014–1022, <https://doi.org/10.1038/AJG.2016.162>.
 97. M. P. M. Adriaanse, G. J. Tack, V. L. Passos, et al., “Serum I-FABP as Marker for Enterocyte Damage in Coeliac Disease and its Relation to Villous Atrophy and Circulating Autoantibodies,” *Alimentary Pharmacology & Therapeutics* 37, no. 4 (2013): 482–490, <https://doi.org/10.1111/APT.12194>.
 98. L. Kivelä, K. Lindfors, K. E. A. Lundin, and K. Størdal, “Review Article: Faecal Biomarkers for Assessing Small Intestinal Damage in Coeliac Disease and Environmental Enteropathy,” *Alimentary Pharmacology & Therapeutics* 60, no. 8 (2024): 988–1004, <https://doi.org/10.1111/APT.18234>.
 99. A. Schieppatti, A. Cappellini, S. Maimaris, et al., “Fecal Calprotectin Measurement as a Biomarker of Severe Disease Phenotype in Celiac Disease and Non-Celiac Enteropathies,” *Digestive and Liver Disease* 57, no. 1 (2025): 308–314, <https://doi.org/10.1016/j.dld.2024.09.010>.
 100. N. J. Hall, G. Rubin, and A. Charnock, “Systematic Review: Adherence to a Gluten-free Diet in Adult Patients With Coeliac Disease,” *Alimentary Pharmacology & Therapeutics* 30, no. 4 (2009): 315–330, <https://doi.org/10.1111/J.1365-2036.2009.04053.X>.
 101. P. Paarlahti, K. Kurppa, A. Ukkola, et al., “Predictors of Persistent Symptoms and Reduced Quality of Life in Treated Coeliac Disease Patients: A Large Cross-Sectional Study,” *BMC Gastroenterology* 13, no. 1 (2013): 75, <https://doi.org/10.1186/1471-230X-13-75>.
 102. J. Villafuerte-Galvez, R. R. Vanga, M. Dennis, et al., “Factors Governing Long-Term Adherence to a Gluten-Free Diet in Adult Patients With Coeliac Disease,” *Alimentary Pharmacology & Therapeutics* 42, no. 6 (2015): 753–760, <https://doi.org/10.1111/APT.13319>.
 103. S. Hopkins and J. M. Soon, “Nutritional Quality, Cost and Availability of Gluten-Free Food in England,” *British Food Journal* 121, no. 11 (2019): 2867–2882, <https://doi.org/10.1108/bfj-09-2018-0607>.
 104. A. Schieppatti, S. Maimaris, M. L. Nicolardi, et al., “Determinants and Trends of Adherence to a Gluten-Free Diet in Adult Celiac Patients on a Long-term Follow-up (2000–2020),” *Clinical Gastroenterology and Hepatology* 20, no. 4 (2022): e741–e749, <https://doi.org/10.1016/J.CGH.2020.12.015>.
 105. N. Trott, A. Rej, S. H. Coleman, and D. S. Sanders, “Adult Celiac Disease With Persistent IBS-Type Symptoms: A Pilot Study of an Adjuvant FODMAP Diet,” *Gastroenterol Hepatol From Bed to Bench* 14, no. 4 (2021): 304–310, <https://pubmed.ncbi.nlm.nih.gov/34659657/>.
 106. F. van Megen, G. I. Skodje, S. Lergenmuller, et al., “A Low FODMAP Diet Reduces Symptoms in Treated Celiac Patients With Ongoing Symptoms—A Randomized Controlled Trial,” *Clinical Gastroenterology and Hepatology* 20, no. 10 (2022): 2258–2266.e3, <https://doi.org/10.1016/J.CGH.2022.01.011>.
 107. M. G. Shiha, A. Schieppatti, F. Manza, S. Maimaris, I. Aziz, and D. S. Sanders, “Global Prevalence of Celiac Disease in Patients With Rome III and Rome IV Irritable Bowel Syndrome: A Systematic Review and Meta-Analysis,” *American Journal of Gastroenterology* 120, no. 12 (2025): 2776–2787, <https://doi.org/10.14309/AJG.0000000000003586>.
 108. S. Parker, O. Palsson, D. S. Sanders, et al., “Functional Gastrointestinal Disorders and Associated Health Impairment in Individuals With Celiac Disease,” *Clinical Gastroenterology and Hepatology* 20, no. 6 (2022): 1315–1325, <https://doi.org/10.1016/j.cgh.2021.07.026>.
 109. P. Usai-Satta, M. Lai, and F. Oppia, “Lactose Malabsorption and Presumed Related Disorders: A Review of Current Evidence,” *Nutrients* 14, no. 3 (2022): 584, <https://doi.org/10.3390/NU14030584>.
 110. V. Ojetti, M. Gabrielli, A. Migneco, et al., “Regression of Lactose Malabsorption in Coeliac Patients After Receiving a Gluten-Free Diet,” *Scandinavian Journal of Gastroenterology* 43, no. 2 (2008): 174–177, <https://doi.org/10.1080/00365520701676138>.
 111. J. Dionne, A. C. Ford, Y. Yuan, et al., “A Systematic Review and Meta-Analysis Evaluating the Efficacy of a Gluten-Free Diet and a Low FODMAPs Diet in Treating Symptoms of Irritable Bowel Syndrome,” *American Journal of Gastroenterology* 113, no. 9 (2018): 1290–1300, <https://doi.org/10.1038/S41395-018-0195-4>.
 112. H. Haghibin, F. Hasan, M. K. Gangwani, et al., “Efficacy of Dietary Interventions for Irritable Bowel Syndrome: A Systematic Review and Network Meta-Analysis,” *Journal of Clinical Medicine* 13, no. 24 (2024): 7531, <https://doi.org/10.3390/JCM13247531>.
 113. F. Lusetti, A. Schieppatti, D. Scalvini, S. Maimaris, and F. Biagi, “Efficacy of a Low-FODMAP Diet for Coeliac Patients With Persistent IBS-like Symptoms Despite a Gluten-Free Diet: A Systematic Review,” *Nutrients* 16, no. 7 (2024): 1094, <https://doi.org/10.3390/NU16071094>.
 114. C. Rose, G. U. Law, and R. A. Howard, “The Psychosocial Experiences of Adults Diagnosed With Coeliac Disease: A Qualitative Evidence Synthesis,” *Quality of Life Research* 33, no. 1 (2024): 1–16, <https://doi.org/10.1007/s11136-023-03483-1>.
 115. S. Hansen, M. Osler, S. M. Thysen, J. J. Rumessen, A. Linneberg, and L. L. Kårhus, “Celiac Disease and Risk of Neuropsychiatric Disorders: A Nationwide Cohort Study,” *Acta Psychiatrica Scandinavica* 148, no. 1 (2023): 60–70, <https://doi.org/10.1111/ACPS.13554>.
 116. F. Zingone, G. L. Swift, T. R. Card, D. S. Sanders, J. F. Ludvigsson, and J. C. Bai, “Psychological Morbidity of Celiac Disease: A Review of the Literature,” *United European Gastroenterology Journal* 3, no. 2 (2015): 136–145, <https://doi.org/10.1177/2050640614560786>.
 117. S. P. Möller, P. Apputhurai, J. A. Tye-Din, and S. R. Knowles, “Quality of Life in Coeliac Disease: Relationship Between Psychosocial Processes and Quality of Life in a Sample of 1697 Adults Living With Coeliac Disease,” *Journal of Psychosomatic Research* 151 (2021): 110652, <https://doi.org/10.1016/J.JPSYCHORES.2021.110652>.
 118. E. Clappison, M. Hadjivassiliou, and P. Zis, “Psychiatric Manifestations of Coeliac Disease, a Systematic Review and Meta-Analysis,” *Nutrients* 12, no. 1 (2020): 142, <https://doi.org/10.3390/NU12010142>.
 119. Z. Nikniaz, S. Beheshti, M. Abbasalizad Farhangi, and L. Nikniaz, “A Systematic Review and Meta-Analysis of the Prevalence and Odds of Eating Disorders in Patients With Celiac Disease and Vice-Versa,” *International Journal of Eating Disorders* 54, no. 9 (2021): 1563–1574, <https://doi.org/10.1002/EAT.23561>.
 120. R. M. Satherley, R. Howard, and S. Higgs, “The Prevalence and Predictors of Disordered Eating in Women With Coeliac Disease,” *Appetite* 107 (2016): 260–267, <https://doi.org/10.1016/j.appet.2016.07.038>.
 121. J. W. Cadenhead, R. L. Wolf, B. Lebowitz, et al., “Diminished Quality of Life Among Adolescents With Coeliac Disease Using Maladaptive Eating Behaviours to Manage a Gluten-Free Diet: A Cross-Sectional, Mixed-Methods Study,” *Journal of Human Nutrition and Dietetics* 32, no. 3 (2019): 311–320, <https://doi.org/10.1111/jhn.12638>.

122. H. G. Gercek, S. A. İpek Bas, A. Kara, and A. Bukulmez, "Examination of Eating Attitudes and Symptoms of Orthorexia Nervosa in Adolescents With and Without Celiac Disease," *Journal of Eating Disorders* 13, no. 1 (2025): 102, <https://doi.org/10.1186/S40337-025-01294-Y>.
123. J. E. Peters, C. Basnayake, G. S. Hebbard, M. R. Salzberg, and M. A. Kamm, "Prevalence of Disordered Eating in Adults With Gastrointestinal Disorders: A Systematic Review," *Neuro-Gastroenterology and Motility* 34, no. 8 (2022): e14278, <https://doi.org/10.1111/nmo.14278>.
124. G. Addolorato, G. De Lorenzi, L. Abenavoli, L. Leggio, E. Capristo, and G. Gasbarrini, "Psychological Support Counselling Improves Gluten-free Diet Compliance in Coeliac Patients With Affective Disorders," *Alimentary Pharmacology & Therapeutics* 20, no. 7 (2004): 777–782, <https://doi.org/10.1111/J.1365-2036.2004.02193.X>.
125. E. P. Halmos, M. Deng, S. R. Knowles, K. Sainsbury, B. Mullan, and J. A. Tye-Din, "Food Knowledge and Psychological State Predict Adherence to a Gluten-Free Diet in a Survey of 5310 Australians and New Zealanders With Coeliac Disease," *Alimentary Pharmacology & Therapeutics* 48, no. 1 (2018): 78–86, <https://doi.org/10.1111/APT.14791>.
126. M. Hadavi, H. Mirhosseini, M. Barakati, M. Hadavi, and R. Bidaki, "Efficacy of Acceptance and Commitment Therapy on Depression and Anxiety of Patients With Celiac (Coeliac) and It's Relation to Therapeutic Response in Yazd," *Psychol Disord Res* 2021, no. 1 (2021): 1–6, <https://doi.org/10.31487/J.PDR.2021.01.03>.
127. L. Ring Jacobsson, M. Friedrichsen, A. Göransson, and C. Hallert, "Does a Coeliac School Increase Psychological Well-Being in Women Suffering From Coeliac Disease, Living on a Gluten-Free Diet?," *Journal of Clinical Nursing* 21, no. 5–6 (2012): 766–775, <https://doi.org/10.1111/J.1365-2702.2011.03953.X>.
128. J. F. Ludvigsson, L. Agreus, C. Ciacci, et al., "Transition From Childhood to Adulthood in Coeliac Disease: The Prague Consensus Report," *Gut* 65, no. 8 (2016): 1242–1251, <https://doi.org/10.1136/gutjnl-2016-311574>.
129. M. L. Mearin, D. Agardh, H. Antunes, et al., "ESPGHAN Position Paper on Management and Follow-up of Children and Adolescents With Celiac Disease," *Journal of Pediatric Gastroenterology and Nutrition* 75, no. 3 (2022): 369–386, <https://doi.org/10.1097/MPG.00000000000003540>.
130. N. R. Reilly, M. L. Hammer, J. F. Ludvigsson, and P. H. Green, "Frequency and Predictors of Successful Transition of Care for Young Adults With Childhood Celiac Disease," *Journal of Pediatric Gastroenterology and Nutrition* 70, no. 2 (2020): 190–194, <https://doi.org/10.1097/MPG.00000000000002568>.
131. L. Kiveli, S. Hekkala, H. Huhtala, K. Kaukinen, and K. Kurppa, "Lack of Long-Term Follow-Up After Paediatric-Adult Transition in Coeliac Disease is Not Associated With Complications, Ongoing Symptoms or Dietary Adherence," *United European Gastroenterology Journal* 8, no. 2 (2020): 157–166, <https://doi.org/10.1177/2050640619900077>.
132. F. Zingone, S. Massa, B. Malamisura, P. Pisano, and C. Ciacci, "Coeliac Disease: Factors Affecting the Transition and a Practical Tool for the Transition to Adult Healthcare," *United European Gastroenterology Journal* 6, no. 9 (2018): 1356–1362, <https://doi.org/10.1177/2050640618787651>.
133. A. Schieppati, S. Maimaris, C. de Queiros Mattoso Archela dos Santos, G. Rusca, S. Costa, and F. Biagi, "Long-Term Adherence to a Gluten-Free Diet and Quality of Life of Celiac Patients After Transition to an Adult Referral Center," *Digestive Diseases and Sciences* 67, no. 8 (2022): 3955–3963, <https://doi.org/10.1007/S10620-021-07231-8>.
134. A. R. Lee, R. Wolf, I. Contento, H. Verdell, and P. H. R. Green, "Coeliac Disease: The Association Between Quality of Life and Social Support Network Participation," *Journal of Human Nutrition and Dietetics* 29, no. 3 (2016): 383–390, <https://doi.org/10.1111/JHN.12319>.
135. G. Perez-Junkera, L. Ruiz de Azua, M. Vázquez-Polo, et al., "Global Approach to Follow-Up of Celiac Disease," *Foods* 13, no. 10 (2024): 1449, <https://doi.org/10.3390/FOODS13101449>.
136. A. Roy, M. Laszkowska, J. Sundström, et al., "Prevalence of Celiac Disease in Patients With Autoimmune Thyroid Disease," *A Meta-Analysis* 26, no. 7 (2016): 880–890, <https://doi.org/10.1089/thy.2016.0108>.
137. X. Sun, L. Lu, R. Yang, Y. Li, L. Shan, and Y. Wang, "Increased Incidence of Thyroid Disease in Patients With Celiac Disease: A Systematic Review and Meta-Analysis," *PLoS One* 11, no. 12 (2016): e0168708, <https://doi.org/10.1371/JOURNAL.PONE.0168708>.
138. T. D'Ambrosio, S. Bianchini, R. Gastaldi, et al., "A Systematic Review of Guidelines on Screening for Celiac Disease in Children With Thyroid Disease and Vice Versa," *Frontiers in Pediatrics* 13 (2025): 1538409, <https://doi.org/10.3389/fped.2025.1538409>.
139. J. R. Garber, R. H. Cobin, H. Gharib, et al., "Clinical Practice Guidelines for Hypothyroidism in Adults: Cosponsored by the American Association of Clinical Endocrinologists and the American Thyroid Association," *Thyroid* 22, no. 12 (2012): 1200–1235, <https://doi.org/10.1089/THY.2012.0205>.
140. L. P. Jelsness-Jørgensen, T. Bernklev, and K. E. A. Lundin, "Fatigue as an Extra-Intestinal Manifestation of Celiac Disease: A Systematic Review," *Nutrients* 10, no. 11 (2018): 1652, <https://doi.org/10.3390/NU10111652>.
141. S. Errichiello, O. Esposito, R. Di Mase, et al., "Celiac Disease: Predictors of Compliance With a Gluten-Free Diet in Adolescents and Young Adults," *Journal of Pediatric Gastroenterology and Nutrition* 50, no. 1 (2010): 54–60, <https://doi.org/10.1097/MPG.0B013E31819DE82A>.
142. H. Söderström, M. Cervin, J. Dereke, et al., "Does a Gluten-Free Diet Lead to Better Glycemic Control in Children With Type 1 Diabetes? Results from a Feasibility Study and Recommendations for Future Trials," *Contemporary Clinical Trials Communications* 26 (2022): 100893, <https://doi.org/10.1016/j.conctc.2022.100893>.
143. I. Eland, L. Klieverik, A. A. Mansour, and A. Al-Toma, "Gluten-Free Diet in Co-Existent Celiac Disease and Type 1 Diabetes Mellitus: Is It Detrimental or Beneficial to Glycemic Control, Vascular Complications, and Quality of Life?," *Nutrients* 15, no. 1 (2022): 199, <https://doi.org/10.3390/NU15010199>.
144. Y. Zhang, S. Yang, and P. Wang, "Gluten-Free Diets for Metabolic Control of Type 1 Diabetes Mellitus in Children and Adolescents: A Systematic Review and Meta-Analysis," *Archives of Endocrinology and Metabolism* 68 (2025): e240165, <https://doi.org/10.20945/2359-4292-2024-0165>.
145. D. I. Weiman, F. H. Mahmud, A. B. M. Clarke, et al., "Impact of a Gluten-Free Diet on Quality of Life and Health Perception in Patients With Type 1 Diabetes and Asymptomatic Celiac Disease," *Journal of Clinical Endocrinology and Metabolism* 106, no. 5 (2021): E1984–E1992, <https://doi.org/10.1210/CLINEM/DGAA977>.
146. J. Svensson, S. M. Sildorf, C. B. Pipper, et al., "Potential Beneficial Effects of a Gluten-Free Diet in Newly Diagnosed Children With Type 1 Diabetes: A Pilot Study," *SpringerPlus* 5, no. 1 (2016): 994, <https://doi.org/10.1186/S40064-016-2641-3>.
147. A. M. Hegab and A. Abou-Taleb, "Effect of Gluten-Free Diet on Metabolic Control and Growth Parameters Among Children and Adolescents With Type 1 Diabetes During the First Year After Diagnosis of Celiac Disease: A Retrospective Case-Control Study," *Pediatric Diabetes* 2025, no. 1 (2025): 1283259, <https://doi.org/10.1155/PEDI/1283259>.
148. E. Mozzillo, R. Franceschi, F. Di Candia, et al., "The Impact of Gluten-Free Diet on Growth, Metabolic Control and Quality of Life in Youth With Type 1 Diabetes and Celiac Disease: A Systematic Review," *Diabetes Research and Clinical Practice* 191 (2022): 110032, <https://doi.org/10.1016/J.DIABRES.2022.110032>.
149. R. Franceschi, R. Pertile, M. Marigliano, et al., "Maintaining a Gluten-Free Diet Is Associated With Quality of Life in Youths With Type 1 Diabetes and Celiac Disease," *Acta Diabetologica* 61, no. 8 (2024): 987–995, <https://doi.org/10.1007/S00592-024-02281-6>.

150. A. Pham-Short, K. C. Donaghue, G. Ambler, S. Garnett, and M. E. Craig, "Quality of Life in Type 1 Diabetes and Celiac Disease: Role of the Gluten-Free Diet," *Journal de Pédiatrie* 179 (2016): 131–138.e1, <https://doi.org/10.1016/j.jpeds.2016.08.105>.
151. D. Pitocco, S. Giubilato, F. Martini, et al., "Combined Atherogenic Effects of Celiac Disease and Type 1 Diabetes Mellitus," *Atherosclerosis* 217, no. 2 (2011): 531–535, <https://doi.org/10.1016/j.atherosclerosis.2011.04.042>.
152. T. R. Rohrer, J. Wolf, S. Liptay, et al., "Microvascular Complications in Childhood-Onset Type 1 Diabetes and Celiac Disease: A Multicenter Longitudinal Analysis of 56,514 Patients From the German-Austrian DPV Database," *Diabetes Care* 38, no. 5 (2015): 801–807, <https://doi.org/10.2337/dc14-0683>.
153. A. Pham-Short, K. C. Donaghue, G. Ambler, S. Garnett, and M. E. Craig, "Greater Postprandial Glucose Excursions and Inadequate Nutrient Intake in Youth With Type 1 Diabetes and Celiac Disease," *Scientific Reports* 7 (2017): 45286, <https://doi.org/10.1038/SREP45286>.
154. F. H. Mahmud, E. N. De Melo, K. Noordin, et al., "The Celiac Disease and Diabetes-Dietary Intervention and Evaluation Trial (CD-DIET) Protocol: A Randomised Controlled Study to Evaluate Treatment of Asymptomatic Coeliac Disease in Type 1 Diabetes," *BMJ Open* 5, no. 5 (2015): e008097, <https://doi.org/10.1136/BMJOPEN-2015-008097>.
155. K. V. Hatlen, T. M. Lysell Lensnes, C. Henriksen, T. J. Berg, I. Nermoen, and K. E. A. Lundin, "Suboptimal Adherence to a Gluten-Free Diet in Adults With Both Type 1 Diabetes and Celiac Disease Using Urinary Gluten Immunogenic Peptide Measurement," *Scandinavian Journal of Gastroenterology* 60, no. 1 (2025): 28–36, <https://doi.org/10.1080/00365521.2024.2442688>.
156. H. Söderström, M. Cervin, J. Dereke, et al., "Does a Gluten-Free Diet Lead to Better Glycemic Control in Children With Type 1 Diabetes? Results From a Feasibility Study and Recommendations for Future Trials," *Contemp Clin Trials Commun* 26 (2022): 100893, <https://doi.org/10.1016/j.conctc.2022.100893>.
157. L. Kivelä, A. Eurén, M. Repo, H. Huhtala, K. Kaukinen, and K. Kurppa, "Coexisting Type 1 Diabetes, Persistent Symptoms, and Financial Issues Associate With Poorer Adherence to a Gluten-Free Diet in Celiac Disease After Transition From Pediatrics to Adult Care," *Frontiers in Nutrition* 9 (2022): 883220, <https://doi.org/10.3389/FNUT.2022.883220>.
158. V. Passananti, A. Santonicola, C. Bucci, et al., "Bone Mass in Women With Celiac Disease: Role of Exercise and Gluten-Free Diet," *Digestive and Liver Disease* 44, no. 5 (2012): 379–383, <https://doi.org/10.1016/j.dld.2011.12.012>.
159. M. R. Jafri, C. W. Nordstrom, J. A. Murray, et al., "Long-Term Fracture Risk in Patients With Celiac Disease: A Population-Based Study in Olmsted County, Minnesota," *Digestive Diseases and Sciences* 53, no. 4 (2008): 964–971, <https://doi.org/10.1007/S10620-007-9976-0>.
160. E. Kamycheva, T. Goto, and C. A. Camargo, "Celiac Disease Is Associated With Reduced Bone Mineral Density and Increased FRAX Scores in the US National Health and Nutrition Examination Survey," *Osteoporosis International* 28, no. 3 (2017): 781–790, <https://doi.org/10.1007/s00198-016-3791-4>.
161. R. Ganji, M. Moghbeli, R. Sadeghi, G. Bayat, and A. Ganji, "Prevalence of Osteoporosis and Osteopenia in Men and Premenopausal Women With Celiac Disease: A Systematic Review," *BioMed Central Ltd.* 18, no. 1 (2019): 9, <https://doi.org/10.1186/s12937-019-0434-6>.
162. D. Meyer, S. Stavropoulos, B. Diamond, E. Shane, and P. H. Green, "Osteoporosis in a North American Adult Population With Celiac Disease," *American Journal of Gastroenterology* 96, no. 1 (2001): 112–119, <https://doi.org/10.1111/j.1572-0241.2001.03507.x>.
163. L. Posthumus and A. Al-Toma, "Duodenal Histopathology and Laboratory Deficiencies Related to Bone Metabolism in Coeliac Disease," *European Journal of Gastroenterology and Hepatology* 29, no. 8 (2017): 897–903, <https://doi.org/10.1097/MEG.0000000000000880>.
164. E. D. Newnham, S. J. Shepherd, B. J. Strauss, P. Hosking, and P. R. Gibson, "Adherence to the Gluten-Free Diet Can Achieve the Therapeutic Goals in Almost all Patients With Coeliac Disease: A 5-Year Longitudinal Study From Diagnosis," *Journal of Gastroenterology and Hepatology* 31, no. 2 (2016): 342–349, <https://doi.org/10.1111/jgh.13060>.
165. J. P. W. Burger, J. J. H. van der Laan, T. A. Jansen, et al., "Low Yield for Routine Laboratory Checks in follow-up of Coeliac Disease," *Journal of Gastrointestinal and Liver Diseases* 27, no. 3 (2018): 233–239, <https://doi.org/10.15403/gld.2014.1121.273.jph>.
166. M. Olmos, M. Antelo, H. Vazquez, E. Smecul, E. Mauriño, and J. C. C. Bai, "Systematic Review and Meta-Analysis of Observational Studies on the Prevalence of Fractures in Coeliac Disease," *Digestive and Liver Disease* 40, no. 1 (2008): 46–53, <https://doi.org/10.1016/j.dld.2007.09.006>.
167. M. W. Davie, I. Gaywood, E. George, et al., "Excess Non-Spine Fractures in Women Over 50 Years With Celiac Disease: A Cross-Sectional, Questionnaire-Based Study," *Osteoporosis International* 16, no. 9 (2005): 1150–1155, <https://doi.org/10.1007/S00198-004-1822-Z>.
168. J. West, R. F. A. Logan, T. R. Card, C. Smith, and R. Hubbard, "Fracture Risk in People With Celiac Disease: A Population-Based Cohort Study," *Gastroenterology* 125, no. 2 (2003): 429–436, [https://doi.org/10.1016/S0016-5085\(03\)00891-6](https://doi.org/10.1016/S0016-5085(03)00891-6).
169. M. Laszkowska, S. Mahadev, J. Sundström, et al., "Systematic Review With Meta-Analysis: The Prevalence of Coeliac Disease in Patients With Osteoporosis," *Alimentary Pharmacology & Therapeutics* 48, no. 6 (2018): 590–597, <https://doi.org/10.1111/apt.14911>.
170. S. Chavda, B. Chavda, and R. Dube, "Osteoporosis Screening and Fracture Risk Assessment Tool: Its Scope and Role in General Clinical Practice," *Cureus* 14, no. 7 (2022), <https://doi.org/10.7759/CUREUS.26518>.
171. J. J. Carey, P. Chih-Hsing Wu, and D. Bergin, "Risk Assessment Tools for Osteoporosis and Fractures in 2022," *Best Practice & Research Clinical Rheumatology* 36, no. 3 (August 2022): 101775, <https://doi.org/10.1016/J.BERH.2022.101775>.
172. A. Rastogi, S. K. Bhadada, A. Bhansali, R. Kochhar, and R. Santosh, "Celiac Disease: A Missed Cause of Metabolic Bone Disease," *Indian J Endocrinol Metab* 16, no. 5 (2012): 780–785, <https://doi.org/10.4103/2230-8210.100674>.
173. G. Evangelatos, K. Kouna, A. Iliopoulos, and G. E. Fragoulis, "Musculoskeletal Complications of Celiac Disease: A Case-Based Review," *Mediterr J Rheumatol* 34, no. 1 (2023): 86–90, <https://doi.org/10.31138/MJR.34.1.86>.
174. I. E. L. W. Bultink and W. F. Lems, "Performance of Vertebral Fracture Assessment in Addition to Dual Energy X-ray Absorptiometry in Patients With Rheumatoid Arthritis," *Rheumatology* 53, no. 5 (2014): 775–776, <https://doi.org/10.1093/rheumatology/ket448>.
175. M. Bandirali, E. Lanza, C. Messina, et al., "Dose Absorption in Lumbar and Femoral Dual Energy x-ray Absorptiometry Examinations Using Three Different Scan Modalities: An Anthropomorphic Phantom Study," *Journal of Clinical Densitometry* 16, no. 3 (2013): 279–282, <https://doi.org/10.1016/j.jocd.2013.02.005>.
176. J. A. Kanis, N. Harvey, C. Cooper, H. Johansson, A. Odén, and E. V. McCloskey, "A Systematic Review of Intervention Thresholds Based on FRAX: A Report Prepared for the National Osteoporosis Guideline Group and the International Osteoporosis Foundation," *Archives of Osteoporosis* 11, no. 1 (2016): 25, <https://doi.org/10.1007/S11657-016-0278-Z>.
177. T. Porcelli, F. Maffezzoni, L. C. Pezzaioli, A. Delbarba, C. Cappelli, and A. Ferlin, "Management of Endocrine Disease: Male Osteoporosis: Diagnosis and Management. Should the Treatment and Target Be the Same as for Female Osteoporosis?," *European Journal of Endocrinology*, no. June 1 (2020), <https://doi.org/10.1530/EJE-20-0034>.

178. B. L. Langdahl, "Osteoporosis in Premenopausal Women," *Current Opinion in Rheumatology* 29, no. 4 (2017): 410–415, <https://doi.org/10.1097/BOR.0000000000000400>.
179. A. Al-Toma, A. Herman, W. F. Lems, C. J. J. Mulder, and C. J. J. Mulder, "The Dietary and Non-Dietary Management of Osteoporosis in Adult-Onset Celiac Disease," *Current Status and Practical Guidance* 14, no. 21 (2022): 4554, <https://doi.org/10.3390/nu14214554>.
180. A. Di Sabatino, M. M. Rosado, P. Cazzola, et al., "Splenic Hypofunction and the Spectrum of Autoimmune and Malignant Complications in Celiac Disease," *Clinical Gastroenterology and Hepatology* 4, no. 2 (2006): 179–186, [https://doi.org/10.1016/S1542-3565\(05\)00982-1](https://doi.org/10.1016/S1542-3565(05)00982-1).
181. A. Di Sabatino, F. Biagi, P. G. Gobbi, and G. R. Corazza, "How I Treat Enteropathy-Associated T-cell Lymphoma," *Blood* 119, no. 11 (2012): 2458–2468, <https://doi.org/10.1182/blood-2011-10-385559>.
182. P. Giuffrida, N. Aronico, M. Rosselli, et al., "Defective Spleen Function in Autoimmune Gastrointestinal Disorders," *Internal and Emergency Medicine* 15, no. 2 (2020): 225–229, <https://doi.org/10.1007/s11739-019-02129-w>.
183. T. van Gils, P. Nijeboer, J. H. T. van Waesberghe, et al., "Splenic Volume Differentiates Complicated and Non-Complicated Celiac Disease," *United European Gastroenterology Journal* 5, no. 3 (2017): 374–379, <https://doi.org/10.1177/2050640616663571>.
184. M. Di Stefano, R. A. Jorizzo, G. Veneto, L. Cecchetti, G. Gasbarrini, and G. R. Corazza, "Bone Mass and Metabolism in Dermatitis Herpetiformis," *Digestive Diseases and Sciences* 44, no. 10 (1999): 2139–2143, <https://doi.org/10.1023/a:1026603309056>.
185. J. F. Ludvigsson, O. Olén, M. Bell, A. Ekbom, and S. M. Montgomery, "Coeliac Disease and Risk of Sepsis," *Gut* 57, no. 8 (2008): 1074–1080, <https://doi.org/10.1136/GUT.2007.133868>.
186. A. Welander, A. R. Tjernberg, S. M. Montgomery, J. Ludvigsson, and J. F. Ludvigsson, "Infectious Disease and Risk of Later Celiac Disease in Childhood," *Pediatrics* 125, no. 3 (2010): e530–e536, <https://doi.org/10.1542/peds.2009-1200>.
187. O. G. Moscatelli, A. K. Russell, L. M. Henneken, et al., "Impaired IgM Memory B Cell Function Is Common in Coeliac Disease But Conjugate Pneumococcal Vaccination Induces Robust Protective Immunity," *Vaccines* 12, no. 2 (2024): 214, <https://doi.org/10.3390/VACCINES12020214>.
188. P. N. Trewby, P. M. Chipping, S. J. Palmer, P. D. Roberts, S. M. Lewis, and J. S. Stewart, "Splenic Atrophy in Adult Coeliac Disease: Is It Reversible?," *Gut* 22, no. 8 (1981): 628–632, <https://doi.org/10.1136/GUT.22.8.628>.
189. G. R. Corazza, M. Frisoni, D. Vaira, and G. Gasbarrini, "Effect of Gluten-Free Diet on Splenic Hypofunction of Adult Coeliac Disease," *Gut* 24, no. 3 (1983): 228–230, <https://doi.org/10.1136/GUT.24.3.228>.
190. M. Kobayashi, J. L. Farrar, R. Gierke, et al., "Use of 15-Valent Pneumococcal Conjugate Vaccine and 20-Valent Pneumococcal Conjugate Vaccine Among U.S. Adults: Updated Recommendations of the Advisory Committee on Immunization Practices - United States, 2022," *MMWR Morb Mortal Wkly Rep* 71, no. 4 (2022): 109–117, <https://doi.org/10.15585/MMWR.MM7104A1>.
191. C. Canova, J. Ludvigsson, V. Baldo, C. Barbiellini Amidei, L. Zanier, and F. Zingone, "Risk of Bacterial Pneumonia and Pneumococcal Infection in Youths With Celiac Disease - A Population-Based Study," *Digestive and Liver Disease* 51, no. 8 (2019): 1101–1105, <https://doi.org/10.1016/J.DLD.2019.02.010>.
192. F. Zingone, A. Abdul Sultan, C. J. Crooks, L. J. Tata, C. Ciacci, and J. West, "The Risk of Community-Acquired Pneumonia Among 9803 Patients With Coeliac Disease Compared to the General Population: A Cohort Study," *Alimentary Pharmacology & Therapeutics* 44, no. 1 (2016): 57–67, <https://doi.org/10.1111/APT.13652>.
193. J. Jang, J. Krishnamurthy, and C. M. Nylund, "Association Between Celiac Disease and Pneumococcal Infections in Hospitalized Pediatric Patients in the United States," *Journal of Pediatric Gastroenterology and Nutrition* 79, no. 2 (2024): 335–342, <https://doi.org/10.1002/JPN3.12250>.
194. M. Hadjivassiliou, I. D. Croall, P. Zis, et al., "Neurologic Deficits in Patients With Newly Diagnosed Celiac Disease Are Frequent and Linked With Autoimmunity to Transglutaminase 6," *Clinical Gastroenterology and Hepatology* 17, no. 13 (2019): 2678–2686.e2, <https://doi.org/10.1016/J.CGH.2019.03.014>.
195. M. Hadjivassiliou, D. D. Sanders, and D. P. Aeschlimann, "Gluten-Related Disorders: Gluten Ataxia," *Digestive Diseases* 33, no. 2 (2015): 264–268, <https://doi.org/10.1159/000369509>.
196. D. Bolotin and V. Petronic-Rosic, "Dermatitis Herpetiformis. Part II. Diagnosis, Management, and Prognosis," *Journal of the American Academy of Dermatology* 64, no. 6 (2011): 1033–1034, <https://doi.org/10.1016/j.jaad.2010.09.776>.
197. A. Laamanen, T. Salmi, H. Huhtala, et al., "Gastrointestinal Symptoms at Diagnosis and During Long-Term Gluten-Free Diet Treatment in Dermatitis Herpetiformis Patients," *Journal of the European Academy of Dermatology and Venereology* 38, no. 1 (2024): e91–e93, <https://doi.org/10.1111/JDV.19443>.
198. T. Reunala, K. Hervonen, and T. Salmi, "Dermatitis Herpetiformis: An Update on Diagnosis and Management," *American Journal of Clinical Dermatology* 22, no. 3 (2021): 329–338, <https://doi.org/10.1007/S40257-020-00584-2>.
199. T. Li, Y. Feng, C. Wang, et al., "Causal Relationships Between Autoimmune Diseases and Celiac Disease: A Mendelian Randomization Analysis," *Biotechnology & Genetic Engineering Reviews* 40, no. 4 (2023): 4611–4626, <https://doi.org/10.1080/02648725.2023.2215039>.
200. P. Acharya and M. Mathur, "Association Between Psoriasis and Celiac Disease: A Systematic Review and Meta-Analysis," *Journal of the American Academy of Dermatology* 82, no. 6 (2020): 1376–1385, <https://doi.org/10.1016/J.JAAD.2019.11.039>.
201. T. van Gils, H. S. Brand, N. K. H. de Boer, C. J. J. Mulder, and G. Bouma, "Gastrointestinal Diseases and Their oro-Dental Manifestations: Part 3: Coeliac Disease," *British Dental Journal* 222, no. 2 (2017): 126–129, <https://doi.org/10.1038/sj.bdj.2017.80>.
202. A. D. Inchingolo, G. Dipalma, F. Viapiano, et al., "Celiac Disease-Related Enamel Defects: A Systematic Review," *Journal of Clinical Medicine* 13, no. 5 (2024): 1382, <https://doi.org/10.3390/JCM13051382>.
203. M. Rashid, M. Zarkadas, A. Anca, and H. Limeback, "Oral Manifestations of Celiac Disease: A Clinical Guide for Dentists," *Journal of Michigan Dental Association* 93, no. 10 (2011): 42–46, <https://pubmed.ncbi.nlm.nih.gov/21507289/>.
204. F. Zingone, J. C. Bai, C. Cellier, and J. F. Ludvigsson, "Celiac Disease-Related Conditions: Who to Test?," *Gastroenterology* 167, no. 1 (March 2024): 64–78, <https://doi.org/10.1053/J.GASTRO.2024.02.044>.
205. B. M. Skjellerudsveen, R. Omdal, A. K. Hetta, et al., "Fatigue: A Frequent and Biologically Based Phenomenon in Newly Diagnosed Celiac Disease," *Scientific Reports* 12, no. 1 (2022): 7281, <https://doi.org/10.1038/S41598-022-11802-8>.
206. A. Rubio-Tapia and J. A. Murray, "The Liver and Celiac Disease," *Clinics in Liver Disease* 23, no. 2 (2019): 167–176, <https://doi.org/10.1016/j.cld.2018.12.001>.
207. A. Seidita, F. Latteri, M. Pistone, et al., "Celiac Disease and Liver Damage: The Gut-Liver Axis Strikes Back (Again)? A Retrospective Analysis in the Light of a Literature Review," *Nutrients* 17, no. 1 (2024): 85, <https://doi.org/10.3390/NU17010085>.
208. M. Aggarwal, R. Garg, P. Kumar, et al., "Bi-directional Relationship Between Celiac Disease and Liver Chemistries: A Systematic

- Review and Meta-Analysis,” *Digestive Diseases and Sciences* 68, no. 4 (2023): 1369–1380, <https://doi.org/10.1007/S10620-022-07663-W>.
209. R. Nurmi, I. Korponay-Szabó, K. Laurila, et al., “Celiac Disease-Type Tissue Transglutaminase Autoantibody Deposits in Kidney Biopsies of Patients With IgA Nephropathy,” *Nutrients* 13, no. 5 (2021): 1594, <https://doi.org/10.3390/NU13051594>.
210. I. Habura, K. Fiedorowicz, A. Woźniak, I. Idasiak-Piechocka, P. Kosikowski, and A. Oko, “IgA Nephropathy Associated With Celiac Disease,” *Cent J Immunol* 44, no. 1 (2019): 106–108, <https://doi.org/10.5114/CEJI.2019.84021>.
211. K. E. Evans, J. S. Leeds, S. Morley, and D. S. Sanders, “Pancreatic Insufficiency in Adult Celiac Disease: Do Patients Require Long-Term Enzyme Supplementation?,” *Digestive Diseases and Sciences* 55, no. 10 (2010): 2999–3004, <https://doi.org/10.1007/S10620-010-1261-Y>.
212. C. Jiang, J. A. J. S. Barkin, and J. A. J. S. Barkin, “Exocrine Pancreatic Insufficiency Is Common in Celiac Disease: A Systematic Review and Meta-Analysis,” *Digestive Diseases and Sciences* 68, no. 8 (2023): 3421–3427, <https://doi.org/10.1007/S10620-023-07965-7>.
213. M. Vujasinovic, N. Blazevic, P. Maisonneuve, et al., “Pancreatic Exocrine Insufficiency Is Not Uncommon in Celiac Disease: A Systematic Review and Meta-Analysis, 2025,” <https://doi.org/10.1002/UEG2.70076>.
214. M. Alkhayyat, M. A. Saleh, M. Abureesh, et al., “The Risk of Acute and Chronic Pancreatitis in Celiac Disease,” *Digestive Diseases and Sciences* 66, no. 8 (2021): 2691–2699, <https://doi.org/10.1007/S10620-020-06546-2>.
215. J. E. Dominguez-Muñoz, M. Vujasinovic, D. de la Iglesia, et al., “European Guidelines for the Diagnosis and Treatment of Pancreatic Exocrine Insufficiency: UEG, EPC, EDS, ESPEN, ESPGHAN, ESDO, and ESPCG Evidence-Based Recommendations,” *United European Gastroenterology Journal* 13, no. 1 (2025): 125–172, <https://doi.org/10.1002/UEG2.12674>.
216. C. L. Seiler, M. Kiflen, J. P. Stefanolo, et al., “Probiotics for Celiac Disease: A Systematic Review and Meta-Analysis of Randomized Controlled Trials,” *American Journal of Gastroenterology* 115, no. 10 (2020): 1584–1595, <https://doi.org/10.14309/AJG.0000000000000749>.
217. M. Mozafarybazargany, M. Khonsari, L. Sokoty, H. S. Ejtahed, and M. Qorbani, “The Effects of Probiotics on Gastrointestinal Symptoms and Microbiota in Patients With Celiac Disease: A Systematic Review and Meta-Analysis on Clinical Trials,” *Clinical and Experimental Medicine* 23, no. 6 (2023): 2773–2788, <https://doi.org/10.1007/S10238-022-00987-X>.
218. A. Ciccone, D. Gabrieli, R. Cardinale, et al., “Metabolic Alterations in Celiac Disease Occurring After Following a Gluten-Free Diet,” *Digestion* 100, no. 4 (2018): 1–7, <https://doi.org/10.1159/000495749>.
219. N. Aggarwal, A. Agarwal, H. Alarouri, et al., “Patients With Celiac Disease Have High Prevalence of Fatty Liver and Metabolic Syndrome,” *Digestive Diseases and Sciences* 69, no. 8 (2024): 3029–3042, <https://doi.org/10.1007/S10620-024-08426-5>.
220. R. Tortora, P. Capone, G. De Stefano, et al., “Metabolic Syndrome in Patients With Celiac Disease on a Gluten-Free Diet,” *Alimentary Pharmacology & Therapeutics* 41, no. 4 (2015): 352–359, <https://doi.org/10.1111/apt.13062>.
221. J. J. Noubiap, J. R. Nansseu, E. Lontchi-Yimagou, et al., “Geographic Distribution of Metabolic Syndrome and Its Components in the General Adult Population: A Meta-Analysis of Global Data From 28 Million Individuals,” *Diabetes Research and Clinical Practice* 188 (2022): 109924, <https://doi.org/10.1016/j.diabres.2022.109924>.
222. G. D. Cazac, B. M. Mihai, G. Ștefănescu, et al., “Celiac Disease, Gluten-Free Diet and Metabolic Dysfunction-Associated Steatotic Liver Disease,” *Nutrients* 16, no. 13 (2024): 2008, <https://doi.org/10.3390/NU16132008>.
223. A. Akinsanya and I. A. González, “Liver Manifestation of Patients With Celiac Disease: A Single Center Experience,” *Annals of Diagnostic Pathology* 71 (2024): 152327, <https://doi.org/10.1016/j.anndiagpath.2024.152327>.
224. C. Roderburg, S. Loosen, K. Kostev, M. Demir, M. S. Joerdens, and T. Luedde, “Nonalcoholic Fatty Liver Disease Is Associated With a Higher Incidence of Coeliac Disease,” *European Journal of Gastroenterology and Hepatology* 34, no. 3 (2022): 328–331, <https://doi.org/10.1097/MEG.0000000000002234>.
225. A. Armandi, H. Bepaljko, A. Mang, et al., “Short-Term Reduction of Dietary Gluten Improves metabolic-dysfunction Associated Steatotic Liver Disease: A Randomised, Controlled Proof-of-Concept Study,” *Alimentary Pharmacology & Therapeutics* 59, no. 10 (2024): 1212–1222, <https://doi.org/10.1111/APT.17941>.
226. G. Saccone, V. Berghella, L. Sarno, et al., “Celiac Disease and Obstetric Complications: A Systematic Review and Metaanalysis,” *American Journal of Obstetrics and Gynecology* 214, no. 2 (2016): 225–234, <https://doi.org/10.1016/J.AJOG.2015.09.080>.
227. C. Tersigni, R. Castellani, C. De waure, et al., “Celiac Disease and Reproductive Disorders: Meta-Analysis of Epidemiologic Associations and Potential Pathogenic Mechanisms,” *Human Reproduction Update* 20, no. 4 (2014): 582–593, <https://doi.org/10.1093/humupd/dmu007>.
228. A. V. Stazi and A. Mantovani, “A Risk Factor for Female Fertility and Pregnancy: Celiac Disease,” *Gynecological Endocrinology* 14, no. 6 (2000): 454–463, <https://doi.org/10.3109/09513590009167719>.
229. A. Tursi, G. Giorgetti, G. Brandimarte, and W. Elisei, “Effect of Gluten-Free Diet on Pregnancy Outcome in Celiac Disease Patients With Recurrent Miscarriages,” *Digestive Diseases and Sciences* 53, no. 11 (2008): 2925–2928, <https://doi.org/10.1007/S10620-008-0242-X>.
230. F. Mecacci, S. Biagioni, S. Ottanelli, et al., “Nutrition in Pregnancy and Lactation: How a Healthy Infant Is Born,” *J Pediatr Neonatal Indiv Med* 4, no. 2 (2015): e040236, <https://doi.org/10.7363/040236>.
231. D. Alecsandru, N. López-Palacios, M. Castaño, P. Aparicio, J. A. García-Velasco, and C. Núñez, “Exploring Undiagnosed Celiac Disease in Women With Recurrent Reproductive Failure: The Gluten-Free Diet Could Improve Reproductive Outcomes,” *American Journal of Reproductive Immunology* 83, no. 2 (2020): e13209, <https://doi.org/10.1111/aji.13209>.
232. M. Peshevska-Sekulovska, M. Gulinac, R. Rangelov, D. Docheva, T. Velikova, and M. Sekulovski, “Navigating the Challenges of Gluten Enteropathy and Infertility: The Role of Celiac-Related Antibodies and Dietary Changes,” *Antibodies* 12, no. 4 (2023): 79, <https://doi.org/10.3390/ANTIB12040079>.
233. R. Tortora, F. Zingone, A. Rispo, et al., “Coeliac Disease in the Elderly in a Tertiary Centre,” *Scandinavian Journal of Gastroenterology* 51, no. 10 (2016): 1179–1183, <https://doi.org/10.1080/00365521.2016.1186222>.
234. A. Vilppula, K. Kaukinen, L. Luostarinen, et al., “Clinical Benefit of Gluten-Free Diet in Screen-Detected Older Celiac Disease Patients,” *BMC Gastroenterology* 11, no. 1 (2011): 136, <https://doi.org/10.1186/1471-230X-11-136>.
235. P. Collin, A. Vilppula, L. Luostarinen, G. K. T. Holmes, and K. Kaukinen, “Review Article: Coeliac Disease in Later Life Must Not Be Missed,” *Alimentary Pharmacology & Therapeutics* 47, no. 5 (2018): 563–572, <https://doi.org/10.1111/APT.14490>.
236. H. J. Freeman, “Adult Celiac Disease in the Elderly,” *World Journal of Gastroenterology* 14, no. 45 (2008): 6911–6914, <https://doi.org/10.3748/wjg.14.6911>.
237. S. Casella, B. Zanini, F. Lanzarotto, V. Villanacci, C. Ricci, and A. Lanzini, “Celiac Disease in Elderly Adults: Clinical, Serological, and Histological Characteristics and the Effect of a Gluten-Free Diet,” *Journal of the American Geriatrics Society* 60, no. 6 (2012): 1064–1069, <https://doi.org/10.1111/J.1532-5415.2012.03997.X>.

238. A. Rubio-Tapia, C. T. Van Dyke, B. D. Lahr, et al., "Predictors of Family Risk for Celiac Disease: A Population-based Study," *Clinical Gastroenterology and Hepatology* 6, no. 9 (2008): 983–987, <https://doi.org/10.1016/j.cgh.2008.04.008>.
239. L. Book, J. J. Zone, and S. L. Neuhausen, "Prevalence of Celiac Disease Among Relatives of Sib Pairs With Celiac Disease in U.S. Families," *American Journal of Gastroenterology* 98, no. 2 (2003): 377–381, <https://doi.org/10.1111/j.1572-0241.2003.07238.x>.
240. P. Singh, S. Arora, S. Lal, T. A. Strand, and G. K. Makharia, "Risk of Celiac Disease in the First- and Second-Degree Relatives of Patients With Celiac Disease: A Systematic Review and Meta-Analysis," *American Journal of Gastroenterology* 110, no. 11 (2015): 1539–1548, <https://doi.org/10.1038/AJG.2015.296>.
241. S. Husby, S. Koletzko, I. Korponay-Szabó, et al., "European Society Paediatric Gastroenterology, Hepatology and Nutrition Guidelines for Diagnosing Coeliac Disease 2020," *Journal of Pediatric Gastroenterology and Nutrition* 70, no. 1 (2020): 141–156, <https://doi.org/10.1097/MPG.0000000000002497>.
242. F. Megiorni and A. Pizzuti, "HLA-DQA1 and HLA-DQB1 in Celiac Disease Predisposition: Practical Implications of the HLA Molecular Typing," *Journal of Biomedical Science* 19, no. 1 (2012): 88, <https://doi.org/10.1186/1423-0127-19-88>.
243. L. M. Sollid, "The Roles of MHC Class II Genes and Post-Translational Modification in Celiac Disease," *Immunogenetics* 69, no. 8–9 (2017): 605–616, <https://doi.org/10.1007/S00251-017-0985-7>.
244. S. Paavola, K. Lindfors, L. Kivelä, et al., "Presence of High-Risk HLA Genotype Is the Most Important Individual Risk Factor for Coeliac Disease Among at-Risk Relatives," *Alimentary Pharmacology & Therapeutics* 54, no. 6 (2021): 805–813, <https://doi.org/10.1111/APT.16534>.
245. C. R. Meijer, R. Auricchio, H. Putter, et al., "Prediction Models for Celiac Disease Development in Children From High-Risk Families: Data From the PreventCD Cohort," *Gastroenterology* 163, no. 2 (2022): 426–436, <https://doi.org/10.1053/j.gastro.2022.04.030>.
246. L. Nisticò, C. Fagnani, I. Coto, et al., "Concordance, Disease Progression, and Heritability of Coeliac Disease in Italian Twins," *Gut* 55, no. 6 (2006): 803–808, <https://doi.org/10.1136/gut.2005.083964>.
247. C. Catassi, S. Gatti, and A. Fasano, "The New Epidemiology of Celiac Disease," supplement, *Journal of Pediatric Gastroenterology and Nutrition* 59, no. S1 (2014): S7–S9, <https://doi.org/10.1097/01.mpg.0000450393.23156.59>.
248. P. Fallahi, S. M. Ferrari, I. Ruffilli, et al., "The Association of Other Autoimmune Diseases in Patients With Autoimmune Thyroiditis: Review of the Literature and Report of a Large Series of Patients," *Autoimmunity Reviews* 15, no. 12 (2016): 1125–1128, <https://doi.org/10.1016/J.AUTREV.2016.09.009>.
249. A. Dhali, R. Maity, H. R. Bharadwaj, S. H. Ali, M. H. Shah, and D. S. Sanders, "Analyzing the Landscape of Coeliac Crisis in Adult and Paediatric Populations: A Systematic Review and Meta-Analysis," *Digestive and Liver Disease* 57, no. 6 (2025): 1149–1159, <https://doi.org/10.1016/j.dld.2025.02.011>.
250. D. V. Balaban, A. Dima, C. Jurcut, A. Popp, and M. Jinga, "Celiac Crisis, a Rare Occurrence in Adult Celiac Disease: A Systematic Review," *World Journal of Clinical Cases* 7, no. 3 (2019): 311–319, <https://doi.org/10.12998/WJCC.V7.I3.311>.
251. S. Jamma, A. Rubio-Tapia, C. P. Kelly, et al., "Celiac Crisis Is a Rare But Serious Complication of Celiac Disease in Adults," *Clinical Gastroenterology and Hepatology* 8, no. 7 (2010): 587–590, <https://doi.org/10.1016/j.cgh.2010.04.009>.
252. M. Guarino, E. Gambuti, F. Alfano, et al., "Life-Threatening Onset of Coeliac Disease: A Case Report and Literature Review," *BMJ open Gastroenterol* 7, no. 1 (2020): e000406, <https://doi.org/10.1136/BMJGAST-2020-000406>.
253. M. Gupta, C. Graham, and S. Gupta, "Immune Checkpoint Inhibitor-Associated Celiac Disease: A Retrospective Analysis and Literature Review," *Dis (Basel, Switzerland)* 12, no. 12 (2024): 315, <https://doi.org/10.3390/DISEASES12120315>.
254. Y. R. Badran, A. Shih, D. Leet, et al., "Immune Checkpoint inhibitor-associated Celiac Disease," *Journal for Immunotherapy of Cancer* 8, no. 1 (2020): e000958, <https://doi.org/10.1136/jitc-2020-000958>.
255. S. R. Del, U. Volta, V. Lougaris, et al., "Histological Features of Celiac-Disease-Like Conditions Related to Immune Checkpoint Inhibitors Therapy: A Signal to Keep in Mind for Pathologists," *Diagnostics* 12, no. 2 (2022), <https://doi.org/10.3390/DIAGNOSTICS12020395>.
256. J. Leblanc, S. Hoibian, A. Boucraut, et al., "Celiac Disease After Administration of Immune Checkpoint Inhibitors: A Case Report," *Frontiers in Immunology* 12 (2021): 799666, <https://doi.org/10.3389/FIMMU.2021.799666>.
257. A. Schieppatti, A. Premoli, S. Maimaris, et al., "Small Bowel Villous Atrophy Due to Immune-Checkpoint Inhibitors: Report of Two Cases and Literature Review," *Drugs In Context* 11 (2022): 1–12, <https://doi.org/10.7573/DIC.2022-6-3>.
258. C. P. Kelly, J. A. Murray, D. A. Leffler, et al., "TAK-101 Nanoparticles Induce Gluten-Specific Tolerance in Celiac Disease: A Randomized, Double-Blind, Placebo-Controlled Study," *Gastroenterology* 161, no. 1 (2021): 66–80.e8, <https://doi.org/10.1053/J.GASTRO.2021.03.014>.
259. D. Schuppan, M. Mäki, K. E. A. Lundin, et al., "A Randomized Trial of a Transglutaminase 2 Inhibitor for Celiac Disease," *New England Journal of Medicine* 385, no. 1 (2021): 35–45, <https://doi.org/10.1056/NEJMOA2032441>.
260. P. Hindryckx, B. G. Levesque, T. Holvoet, et al., "Disease Activity Indices in Coeliac Disease: Systematic Review and Recommendations for Clinical Trials," *Gut* 67, no. 1 (2018): 61–69, <https://doi.org/10.1136/GUTJNL-2016-312762>.
261. W. J. Canestaro, T. C. Edwards, and D. L. Patrick, "Systematic Review: Patient-Reported Outcome Measures in Coeliac Disease for Regulatory Submissions," *Alimentary Pharmacology & Therapeutics* 44, no. 4 (2016): 313–331, <https://doi.org/10.1111/apt.13703>.
262. J. F. Ludvigsson, C. Ciacci, P. H. R. Green, et al., "Outcome Measures in Coeliac Disease Trials: The Tampere Recommendations," *Gut* 67, no. 8 (2018): 1410–1424, <https://doi.org/10.1136/GUTJNL-2017-314853>.
263. J. König, S. Holster, M. J. Bruins, and R. J. Brummer, "Randomized Clinical Trial: Effective Gluten Degradation by Aspergillus Niger-Derived Enzyme in a Complex Meal Setting," *Scientific Reports* 7, no. 1 (2017): 13100, <https://doi.org/10.1038/s41598-017-13587-7>.
264. Y. E. Dunaevsky, V. F. Tereshchenkova, M. A. Belozersky, I. Y. Filippova, B. Oppert, and E. N. Elpidina, "Review Effective Degradation of Gluten and Its Fragments by Gluten-Specific Peptidases: A Review on Application for the Treatment of Patients With Gluten Sensitivity," *Pharmaceutics* 13, no. 10 (2021): 1603, <https://doi.org/10.3390/PHARMACEUTICS13101603>.
265. B. Lebwohl, J. A. Murray, A. Rubio-Tapia, P. H. R. L. J. Green, and J. F. Ludvigsson, "Predictors of Persistent Villous Atrophy in Coeliac Disease: A Population-Based Study," *Alimentary Pharmacology & Therapeutics* 39, no. 5 (2014): 488–495, <https://doi.org/10.1111/apt.12621>.
266. P. J. Wahab, J. W. R. Meijer, and C. J. J. Mulder, "Histologic follow-up of People With Celiac Disease on a Gluten-Free Diet: Slow and Incomplete Recovery," *American Journal of Clinical Pathology* 118, no. 3 (2002): 459–463, <https://doi.org/10.1309/EVXT-851X-WHLC-RLX9>.
267. W. Dickey, D. F. Hughes, and S. A. McMillan, "Disappearance of Endomysial Antibodies in Treated Celiac Disease Does Not Indicate Histological Recovery," *American Journal of Gastroenterology* 95, no. 3 (2000): 712–714, <https://doi.org/10.1111/j.1572-0241.2000.01838.x>.

268. J. Lexner, H. Hjortswang, R. Ekesbo, and K. Sjöberg, "Well-Being and Dietary Adherence in Patients With Coeliac Disease Depending on Follow-Up," *Scandinavian Journal of Gastroenterology* 56, no. 4 (2021): 382–390, <https://doi.org/10.1080/00365521.2021.1889024>.
269. C. J. J. Mulder, L. Elli, B. Lebwohl, et al., "Follow-Up of Celiac Disease in Adults: "When, What, Who, and Where,"" *Nutrients* 15, no. 9 (2023): 2048, <https://doi.org/10.3390/NU15092048>.
270. J. J. Hughey, B. K. Ray, A. R. Lee, K. N. Voorhees, C. P. Kelly, and D. Schuppan, "Self-Reported Dietary Adherence, Disease-specific Symptoms, and Quality of Life Are Associated With Healthcare Provider Follow-Up in Celiac Disease," *BMC Gastroenterology* 17, no. 1 (2017): 156, <https://doi.org/10.1186/S12876-017-0713-7>.
271. R. Mandile, M. Maglio, C. Mosca, et al., "Mucosal Healing in Celiac Disease: Villous Architecture and Immunohistochemical Features in Children on a Long-Term Gluten Free Diet," *Nutrients* 14, no. 18 (2022): 3696, <https://doi.org/10.3390/NU14183696>.
272. P. Hære, O. Høie, T. Schulz, I. Schönhardt, M. Raki, and K. E. A. Lundin, "Long-Term Mucosal Recovery and Healing in Celiac Disease Is the Rule - Not the Exception," *Scandinavian Journal of Gastroenterology* 51, no. 12 (2020): 1439–1446, <https://doi.org/10.1080/00365521.2016.1218540>.
273. A. Rej, N. Trott, M. Kurien, et al., "Is Peer Support in Group Clinics as Effective as Traditional Individual Appointments? The First Study in Patients With Celiac Disease," *Clinical and Translational Gastroenterology* 11, no. 1 (2020): e00121, <https://doi.org/10.14309/CTG.000000000000121>.
274. S. Vriezinga, A. Borghorst, E. van den Akker-van Marle, et al., "E-Healthcare for Celiac Disease-A Multicenter Randomized Controlled Trial," *Jornal de Pediatria* 195 (2018): 154–160.e7, <https://doi.org/10.1016/J.JPEDS.2017.10.027>.
275. G. Perez-Junkera, M. Vázquez-Polo, F. J. Eizagirre, et al., "Application of a Platform for Gluten-Free Diet Evaluation and Dietary Advice: From Theory to Practice," *Sensors* 22, no. 3 (2022): 732, <https://doi.org/10.3390/s22030732>.
276. Y. Wang, B. Chen, E. J. Ciaccio, et al., "Celiac Disease and the Risk of Cardiovascular Diseases," *International Journal of Molecular Sciences* 24, no. 12 (2023): 9974, <https://doi.org/10.3390/IJMS24129974>.
277. L. Emilsson, B. Lebwohl, J. Sundström, and J. F. Ludvigsson, "Cardiovascular Disease in Patients With Coeliac Disease: A Systematic Review and Meta-Analysis," *Digestive and Liver Disease* 47, no. 10 (2015): 847–852, <https://doi.org/10.1016/j.dld.2015.06.004>.
278. M. D. E. Potter, S. C. Briennes, M. M. Walker, A. Boyle, and N. J. Talley, "Effect of the Gluten-Free Diet on Cardiovascular Risk Factors in Patients With Coeliac Disease: A Systematic Review," *Journal of Gastroenterology and Hepatology* 33, no. 4 (2018): 781–791, <https://doi.org/10.1111/JGH.14039>.
279. B. Zanini, E. Mazzoncin, F. Lanzarotto, et al., "Impact of Gluten-Free Diet on Cardiovascular Risk Factors: A Retrospective Analysis in a Large Cohort of Coeliac Patients," *Digestive and Liver Disease* 45, no. 10 (2013): 810–815, <https://doi.org/10.1016/j.dld.2013.04.001>.
280. P. Baharvand, M. Hormozi, and A. Aaliehpor, "Comparison of Thyroid Disease Prevalence in Patients With Celiac Disease and Controls," *Gastroenterol Hepatol from bed to bench* 13, no. 1 (2020): 44–49, <https://doi.org/10.22037/ghfbv.v13i1.1743>.
281. E. Sugai, F. Nachman, H. Vázquez, et al., "Dynamics of Celiac Disease-Specific Serology After Initiation of a Gluten-Free Diet and Use in the Assessment of Compliance With Treatment," *Digestive and Liver Disease* 42, no. 5 (2010): 352–358, <https://doi.org/10.1016/J.DLD.2009.07.011>.
282. F. Nachman, E. Sugai, H. Vázquez, et al., "Serological Tests for Celiac Disease as Indicators of long-Term Compliance With the Gluten-Free Diet," *European Journal of Gastroenterology and Hepatology* 23, no. 6 (2011): 473–480, <https://doi.org/10.1097/MEG.0b013e328346e0f1>.
283. B. Zanini, F. Lanzarotto, A. Mora, et al., "Five Year Time Course of Celiac Disease Serology During Gluten Free Diet: Results of a Community Based "CD-Watch" Program," *Digestive and Liver Disease* 42, no. 12 (2010): 865–870, <https://doi.org/10.1016/j.dld.2010.05.009>.
284. L. F. Risnes, H. M. Reims, R. M. Doyle, et al., "Gluten-Free Diet Induces Rapid Changes in Phenotype and Survival Properties of Gluten-Specific T Cells in Celiac Disease," *Gastroenterology* 167, no. 2 (2024): 250–263, <https://doi.org/10.1053/j.gastro.2024.03.027>.
285. K. Vahedi, F. Mascart, J. Y. Mary, et al., "Reliability of Anti-transglutaminase Antibodies as Predictors of Gluten-Free Diet Compliance in Adult Celiac Disease," *American Journal of Gastroenterology* 98, no. 5 (2003): 1079–1087, <https://doi.org/10.1111/j.1572-0241.2003.07284.x>.
286. W. Dickey and D. Hughes, "Disappointing Sensitivity of Endoscopic Markers for Villous Atrophy in a High-Risk Population: Implications for Celiac Disease Diagnosis During Routine Endoscopy," *American Journal of Gastroenterology* 96, no. 7 (2001): 2126–2128, <https://doi.org/10.1111/j.1572-0241.2001.03947.x>.
287. I. R. Korponay-Szabó, I. Dahlbom, K. Laurila, et al., "Elevation of IgG Antibodies Against Tissue Transglutaminase as a Diagnostic Tool for Coeliac Disease in Selective IgA Deficiency," *Gut* 52, no. 11 (2003): 1567–1571, <https://doi.org/10.1136/GUT.52.11.1567>.
288. F. Cataldo, D. Lio, V. Marino, et al., "IgG1 Antiendomysium and IgG Antitissue Transglutaminase (anti-tTG) Antibodies in Coeliac Patients With Selective IgA Deficiency," *Gut* 47, no. 3 (2000): 366–369, <https://doi.org/10.1136/gut.47.3.366>.
289. K. Kaukinen, M. Peraaho, K. Lindfors, et al., "Persistent Small Bowel Mucosal Villous Atrophy Without Symptoms in Coeliac Disease," *Alimentary Pharmacology & Therapeutics* 25, no. 10 (2007): 1237–1245, <https://doi.org/10.1111/j.1365-2036.2007.03311.x>.
290. B. Lebwohl, F. Granath, A. Ekbo, et al., "Mucosal Healing and Mortality in Coeliac Disease," *Alimentary Pharmacology & Therapeutics* 37, no. 3 (2013): 332–339, <https://doi.org/10.1111/apt.12164>.
291. J. M. Hutchinson, N. P. West, G. G. Robins, and P. D. Howdle, "Long-Term Histological follow-up of People With Coeliac Disease in a UK Teaching Hospital," *QJM* 103, no. 7 (2010): 511–517, <https://doi.org/10.1093/qjmed/hcq076>.
292. H. Pekki, K. Kurppa, M. Mäki, et al., "Performing Routine follow-up Biopsy 1 Year After Diagnosis Does Not Affect long-term Outcomes in Coeliac Disease," *Alimentary Pharmacology & Therapeutics* 45, no. 11 (2017): 1459–1468, <https://doi.org/10.1111/apt.14048>.
293. A. Rubio-Tapia, M. W. Rahim, J. A. See, B. D. Lahr, T.-T. Wu, and J. A. Murray, "Mucosal Recovery and Mortality in Adults With Celiac Disease After Treatment With a Gluten-free Diet," *American Journal of Gastroenterology* 105, no. 6 (2010): 1412–1420, <https://doi.org/10.1038/ajg.2010.10>.
294. E. Stasi, I. Marafini, R. Caruso, et al., "Frequency and Cause of Persistent Symptoms in Celiac Disease Patients on a Long-Term Gluten-Free Diet," *Journal of Clinical Gastroenterology* 50, no. 3 (2016): 239–243, <https://doi.org/10.1097/MCG.0000000000000392>.
295. P. Laurikka, T. Salmi, P. Collin, et al., "Gastrointestinal Symptoms in Celiac Disease Patients on a Long-Term Gluten-Free Diet," *Nutrients* 8, no. 7 (2016): 429, <https://doi.org/10.3390/NU8070429>.
296. D. C. Adelman, J. Murray, T.-T. Wu, M. Mäki, P. H. Green, and C. P. Kelly, "Measuring Change in Small Intestinal Histology in Patients With Celiac Disease," *American Journal of Gastroenterology* 113, no. 3 (2018): 339–347, <https://doi.org/10.1038/ajg.2017.480>.
297. S. K. Lee, W. Lo, L. Memeo, H. Rotterdam, and P. H. R. Green, "Duodenal Histology in Patients With Celiac Disease After Treatment With a Gluten-Free Diet," *Gastrointestinal Endoscopy* 57, no. 2 (2003): 187–191, <https://doi.org/10.1067/mge.2003.54>.

298. J. A. Tye-Din, "Review Article: Follow-Up of Coeliac Disease," supplement, *Alimentary Pharmacology & Therapeutics* 56, no. Suppl 1 (2022): S49–S63, <https://doi.org/10.1111/APT.16847>.
299. H. A. Penny, A. Rej, E. M. R. Baggus, et al., "Non-Responsive and Refractory Coeliac Disease: Experience From the NHS England National Centre," *Nutrients* 14, no. 13 (2022): 2776, <https://doi.org/10.3390/NU14132776>.
300. A. Rubio-Tapia, I. D. Hill, C. Semrad, et al., "American College of Gastroenterology Guidelines Update: Diagnosis and Management of Celiac Disease," *American Journal of Gastroenterology* 118, no. 1 (2023): 59–76, <https://doi.org/10.14309/AJG.0000000000002075>.
301. A. Rubio-Tapia and J. A. Murray, "Classification and Management of Refractory Coeliac Disease," *Gut* 59, no. 4 (2010): 547–557, <https://doi.org/10.1136/gut.2009.195131>.
302. D. H. Dewar, S. C. Donnelly, S. D. McLaughlin, M. W. Johnson, H. J. Ellis, and P. J. Ciclitira, "Celiac Disease: Management of Persistent Symptoms in Patients on a Gluten-Free Diet," *World Journal of Gastroenterology* 18, no. 12 (2012): 1348–1356, <https://doi.org/10.3748/WJG.V18.I12.1348>.
303. N. J. Hall, G. P. Rubin, and A. Charnock, "Intentional and Inadvertent Non-Adherence in Adult Coeliac Disease. A Cross-Sectional Survey," *Appetite* 68 (2013): 56–62, <https://doi.org/10.1016/J.APPET.2013.04.016>.
304. M. I. Pinto-Sanchez, C. L. Seiler, N. Santesso, et al., "Association Between Inflammatory Bowel Diseases and Celiac Disease: A Systematic Review and Meta-Analysis," *Gastroenterology* 159, no. 3 (2020): 884–903. e31, <https://doi.org/10.1053/j.gastro.2020.05.016>.
305. P. Vijayvargiya, D. Gonzalez Izundegui, G. Calderon, et al., "Increased Fecal Bile Acid Excretion in a Significant Subset of Patients With Other Inflammatory Diarrheal Diseases," *Digestive Diseases and Sciences* 67, no. 6 (2022): 2413–2419, <https://doi.org/10.1007/S10620-021-06993-5>.
306. R. R. Wenzel and C. Datz, "Association of Sprue-Like Enteropathy and Angiotensin receptor-1 Antagonists," *Wiener Klinische Wochenschrift* 131, no. 19–20 (2019): 493–501, <https://doi.org/10.1007/S00508-019-01539-2>.
307. N. Burbure, B. Lebowohl, C. Arguelles-Grande, P. H. R. Green, G. Bhagat, and S. Lagana, "Olmesartan-Associated Sprue-Like Enteropathy: A Systematic Review With Emphasis on Histopathology," *Human Pathology* 50 (2016): 127–134, <https://doi.org/10.1016/j.humpath.2015.12.001>.
308. T. van Gils, P. Nijeboer, L. I. Overbeek, et al., "Risks for Lymphoma and Gastrointestinal Carcinoma in Patients With Newly Diagnosed Adult-onset Celiac Disease: Consequences for Follow-Up: Celiac Disease, Lymphoma and GI Carcinoma," *United European Gastroenterology Journal* 6, no. 10 (2018): 1485–1495, <https://doi.org/10.1177/2050640618800540>.
309. A. Al-Toma, W. H. M. Verbeek, and C. J. J. Mulder, "Update on the Management of Refractory Coeliac Disease," *Journal of Gastrointestinal and Liver Diseases* 16, no. 1 (2007): 57–63, <https://doi.org/10.1159/000128672>.
310. M. L. Sanford and A. K. Nagel, "A Review of Current Evidence of Olmesartan Medoxomil Mimicking Symptoms of Celiac Disease," *Journal of Pharmacy Practice* 28, no. 2 (2015): 189–192, <https://doi.org/10.1177/0897190014527320>.
311. A. Schieppatti, D. S. Sanders, P. Baiardi, et al., "Nomenclature and Diagnosis of Seronegative Coeliac Disease and Chronic Non-Coeliac Enteropathies in Adults: The Paris consensus," *Gut* 71, no. 11 (2022): 2218–2225, <https://doi.org/10.1136/GUTJNL-2021-326645>.
312. A. F. Costa, E. Sugai, M. De La Paz Temprano, et al., "Gluten Immunogenic Peptide Excretion Detects Dietary Transgressions in Treated Celiac Disease Patients," *World Journal of Gastroenterology* 25, no. 11 (2019): 1409–1420, <https://doi.org/10.3748/WJG.V25.I11.1409>.
313. J. P. Stefanolo, M. Temprano, P. de la, et al., "Comparison of Weekly Gluten Immunogenic Peptide Measurement and Conventional Tools to Assess Adherence to the Gluten-Free Diet in Celiac Disease: An Observational Prospective Study," *American Journal of Clinical Nutrition* 118, no. 6 (2023): 1106–1112, <https://doi.org/10.1016/J.AJCNUT.2023.10.001>.
314. J. A. Silvester, I. Comino, C. P. Kelly, et al., "Most Patients With Celiac Disease on Gluten-Free Diets Consume Measurable Amounts of Gluten," *Gastroenterology* 158, no. 5 (2020): 1497–1499.e1, <https://doi.org/10.1053/J.GASTRO.2019.12.016>.
315. F. Fernández-Bañares, B. Beltrán, A. Salas, et al., "Persistent Villous Atrophy in De Novo Adult Patients With Celiac Disease and Strict Control of Gluten-Free Diet Adherence: A Multicenter Prospective Study (CADER Study)," *American Journal of Gastroenterology* 116, no. 5 (2021): 1036–1043, <https://doi.org/10.14309/AJG.0000000000001139>.
316. G. Malamut, P. Afchain, V. Verkarre, et al., "Presentation and Long-Term Follow-up of Refractory Celiac Disease: Comparison of Type I With Type II," *Gastroenterology* 136, no. 1 (2009): 81–90, <https://doi.org/10.1053/J.GASTRO.2008.09.069>.
317. C. Cellier, N. Patey, L. Mauvieux, et al., "Abnormal Intestinal Intraepithelial Lymphocytes in Refractory Sprue," *Gastroenterology* 114, no. 3 (1998): 471–481, [https://doi.org/10.1016/S0016-5085\(98\)70530-X](https://doi.org/10.1016/S0016-5085(98)70530-X).
318. P. H. R. Green, S. Paski, C. W. Ko, and A. Rubio-Tapia, "AGA Clinical Practice Update on Management of Refractory Celiac Disease: Expert Review," *Gastroenterology* 163, no. 5 (2022): 1461–1469, <https://doi.org/10.1053/J.GASTRO.2022.07.086>.
319. T. Ilus, K. Kaukinen, L. J. Virta, et al., "Refractory Coeliac Disease in a Country With a High Prevalence of Clinically-Diagnosed Coeliac Disease," *Alimentary Pharmacology & Therapeutics* 39, no. 4 (2014): 418–425, <https://doi.org/10.1111/APT.12606>.
320. G. Malamut and C. Cellier, "Refractory Celiac Disease: Epidemiology and Clinical Manifestations," *Digestive Diseases* 33, no. 2 (2015): 221–226, <https://doi.org/10.1159/000369519>.
321. A. Al-Toma, W. H. M. Verbeek, and C. J. J. Mulder, "The Management of Complicated Celiac Disease," *Digestive Diseases* 25, no. 3 (2007): 230–236, <https://doi.org/10.1159/000103891>.
322. F. Branchi, J. J. Wiese, C. Heldt, et al., "The Combination of Clinical Parameters and Immunophenotyping of Intraepithelial Lymphocytes Allows to Assess Disease Severity in Refractory Celiac Disease," *Digestive and Liver Disease* 54, no. 12 (2022): 1649–1656, <https://doi.org/10.1016/J.DLD.2022.06.024>.
323. G. Malamut, B. Meresse, C. Cellier, and N. Cerf-Bensussan, "Refractory Celiac Disease: From Bench to Bedside," *Seminars in Immunopathology* 34, no. 4 (2012): 601–613, <https://doi.org/10.1007/s00281-012-0322-z>.
324. J. M. W. Van De Water, P. Nijeboer, L. R. De Baaij, et al., "Surgery in (Pre)Malignant Celiac Disease," *World Journal of Gastroenterology* 21, no. 43 (2015): 12403–12409, <https://doi.org/10.3748/WJG.V21.I43.12403>.
325. S. Hussein, T. Gindin, S. M. Lagana, et al., "Clonal T Cell Receptor Gene Rearrangements in Coeliac Disease: Implications for Diagnosing Refractory Coeliac Disease," *Journal of Clinical Pathology* 71, no. 9 (2018): 825–831, <https://doi.org/10.1136/JCLINPATH-2018-205023>.
326. C. García-Hoz, L. Crespo, N. Lopez, et al., "The Intracellular Intensity of CD3 on Aberrant Intraepithelial Lymphocytes Is a Prognostic Factor of the Progression to Overt Lymphoma in Refractory Celiac Disease Type II (Pre-Enteropathy-Associated T Cell Lymphoma)," *Digestive Diseases* 38, no. 6 (2020): 490–499, <https://doi.org/10.1159/000506305>.
327. C. Cellier, E. Delabesse, C. Helmer, et al., "Refractory Sprue, Coeliac Disease, and enteropathy-associated T-cell Lymphoma. French Coeliac Disease Study Group," *Lancet (London, England)* 356, no. 9225 (2000): 203–208, [https://doi.org/10.1016/S0140-6736\(00\)02481-8](https://doi.org/10.1016/S0140-6736(00)02481-8).

328. L. Pironi, D. Konrad, C. Brandt, et al., "Clinical Classification of Adult Patients With Chronic Intestinal Failure due to Benign Disease: An International Multicenter Cross-Sectional," *survey* 37, no. 2 (2018): 728–738, <https://doi.org/10.1016/j.clnu.2017.04.013>.
329. D. G. A. M. Bianchetti, G. S. Amelio, S. A. G. Lava, et al., "D-lactic Acidosis in Humans: Systematic Literature Review," *Pediatric Nephrology* 33, no. 4 (2018): 673–681, <https://doi.org/10.1007/S00467-017-3844-8>.
330. W. H. M. Verbeek, M. W. Schreurs, O. J. Visser, B. M. E. von Blomberg, A. Al-Toma, and C. J. J. Mulder, "Novel Approaches in the Management of Refractory Celiac Disease," *Expert Review of Clinical Immunology* 4, no. 2 (2008): 205–219, <https://doi.org/10.1586/1744666X.4.2.205>.
331. S. Daum, C. Cellier, and C. J. J. Mulder, "Refractory Coeliac Disease," *Best Practice & Research Clinical Gastroenterology* 19, no. 3 (2005): 413–424, <https://doi.org/10.1016/j.bpg.2005.02.001>.
332. J. Pastré, K. Juvin, G. Malamut, C. Derrieux, C. Cellier, and D. Israël-Biet, "Phenotypically Aberrant Clonal T Cells in the Lungs of Patients With Type II Refractory Celiac Disease," *Blood* 123, no. 23 (2014): 3674–3675, <https://doi.org/10.1182/BLOOD-2014-04-566513>.
333. A. A. Maguire, J. K. Greenson, G. Y. Lauwers, et al., "Collagenous Sprue: A Clinicopathologic Study of 12 Cases," *American Journal of Surgical Pathology* 33, no. 10 (2009): 1440–1449, <https://doi.org/10.1097/PAS.0B013E3181AE2545>.
334. V. L. Kung, T. C. Liu, and C. Ma, "Collagenous Enteritis is Unlikely a Form of Aggressive Celiac Disease Despite Sharing HLA-DQ2/DQ8 Genotypes," *American Journal of Surgical Pathology* 42, no. 4 (2018): 545–552, <https://doi.org/10.1097/PAS.0000000000001022>.
335. E. Vakiani, C. Arguelles-Grande, M. M. Mansukhani, et al., "Collagenous Sprue is Not Always Associated With Dismal Outcomes: A Clinicopathological Study of 19 Patients," *Modern Pathology* 23, no. 1 (2010): 12–26, <https://doi.org/10.1038/MODPATHOL.2009.151>.
336. M. Barret, G. Malamut, G. Rahmi, et al., "Diagnostic Yield of Capsule Endoscopy in Refractory Celiac Disease," *American Journal of Gastroenterology* 107, no. 10 (2012): 1546–1553, <https://doi.org/10.1038/ajg.2012.199>.
337. M. Hadithi, A. Al-toma, J. Oudejans, A. A. A. Van Bodegraven, C. J. C. J. Mulder, and M. Jacobs, "The Value of Double-Balloon Enteroscopy in Patients With Refractory Celiac Disease," *American Journal of Gastroenterology* 102, no. 5 (2007): 987–996, <https://doi.org/10.1111/j.1572-0241.2007.01122.x>.
338. A. Al-Toma, H. Beaumont, J. J. Koornstra, et al., "Motorized Spiral Enteroscopy: Multicenter Prospective Study on Performance and Safety Including in Patients With Surgically-Altered Gastrointestinal Anatomy," *Endoscopy* 54, no. 11 (February 2022): 1034–1042, <https://doi.org/10.1055/a-1783-4802>.
339. W. H. M. Verbeek, M. S. Goerres, B. M. E. von Blomberg, et al., "Flow Cytometric Determination of Aberrant Intra-Epithelial Lymphocytes Predicts T-cell Lymphoma Development More Accurately than T-cell Clonality Analysis in Refractory Celiac Disease," *Clinical Immunology* 126, no. 1 (2008): 48–56, <https://doi.org/10.1016/j.clim.2007.09.002>.
340. R. L. J. van Wanrooij, M. W. J. Schreurs, G. Bouma, et al., "Accurate Classification of RCD Requires Flow Cytometry," *Gut* 59, no. 12 (2010): 1732, <https://doi.org/10.1136/gut.2010.223438>.
341. G. Malamut, B. Meresse, S. Kaltenbach, et al., "Small Intestinal CD4+ T-cell Lymphoma is a Heterogenous Entity With Common Pathology Features," *Clinical Gastroenterology and Hepatology* 12, no. 4 (2014): 599–608.e1, <https://doi.org/10.1016/j.CGH.2013.11.028>.
342. A. Al-Toma, W. H. M. Verbeek, M. Hadithi, B. M. E. von Blomberg, and C. J. J. Mulder, "Survival in Refractory Coeliac Disease and Enteropathy-Associated T-cell Lymphoma: Retrospective Evaluation of single-centre Experience," *Gut* 56, no. 10 (2007): 1373–1378, <https://doi.org/10.1136/gut.2006.114512>.
343. M. Cheminant, J. Bruneau, G. Malamut, et al., "NKp46 is a Diagnostic Biomarker and may be a Therapeutic Target in Gastrointestinal T-cell Lymphoproliferative Diseases: A CELAC Study," *Gut* 68, no. 8 (November 2018): 1396–1405, <https://doi.org/10.1136/gutjnl-2018-317371>.
344. G. Malamut and C. Cellier, "Refractory Celiac Disease," *Gastroenterology Clinics of North America* 48, no. 1 (2019): 137–144, <https://doi.org/10.1016/J.GTC.2018.09.010>.
345. G. Malamut and C. Cellier, "Refractory Coeliac Disease," *Current Opinion in Oncology* 25, no. 5 (2013): 445–451, <https://doi.org/10.1097/01.CCO.0000432526.47228.B6>.
346. A. Al-toma, O. J. Visser, H. M. Van Roessel, et al., "Autologous Hematopoietic Stem Cell Transplantation in Refractory Celiac Disease With Aberrant T Cells," *Blood* 109, no. 5 (2007): 2243–2249, <https://doi.org/10.1182/blood-2006-08-042820>.
347. F. Charbit-Henrion, M. Haas, S. Chaussade, et al., "Genetic Diagnosis Guides Treatment of Autoimmune Enteropathy," *Clinical Gastroenterology and Hepatology* 21, no. 5 (2023): 1368–1371.e2, <https://doi.org/10.1016/J.CGH.2022.07.030>.
348. S. Cording, L. Lhermitte, G. Malamut, et al., "Oncogenetic Landscape of Lymphomagenesis in Coeliac Disease," *Gut* 71, no. 3 (2022): 497–508, <https://doi.org/10.1136/GUTJNL-2020-322935>.
349. G. Malamut, R. El Machhour, N. Montcuquet, et al., "IL-15 Triggers an Antiapoptotic Pathway in Human Intraepithelial Lymphocytes That is a Potential New Target in Celiac Disease-Associated Inflammation and Lymphomagenesis," *Journal of Clinical Investigation* 120, no. 6 (2010): 2131–2143, <https://doi.org/10.1172/JCI41344>.
350. C. Cellier, G. Bouma, T. van Gils, et al., "Safety and Efficacy of AMG 714 in Patients With Type 2 Refractory Coeliac Disease: A Phase 2a, Randomised, Double-Blind, Placebo-Controlled, Parallel-Group Study," *Lancet Gastroenterology & Hepatology* 4, no. 12 (2019): 960–970, [https://doi.org/10.1016/S2468-1253\(19\)30265-1](https://doi.org/10.1016/S2468-1253(19)30265-1).
351. M. Singh, R. H. Y. Louie, J. Samir, et al., "Expanded T Cell Clones With Lymphoma Driver Somatic Mutations Accumulate in Refractory Celiac Disease," *Science Translational Medicine* 17, no. 798 (2025): eadp6812, <https://doi.org/10.1126/SCITRANSLMED.ADP6812>.
352. M. Hoffmann, H. Vogelsang, K. Kletter, G. Zettinig, A. Chott, and M. Raderer, "18F-Fluoro-Deoxy-Glucose Positron Emission Tomography (18F-FDG-PET) for Assessment of Enteropathy-Type T Cell Lymphoma," *Gut* 52, no. 3 (2003): 347–351, <https://doi.org/10.1136/GUT.52.3.347>.
353. T. S. Y. Chan, E. Lee, P.-L. Khong, E. W. C. Tse, and Y.-L. Kwong, "Positron Emission Tomography Computed Tomography Features of Monomorphic Epitheliotropic Intestinal T-Cell Lymphoma," *Hematology* 23, no. 1 (2018): 10–16, <https://doi.org/10.1080/10245332.2017.1335979>.
354. S. J. B. Van Weyenberg, C. J. J. Mulder, and J. H. T. M. Van Waesberghe, "Small Bowel Imaging in Celiac Disease," *Digestive Diseases* 33, no. 2 (2015): 252–259, <https://doi.org/10.1159/000369516>.
355. N. J. Wierdsma, P. Nijeboer, M. A. E. de van der Schueren, M. Berkenpas, A. A. van Bodegraven, and C. J. J. Mulder, "Refractory Celiac Disease and EATL Patients Show Severe Malnutrition and Malabsorption at Diagnosis," *Clinical Nutrition* 35, no. 3 (2016): 685–691, <https://doi.org/10.1016/j.clnu.2015.04.014>.
356. S. S. Mukewar, A. Sharma, A. Rubio-Tapia, T.-T. Wu, B. Jabri, and J. A. Murray, "Open-Capsule Budesonide for Refractory Celiac Disease," *American Journal of Gastroenterology* 112, no. 6 (2017): 959–967, <https://doi.org/10.1038/ajg.2017.71>.
357. D. Saitta, L. M. Henneken, P. Apputhurai, S. L. Chen Yi Mei, and J. A. Tye-Din, "Budesonide Induces Favourable Histologic and Symptomatic Recovery in Patients With Non-responsive and Refractory

- Coeliac Disease When Given in an Open Capsule Format,” *Digestive Diseases and Sciences* 69, no. 7 (2024): 2548–2557, <https://doi.org/10.1007/S10620-024-08436-3>.
358. G. J. Tack, W. H. M. Verbeek, A. Al-Toma, et al., “Evaluation of Cladribine Treatment in Refractory Celiac Disease Type II,” *World Journal of Gastroenterology* 17, no. 4 (2011): 506–513, <https://doi.org/10.3748/wjg.v17.i4.506>.
359. G. J. Tack, M. J. Wondergem, A. Al-Toma, et al., “Auto-SCT in Refractory Celiac Disease Type II Patients Unresponsive to Cladribine Therapy,” *Bone Marrow Transplantation* 46, no. 6 (2011): 840–846, <https://doi.org/10.1038/bmt.2010.199>.
360. A. Al-toma, M. S. Goerres, J. W. R. Meijer, et al., “Cladribine Therapy in Refractory Celiac Disease With Aberrant T Cells,” *Clinical Gastroenterology and Hepatology* 4, no. 11 (2006): 1322–1327, <https://doi.org/10.1016/j.cgh.2006.07.007>.
361. M. S. Goerres, J. W. R. Meijer, P. J. Wahab, et al., “Azathioprine and Prednisone Combination Therapy in Refractory Coeliac Disease,” *Alimentary Pharmacology & Therapeutics* 18, no. 5 (2003): 487–494, <https://doi.org/10.1046/j.1365-2036.2003.01687.x>.
362. T. Dieckman, M. Schumann, H. Beaumont, et al., “Enduring Clinical Remission in Refractory Celiac Disease Type II With Tofacitinib: An Open-Label Clinical Study,” *Clinical Gastroenterology and Hepatology* 22, no. 11 (May 2024): 2334–2336, <https://doi.org/10.1016/J.CGH.2024.05.022>.
363. A. Rubio-Tapia, D. G. Kelly, B. D. Lahr, A. Dogan, T. Wu, and J. A. Murray, “Clinical Staging and Survival in Refractory Celiac Disease: A Single Center Experience,” *Gastroenterology* 136, no. 1 (2009): 99–107, <https://doi.org/10.1053/j.gastro.2008.10.013>.
364. G. J. Tack, D. P. van Asseldonk, R. L. J. van Wanrooij, A. A. van Bodegraven, and C. J. Mulder, “Tioguanine in the Treatment of Refractory Coeliac Disease—A Single Centre Experience,” *Alimentary Pharmacology & Therapeutics* 36, no. 3 (2012): 274–281, <https://doi.org/10.1111/j.1365-2036.2012.05154.x>.
365. H. R. Gillett, I. D. R. Arnott, M. McIntyre, et al., “Successful Infliximab Treatment for Steroid-Refractory Celiac Disease: A Case Report,” *Gastroenterology* 122, no. 3 (2002): 800–805, <https://doi.org/10.1053/gast.2002.31874>.
366. S. M. Turner, M. Moorghen, and C. S. J. Probert, “Refractory Coeliac Disease: Remission With Infliximab and Immunomodulators,” *European Journal of Gastroenterology and Hepatology* 17, no. 6 (2005): 667–669, <https://doi.org/10.1097/00042737-200506000-00012>.
367. A. Al-Toma and H. R. Koene, “Hematopoietic Stem Cell Transplantation in Refractory Celiac Disease: An Overview With Focus on Infectious Complications,” *OBM Transplant* 4, no. 1 (2020): 1–17, <https://doi.org/10.21926/OBM.TRANSPLANT.2001101>.
368. A. Al-Toma, P. Nijeboer, G. Bouma, O. Visser, and C. J. J. Mulder, “Hematopoietic Stem Cell Transplantation for Non-Malignant Gastrointestinal Diseases,” *World Journal of Gastroenterology* 20, no. 46 (2014): 17368–17375, <https://doi.org/10.3748/wjg.v20.i46.17368>.
369. R. Ciccocioppo, A. Gallia, M. A. Avanzini, et al., “A Refractory Celiac Patient Successfully Treated With Mesenchymal Stem Cell Infusions,” *Mayo Clinic Proceedings* 91, no. 6 (2016): 812–819, <https://doi.org/10.1016/j.mayocp.2016.03.001>.
370. W. H. M. Verbeek, J. M. W. Van De Water, A. Al-Toma, J. J. Oudejans, C. J. J. Mulder, and V. M. H. Coupé, “Incidence of Enteropathy - Associated T-cell Lymphoma: A Nation-Wide Study of a Population-Based Registry in the Netherlands,” *Scandinavian Journal of Gastroenterology* 43, no. 11 (2008): 1322–1328, <https://doi.org/10.1080/00365520802240222>.
371. G. Costantino, A. della Torre, M. A. Lo Presti, R. Caruso, E. Mazzon, and W. Fries, “Treatment of Life-Threatening Type I Refractory Coeliac Disease With long-term Infliximab,” *Digestive and Liver Disease* 40, no. 1 (2008): 74–77, <https://doi.org/10.1016/J.DLD.2006.10.017>.
372. J. K. Grewal, A. Kassardjian, and G. A. Weiss, “Successful Novel Use of Tofacitinib for Type II Refractory Coeliac Disease,” *BMJ Case Reports* 15, no. 4 (2022): e244692, <https://doi.org/10.1136/bcr-2021-244692>.
373. G. Malamut, O. Chandesris, V. Verkarre, et al., “Enteropathy Associated T Cell Lymphoma in Celiac Disease: A Large Retrospective Study,” *Digestive and Liver Disease* 45, no. 5 (2013): 377–384, <https://doi.org/10.1016/J.DLD.2012.12.001>.
374. A. Al-Toma, W. H. M. Verbeek, O. J. Visser, et al., “Disappointing Outcome of Autologous Stem Cell Transplantation for enteropathy-associated T-cell Lymphoma,” *Digestive and Liver Disease* 39, no. 7 (2007): 634–641, <https://doi.org/10.1016/j.dld.2007.03.009>.
375. B. Lebwohl, P. H. R. Green, L. Emilsson, et al., “Cancer Risk in 47,241 Individuals With Celiac Disease: A Nationwide Cohort Study,” *Clinical Gastroenterology and Hepatology* 20, no. 2 (2022): e111–e131, <https://doi.org/10.1016/j.cgh.2021.05.034>.
376. B. Lebwohl, P. H. R. Green, J. Söderling, B. Roelstraete, and J. F. Ludvigsson, “Association Between Celiac Disease and Mortality Risk in a Swedish Population,” *Journal of the American Medical Association* 323, no. 13 (2020): 1277–1285, <https://doi.org/10.1001/jama.2020.1943>.
377. E. Jantunen, A. Boumendil, H. Finel, et al., “Autologous Stem Cell Transplantation for Enteropathy-associated T-cell Lymphoma: A Retrospective Study by the EBMT,” *Blood* 121, no. 13 (2013): 2529–2532, <https://doi.org/10.1182/blood-2012-11-466839>.
378. J. K. C. Chan, A. C. L. Chan, W. Cheuk, et al., “Type II enteropathy-associated T-cell Lymphoma: A Distinct Aggressive Lymphoma With Frequent $\gamma\delta$ T-cell Receptor Expression,” *American Journal of Surgical Pathology* 35, no. 10 (2011): 1557–1569, <https://doi.org/10.1097/PAS.0b013e318222dfdc>.
379. P. Nijeboer, G. Malamut, C. J. Mulder, et al., “Enteropathy-Associated T-cell Lymphoma: Improving Treatment Strategies,” *Digestive Diseases* 33, no. 2 (2015): 231–235, <https://doi.org/10.1159/000369542>.
380. S. A. Rowinski and E. Christensen, “Epidemiologic and Therapeutic Aspects of Refractory Coeliac Disease - A Systematic Review,” *Dan Med J* 63, no. 12 (2016): A5307, <https://pubmed.ncbi.nlm.nih.gov/27910801/>.
381. J. D. G. Linssen, P. J. M. Schafrat, T. R. de Back, et al., “Predisposing Conditions in Patients With Small Intestinal Adenocarcinomas in the Netherlands: A 20-Year Nationwide Cohort Study,” *International Journal of Cancer* 157, no. 2 (2025): 218–231, <https://doi.org/10.1002/IJC.35354>.
382. T. Aparicio, J. Henriques, S. Manfredi, et al., “Small Bowel Adenocarcinoma: Results From a Nationwide Prospective ARCAD-NADEGE Cohort Study of 347 Patients,” *International Journal of Cancer* 147, no. 4 (2020): 967–977, <https://doi.org/10.1002/IJC.32860>.
383. G. Santacroce, A. Vanoli, N. Aronico, et al., “Different Survival Outcomes of Small Bowel Adenocarcinomas and T-Cell Lymphomas Associated With Celiac Disease,” *American Journal of Gastroenterology* 120, no. 12 (2025): 2949–2953, <https://doi.org/10.14309/AJG.0000000000003574>.

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